# (19) World Intellectual Property Organization International Bureau





## (43) International Publication Date 20 December 2001 (20.12.2001)

## **PCT**

# (10) International Publication Number WO 01/96547 A2

(51) International Patent Classification<sup>7</sup>: C12N 9/00

(21) International Application Number: PCT/US01/19444

(22) International Filing Date: 14 June 2001 (14.06.2001)

(25) Filing Language: English

(26) Publication Language: English

(30) Priority Data:

60/212,073	15 June 2000 (15.06.2000)	US
60/213,467	23 June 2000 (23.06.2000)	US
60/215,651	30 June 2000 (30.06.2000)	US
60/216,605	7 July 2000 (07.07.2000)	$\mathbf{U}\mathbf{S}$
60/218,372	13 July 2000 (13.07.2000)	US
60/228,056	25 August 2000 (25.08.2000)	US

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- (81) Designated States (national): AE, AG, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, BZ, CA, CH, CN, CO, CR, CU, CZ, DE, DK, DM, DZ, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, MZ, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZW.
- (84) Designated States (regional): ARIPO patent (GH, GM, KE, LS, MW, MZ, SD, SL, SZ, TZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE, TR), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).

## Published:

 without international search report and to be republished upon receipt of that report

[Continued on next page]

(54) Title: HUMAN KINASES

(57) Abstract: The invention provides human kinases (PKIN) and polynucleotides which identify and encode PKIN. The invention also provides expression vectors, host cells, antibodies, agonists, and antagonists. The invention also provides methods for diagnosing, treating or prevention disorders associated with aberrant expression of PKIN.



VO 01/96



For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.

#### **HUMAN KINASES**

#### TECHNICAL FIELD

This invention relates to nucleic acid and amino acid sequences of human kinases and to the use of these sequences in the diagnosis, treatment, and prevention of cancer, immune disorders, disorders affecting growth and development, cardiovascular diseases, and lipid disorders, and in the assessment of the effects of exogenous compounds on the expression of nucleic acid and amino acid sequences of human kinases.

#### BACKGROUND OF THE INVENTION

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Kinases comprise the largest known enzyme superfamily and vary widely in their target molecules. Kinases catalyze the transfer of high energy phosphate groups from a phosphate donor to a phosphate acceptor. Nucleotides usually serve as the phosphate donor in these reactions, with most kinases utilizing adenosine triphosphate (ATP). The phosphate acceptor can be any of a variety of molecules, including nucleosides, nucleotides, lipids, carbohydrates, and proteins. Proteins are phosphorylated on hydroxyamino acids. Addition of a phosphate group alters the local charge on the acceptor molecule, causing internal conformational changes and potentially influencing intermolecular contacts. Reversible protein phosphorylation is the primary method for regulating protein activity in eukaryotic cells. In general, proteins are activated by phosphorylation in response to extracellular signals such as hormones, neurotransmitters, and growth and differentiation factors. The activated proteins initiate the cell's intracellular response by way of intracellular signaling pathways and second messenger molecules such as cyclic nucleotides, calcium-calmodulin, inositol, and various mitogens, that regulate protein phosphorylation.

Kinases are involved in all aspects of a cell's function, from basic metabolic processes, such as glycolysis, to cell-cycle regulation, differentiation, and communication with the extracellular environment through signal transduction cascades. Inappropriate phosphorylation of proteins in cells has been linked to changes in cell cycle progression and cell differentiation. Changes in the cell cycle have been linked to induction of apoptosis or cancer. Changes in cell differentiation have been linked to diseases and disorders of the reproductive system, immune system, and skeletal muscle.

There are two classes of protein kinases. One class, protein tyrosine kinases (PTKs), phosphorylates tyrosine residues, and the other class, protein serine/threonine kinases (STKs), phosphorylates serine and threonine residues. Some PTKs and STKs possess structural characteristics of both families and have dual specificity for both tyrosine and serine/threonine residues. Almost all kinases contain a conserved 250-300 amino acid catalytic domain containing specific residues and

sequence motifs characteristic of the kinase family. The protein kinase catalytic domain can be further divided into 11 subdomains. N-terminal subdomains I-IV fold into a two-lobed structure which binds and orients the ATP donor molecule, and subdomain V spans the two lobes. C-terminal subdomains VI-XI bind the protein substrate and transfer the gamma phosphate from ATP to the hydroxyl group of a tyrosine, serine, or threonine residue. Each of the 11 subdomains contains specific catalytic residues or amino acid motifs characteristic of that subdomain. For example, subdomain I contains an 8-amino acid glycine-rich ATP binding consensus motif, subdomain II contains a critical lysine residue required for maximal catalytic activity, and subdomains VI through IX comprise the highly conserved catalytic core. PTKs and STKs also contain distinct sequence motifs in subdomains VI and VIII which may confer hydroxyamino acid specificity.

In addition, kinases may also be classified by additional amino acid sequences, generally between 5 and 100 residues, which either flank or occur within the kinase domain. These additional amino acid sequences regulate kinase activity and determine substrate specificity. (Reviewed in Hardie, G. and S. Hanks (1995) The Protein Kinase Facts Book, Vol I, pp. 17-20 Academic Press, San Diego CA.). In particular, two protein kinase signature sequences have been identified in the kinase domain, the first containing an active site lysine residue involved in ATP binding, and the second containing an aspartate residue important for catalytic activity. If a protein analyzed includes the two protein kinase signatures, the probability of that protein being a protein kinase is close to 100% (PROSITE: PDOC00100, November 1995).

## 20 Protein Tyrosine Kinases

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Protein tyrosine kinases (PTKs) may be classified as either transmembrane, receptor PTKs or nontransmembrane, nonreceptor PTK proteins. Transmembrane tyrosine kinases function as receptors for most growth factors. Growth factors bind to the receptor tyrosine kinase (RTK), which causes the receptor to phosphorylate itself (autophosphorylation) and specific intracellular second messenger proteins. Growth factors (GF) that associate with receptor PTKs include epidermal GF, platelet-derived GF, fibroblast GF, hepatocyte GF, insulin and insulin-like GFs, nerve GF, vascular endothelial GF, and macrophage colony stimulating factor.

Nontransmembrane, nonreceptor PTKs lack transmembrane regions and, instead, form signaling complexes with the cytosolic domains of plasma membrane receptors. Receptors that function through non-receptor PTKs include those for cytokines and hormones (growth hormone and prolactin), and antigen-specific receptors on T and B lymphocytes.

Many PTKs were first identified as oncogene products in cancer cells in which PTK activation was no longer subject to normal cellular controls. In fact, about one third of the known oncogenes encode PTKs. Furthermore, cellular transformation (oncogenesis) is often accompanied by increased

tyrosine phosphorylation activity (Charbonneau, H. and N.K. Tonks (1992) Annu. Rev. Cell Biol. 8:463-493). Regulation of PTK activity may therefore be an important strategy in controlling some types of cancer.

#### Protein Serine/Threonine Kinases

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Protein serine/threonine kinases (STKs) are nontransmembrane proteins. A subclass of STKs are known as ERKs (extracellular signal regulated kinases) or MAPs (mitogen-activated protein kinases) and are activated after cell stimulation by a variety of hormones and growth factors. Cell stimulation induces a signaling cascade leading to phosphorylation of MEK (MAP/ERK kinase) which, in turn, activates ERK via serine and threonine phosphorylation. A varied number of proteins represent the downstream effectors for the active ERK and implicate it in the control of cell proliferation and differentiation, as well as regulation of the cytoskeleton. Activation of ERK is normally transient, and cells possess dual specificity phosphatases that are responsible for its down-regulation. Also, numerous studies have shown that elevated ERK activity is associated with some cancers. Other STKs include the second messenger dependent protein kinases such as the cyclic-AMP dependent protein kinases (PKA), calcium-calmodulin (CaM) dependent protein kinases, and the mitogen-activated protein kinases (MAP); the cyclin-dependent protein kinases; checkpoint and cell cycle kinases; Numb-associated kinase (Nak); human Fused (hFu); proliferation-related kinases; 5'-AMP-activated protein kinases; and kinases involved in apoptosis.

The second messenger dependent protein kinases primarily mediate the effects of second messengers such as cyclic AMP (cAMP), cyclic GMP, inositol triphosphate, phosphatidylinositol, 3,4,5-triphosphate, cyclic ADP ribose, arachidonic acid, diacylglycerol and calcium-calmodulin. The PKAs are involved in mediating hormone-induced cellular responses and are activated by cAMP produced within the cell in response to hormone stimulation. cAMP is an intracellular mediator of hormone action in all animal cells that have been studied. Hormone-induced cellular responses include thyroid hormone secretion, cortisol secretion, progesterone secretion, glycogen breakdown, bone resorption, and regulation of heart rate and force of heart muscle contraction. PKA is found in all animal cells and is thought to account for the effects of cAMP in most of these cells. Altered PKA expression is implicated in a variety of disorders and diseases including cancer, thyroid disorders, diabetes, atherosclerosis, and cardiovascular disease (Isselbacher, K.J. et al. (1994) Harrison's Principles of Internal Medicine, McGraw-Hill, New York NY, pp. 416-431, 1887).

The casein kinase I (CKI) gene family is another subfamily of serine/threonine protein kinases. This continuously expanding group of kinases have been implicated in the regulation of numerous cytoplasmic and nuclear processes, including cell metabolism, and DNA replication and repair. CKI enzymes are present in the membranes, nucleus, cytoplasm and cytoskeleton of eukaryotic cells, and on

the mitotic spindles of mammalian cells (Fish, K.J. et al. (1995) J. Biol. Chem. 270:14875-14883).

The CKI family members all have a short amino-terminal domain of 9-76 amino acids, a highly conserved kinase domain of 284 amino acids, and a variable carboxyl-terminal domain that ranges from 24 to over 200 amino acids in length (Cegielska, A. et al. (1998) J. Biol. Chem. 273:1357-1364). The CKI family is comprised of highly related proteins, as seen by the identification of isoforms of casein kinase I from a variety of sources. There are at least five mammalian isoforms,  $\alpha$ ,  $\beta$ ,  $\gamma$ ,  $\delta$ , and  $\epsilon$ . Fish et al., identified CKI-epsilon from a human placenta cDNA library. It is a basic protein of 416 amino acids and is closest to CKI-delta. Through recombinant expression, it was determined to phosphorylate known CKI substrates and was inhibited by the CKI-specific inhibitor CKI-7. The human gene for CKI-epsilon was able to rescue yeast with a slow-growth phenotype caused by deletion of the yeast CKI locus, HRR250 (Fish et al., supra).

The mammalian circadian mutation tau was found to be a semidominant autosomal allele of CKI-epsilon that markedly shortens period length of circadian rhythms in Syrian hamsters. The tau locus is encoded by casein kinase I-epsilon, which is also a homolog of the Drosophila circadian gene double-time. Studies of both the wildtype and tau mutant CKI-epsilon enzyme indicated that the mutant enzyme has a noticeable reduction in the maximum velocity and autophosphorylation state. Further, *in vitro*, CKI-epsilon is able to interact with mammalian PERIOD proteins, while the mutant enzyme is deficient in its ability to phosphorylate PERIOD. Lowrey et al., have proposed that CKI-epsilon plays a major role in delaying the negative feedback signal within the transcription-translation-based autoregulatory loop that composes the core of the circadian mechanism. Therefore the CKI-epsilon enzyme is an ideal target for pharmaceutical compounds influencing circadian rhythms, jet-lag and sleep, in addition to other physiologic and metabolic processes under circadian regulation (Lowrey, P.L. et al. (2000) Science 288:483-491).

Homeodomain-interacting protein kinases (HIPKs) are serine/threonine kinases and novel members of the DYRK kinase subfamily (Hofmann, T.G. et al. (2000) Biochimie 82:1123-1127). HIPKs contain a conserved protein kinase domain separated from a domain that interacts with homeoproteins. HIPKs are nuclear kinases, and HIPK2 is highly expressed in neuronal tissue (Kim, Y.H. et al. (1998) J. Biol. Chem. 273:25875-25879; Wang, Y. et al. (2001) Biochim. Biophys. Acta 1518:168-172). HIPKs act as corepressors for homeodomian transcription factors. This corepressor activity is seen in posttranslational modifications such as ubiquitination and phosphorylation, each of which are important in the regulation of cellular protein function (Kim, Y.H. et al. (1999) Proc. Natl. Acad. Sci. USA 96:12350-12355).

## Calcium-Calmodulin Dependent Protein Kinases

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Calcium-calmodulin dependent (CaM) kinases are involved in regulation of smooth muscle

contraction, glycogen breakdown (phosphorylase kinase), and neurotransmission (CaM kinase I and CaM kinase II). CaM dependent protein kinases are activated by calmodulin, an intracellular calcium receptor, in response to the concentration of free calcium in the cell. Many CaM kinases are also activated by phosphorylation. Some CaM kinases are also activated by autophosphorylation or by other regulatory kinases. CaM kinase I phosphorylates a variety of substrates including the neurotransmitter-related proteins synapsin I and II, the gene transcription regulator, CREB, and the cystic fibrosis conductance regulator protein, CFTR (Haribabu, B. et al. (1995) EMBO J. 14:3679-3686). CaM kinase II also phosphorylates synapsin at different sites and controls the synthesis of catecholamines in the brain through phosphorylation and activation of tyrosine hydroxylase. CaM kinase II controls the synthesis of catecholamines and seratonin, through phosphorylation/activation of tyrosine hydroxylase and tryptophan hydroxylase, respectively (Fujisawa, H. (1990) BioEssays 12:27-29). The mRNA encoding a calmodulin-binding protein kinase-like protein was found to be enriched in mammalian forebrain. This protein is associated with vesicles in both axons and dendrites and accumulates largely postnatally. The amino acid sequence of this protein is similar to CaM-dependent STKs, and the protein binds calmodulin in the presence of calcium (Godbout, M. et al. (1994) J. Neurosci, 14:1-13).

#### Mitogen-Activated Protein Kinases

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The mitogen-activated protein kinases (MAP) which mediate signal transduction from the cell surface to the nucleus via phosphorylation cascades are another STK family that regulates intracellular signaling pathways. Several subgroups have been identified, and each manifests different substrate specificities and responds to distinct extracellular stimuli (Egan, S.E. and R.A. Weinberg (1993) Nature 365:781-783). MAP kinase signaling pathways are present in mammalian cells as well as in yeast. The extracellular stimuli which activate MAP kinase pathways include epidermal growth factor (EGF), ultraviolet light, hyperosmolar medium, heat shock, endotoxic lipopolysaccharide (LPS), and proinflammatory cytokines such as tumor necrosis factor (TNF) and interleukin-1 (IL-1). Altered MAP kinase expression is implicated in a variety of disease conditions including cancer, inflammation, immune disorders, and disorders affecting growth and development.

## Cyclin-Dependent Protein Kinases

The cyclin-dependent protein kinases (CDKs) are STKs that control the progression of cells through the cell cycle. The entry and exit of a cell from mitosis are regulated by the synthesis and destruction of a family of activating proteins called cyclins. Cyclins are small regulatory proteins that bind to and activate CDKs, which then phosphorylate and activate selected proteins involved in the mitotic process. CDKs are unique in that they require multiple inputs to become activated. In addition to cyclin binding, CDK activation requires the phosphorylation of a specific threonine residue and the

dephosphorylation of a specific tyrosine residue on the CDK.

Another family of STKs associated with the cell cycle are the NIMA (never in mitosis)-related kinases (Neks). Both CDKs and Neks are involved in duplication, maturation, and separation of the microtubule organizing center, the centrosome, in animal cells (Fry, A.M. et al. (1998) EMBO J. 17:470-481).

#### Checkpoint and Cell Cycle Kinases

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In the process of cell division, the order and timing of cell cycle transitions are under control of cell cycle checkpoints, which ensure that critical events such as DNA replication and chromosome segregation are carried out with precision. If DNA is damaged, e.g. by radiation, a checkpoint pathway is activated that arrests the cell cycle to provide time for repair. If the damage is extensive, apoptosis is induced. In the absence of such checkpoints, the damaged DNA is inherited by aberrant cells which may cause proliferative disorders such as cancer. Protein kinases play an important role in this process. For example, a specific kinase, checkpoint kinase 1 (Chk1), has been identified in yeast and mammals, and is activated by DNA damage in yeast. Activation of Chk1 leads to the arrest of the cell at the G2/M transition (Sanchez, Y. et al. (1997) Science 277:1497-1501). Specifically, Chk1 phosphorylates the cell division cycle phosphatase CDC25, inhibiting its normal function which is to dephosphorylate and activate the cyclin-dependent kinase Cdc2. Cdc2 activation controls the entry of cells into mitosis (Peng, C.-Y. et al. (1997) Science 277:1501-1505). Thus, activation of Chk1 prevents the damaged cell from entering mitosis. A similar deficiency in a checkpoint kinase, such as Chk1, may also contribute to cancer by failure to arrest cells with damaged DNA at other checkpoints such as G2/M.

### Proliferation-Related Kinases

Proliferation-related kinase is a serum/cytokine inducible STK that is involved in regulation of the cell cycle and cell proliferation in human megakarocytic cells (Li, B. et al. (1996) J. Biol. Chem. 271:19402-19408). Proliferation-related kinase is related to the polo (derived from <u>Drosophila</u> polo gene) family of STKs implicated in cell division. Proliferation-related kinase is downregulated in lung tumor tissue and may be a proto-oncogene whose deregulated expression in normal tissue leads to oncogenic transformation.

#### 5'-AMP-activated protein kinase

A ligand-activated STK protein kinase is 5'-AMP-activated protein kinase (AMPK) (Gao, G. et al. (1996) J. Biol Chem. 271:8675-8681). Mammalian AMPK is a regulator of fatty acid and sterol synthesis through phosphorylation of the enzymes acetyl-CoA carboxylase and hydroxymethylglutaryl-CoA reductase and mediates responses of these pathways to cellular stresses such as heat shock and depletion of glucose and ATP. AMPK is a heterotrimeric complex comprised of

a catalytic alpha subunit and two non-catalytic beta and gamma subunits that are believed to regulate the activity of the alpha subunit. Subunits of AMPK have a much wider distribution in non-lipogenic tissues such as brain, heart, spleen, and lung than expected. This distribution suggests that its role may extend beyond regulation of lipid metabolism alone.

#### 5 Kinases in Apoptosis

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Apoptosis is a highly regulated signaling pathway leading to cell death that plays a crucial role in tissue development and homeostasis. Deregulation of this process is associated with the pathogenesis of a number of diseases including autoimmune disease, neurodegenerative disorders, and cancer. Various STKs play key roles in this process. ZIP kinase is an STK containing a C-terminal leucine zipper domain in addition to its N-terminal protein kinase domain. This C-terminal domain appears to mediate homodimerization and activation of the kinase as well as interactions with transcription factors such as activating transcription factor, ATF4, a member of the cyclic-AMP responsive element binding protein (ATF/CREB) family of transcriptional factors (Sanjo, H. et al. (1998) J. Biol. Chem. 273:29066-29071). DRAK1 and DRAK2 are STKs that share homology with the death-associated protein kinases (DAP kinases), known to function in interferon-γ induced apoptosis (Sanjo et al., supra). Like ZIP kinase, DAP kinases contain a C-terminal protein-protein interaction domain, in the form of ankyrin repeats, in addition to the N-terminal kinase domain. ZIP, DAP, and DRAK kinases induce morphological changes associated with apoptosis when transfected into NIH3T3 cells (Sanjo et al., supra). However, deletion of either the N-terminal kinase catalytic domain or the C-terminal domain of these proteins abolishes apoptosis activity, indicating that in addition to the kinase activity, activity in the C-terminal domain is also necessary for apoptosis, possibly as an interacting domain with a regulator or a specific substrate.

RICK is another STK recently identified as mediating a specific apoptotic pathway involving the death receptor, CD95 (Inohara, N. et al. (1998) J. Biol. Chem. 273:12296-12300). CD95 is a member of the tumor necrosis factor receptor superfamily and plays a critical role in the regulation and homeostasis of the immune system (Nagata, S. (1997) Cell 88:355-365). The CD95 receptor signaling pathway involves recruitment of various intracellular molecules to a receptor complex following ligand binding. This process includes recruitment of the cysteine protease caspase-8 which, in turn, activates a caspase cascade leading to cell death. RICK is composed of an N-terminal kinase catalytic domain and a C-terminal "caspase-recruitment" domain that interacts with caspase-like domains, indicating that RICK plays a role in the recruitment of caspase-8. This interpretation is supported by the fact that the expression of RICK in human 293T cells promotes activation of caspase-8 and potentiates the induction of apoptosis by various proteins involved in the CD95 apoptosis pathway (Inohara et al., supra).

Mitochondrial Protein Kinases

A novel class of eukaryotic kinases, related by sequence to prokaryotic histidine protein kinases, are the mitochondrial protein kinases (MPKs) which seem to have no sequence similarity with other eukaryotic protein kinases. These protein kinases are located exclusively in the mitochondrial matrix space and may have evolved from genes originally present in respiration-dependent bacteria which were endocytosed by primitive eukaryotic cells. MPKs are responsible for phosphorylation and inactivation of the branched-chain alpha-ketoacid dehydrogenase and pyruvate dehydrogenase complexes (Harris, R.A. et al. (1995) Adv. Enzyme Regul. 34:147-162). Five MPKs have been identified. Four members correspond to pyruvate dehydrogenase kinase isozymes, regulating the activity of the pyruyate dehydrogenase complex, which is an important regulatory enzyme at the interface between glycolysis and the citric acid cycle. The fifth member corresponds to a branchedchain alpha-ketoacid dehydrogenase kinase, important in the regulation of the pathway for the disposal of branched-chain amino acids. (Harris, R.A. et al. (1997) Adv. Enzyme Regul. 37:271-293). Both starvation and the diabetic state are known to result in a great increase in the activity of the pyruvate dehydrogenase kinase in the liver, heart and muscle of the rat. This increase contributes in both disease states to the phosphorylation and inactivation of the pyruvate dehydrogenase complex and conservation of pyruvate and lactate for gluconeogenesis (Harris (1995) supra).

### KINASES WITH NON-PROTEIN SUBSTRATES

### 20 Lipid and Inositol kinases

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Lipid kinases phosphorylate hydroxyl residues on lipid head groups. A family of kinases involved in phosphorylation of phosphatidylinositol (PI) has been described, each member phosphorylating a specific carbon on the inositol ring (Leevers, S.J. et al. (1999) Curr. Opin. Cell. Biol. 11:219-225). The phosphorylation of phosphatidylinositol is involved in activation of the protein kinase C signaling pathway. The inositol phospholipids (phosphoinositides) intracellular signaling pathway begins with binding of a signaling molecule to a G-protein linked receptor in the plasma membrane. This leads to the phosphorylation of phosphatidylinositol (PI) residues on the inner side of the plasma membrane by inositol kinases, thus converting PI residues to the biphosphate state (PIP<sub>2</sub>). PIP<sub>2</sub> is then cleaved into inositol triphosphate (IP<sub>3</sub>) and diacylglycerol. These two products act as mediators for separate signaling pathways. Cellular responses that are mediated by these pathways are glycogen breakdown in the liver in response to vasopressin, smooth muscle contraction in response to acetylcholine, and thrombin-induced platelet aggregation.

PI 3-kinase (PI3K), which phosphorylates the D3 position of PI and its derivatives, has a central role in growth factor signal cascades involved in cell growth, differentiation, and metabolism.

PI3K is a heterodimer consisting of an adapter subunit and a catalytic subunit. The adapter subunit acts as a scaffolding protein, interacting with specific tyrosine-phosphorylated proteins, lipid moieties, and other cytosolic factors. When the adapter subunit binds tyrosine phosphorylated targets, such as the insulin responsive substrate (IRS)-1, the catalytic subunit is activated and converts PI (4,5)

5 bisphosphate (PIP<sub>2</sub>) to PI (3,4,5) P<sub>3</sub> (PIP<sub>3</sub>). PIP<sub>3</sub> then activates a number of other proteins, including PKA, protein kinase B (PKB), protein kinase C (PKC), glycogen synthase kinase (GSK)-3, and p70 ribosomal s6 kinase. PI3K also interacts directly with the cytoskeletal organizing proteins, Rac, rho, and cdc42 (Shepherd, P.R. et al. (1998) Biochem. J. 333:471-490). Animal models for diabetes, such as *obese* and *fat* mice, have altered PI3K adapter subunit levels. Specific mutations in the adapter subunit have also been found in an insulin-resistant Danish population, suggesting a role for PI3K in type-2 diabetes (Shepard, supra).

An example of lipid kinase phosphorylation activity is the phosphorylation of D-erythro-sphingosine to the sphingolipid metabolite, sphingosine-1-phosphate (SPP). SPP has emerged as a novel lipid second-messenger with both extracellular and intracellular actions (Kohama, T. et al. (1998) J. Biol. Chem. 273:23722-23728). Extracellularly, SPP is a ligand for the G-protein coupled receptor EDG-1 (endothelial-derived, G-protein coupled receptor). Intracellularly, SPP regulates cell growth, survival, motility, and cytoskeletal changes. SPP levels are regulated by sphingosine kinases that specifically phosphorylate D-erythro-sphingosine to SPP. The importance of sphingosine kinase in cell signaling is indicated by the fact that various stimuli, including platelet-derived growth factor (PDGF), nerve growth factor, and activation of protein kinase C, increase cellular levels of SPP by activation of sphingosine kinase, and the fact that competitive inhibitors of the enzyme selectively inhibit cell proliferation induced by PDGF (Kohama et al., supra).

#### Purine Nucleotide Kinases

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The purine nucleotide kinases, adenylate kinase (ATP:AMP phosphotransferase, or AdK) and guanylate kinase (ATP:GMP phosphotransferase, or GuK) play a key role in nucleotide metabolism and are crucial to the synthesis and regulation of cellular levels of ATP and GTP, respectively. These two molecules are precursors in DNA and RNA synthesis in growing cells and provide the primary source of biochemical energy in cells (ATP), and signal transduction pathways (GTP). Inhibition of various steps in the synthesis of these two molecules has been the basis of many antiproliferative drugs for cancer and antiviral therapy (Pillwein, K. et al. (1990) Cancer Res. 50:1576-1579).

AdK is found in almost all cell types and is especially abundant in cells having high rates of ATP synthesis and utilization such as skeletal muscle. In these cells AdK is physically associated with mitochondria and myofibrils, the subcellular structures that are involved in energy production and utilization, respectively. Recent studies have demonstrated a major function for AdK in transferring

high energy phosphoryls from metabolic processes generating ATP to cellular components consuming ATP (Zeleznikar, R.J. et al. (1995) J. Biol. Chem. 270:7311-7319). Thus AdK may have a pivotal role in maintaining energy production in cells, particularly those having a high rate of growth or metabolism such as cancer cells, and may provide a target for suppression of its activity to treat certain cancers.

Alternatively, reduced AdK activity may be a source of various metabolic, muscle-energy disorders that can result in cardiac or respiratory failure and may be treatable by increasing AdK activity.

GuK, in addition to providing a key step in the synthesis of GTP for RNA and DNA synthesis, also fulfills an essential function in signal transduction pathways of cells through the regulation of GDP and GTP. Specifically, GTP binding to membrane associated G proteins mediates the activation of cell receptors, subsequent intracellular activation of adenyl cyclase, and production of the second messenger, cyclic AMP. GDP binding to G proteins inhibits these processes. GDP and GTP levels also control the activity of certain oncogenic proteins such as p21<sup>ras</sup> known to be involved in control of cell proliferation and oncogenesis (Bos, J.L. (1989) Cancer Res. 49:4682-4689). High ratios of GTP:GDP caused by suppression of GuK cause activation of p21<sup>ras</sup> and promote oncogenesis. Increasing GuK activity to increase levels of GDP and reduce the GTP:GDP ratio may provide a therapeutic strategy to reverse oncogenesis.

GuK is an important enzyme in the phosphorylation and activation of certain antiviral drugs useful in the treatment of herpes virus infections. These drugs include the guanine homologs acyclovir and buciclovir (Miller, W.H. and R.L. Miller (1980) J. Biol. Chem. 255:7204-7207; Stenberg, K. et al. (1986) J. Biol. Chem. 261:2134-2139). Increasing GuK activity in infected cells may provide a therapeutic strategy for augmenting the effectiveness of these drugs and possibly for reducing the necessary dosages of the drugs.

#### Pyrimidine Kinases

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The pyrimidine kinases are deoxycytidine kinase and thymidine kinase 1 and 2. Deoxycytidine kinase is located in the nucleus, and thymidine kinase 1 and 2 are found in the cytosol (Johansson, M. et al. (1997) Proc. Natl. Acad. Sci. USA 94:11941-11945). Phosphorylation of deoxyribonucleosides by pyrimidine kinases provides an alternative pathway for <u>de novo</u> synthesis of DNA precursors. The role of pyrimidine kinases, like purine kinases, in phosphorylation is critical to the activation of several chemotherapeutically important nucleoside analogues (Arner E.S. and S. Eriksson (1995) Pharmacol. Ther. 67:155-186).

The discovery of new human kinases and the polynucleotides encoding them satisfies a need in the art by providing new compositions which are useful in the diagnosis, prevention, and treatment of cancer, immune disorders, disorders affecting growth and development, cardiovascular diseases, and lipid disorders, and in the assessment of the effects of exogenous compounds on the expression of

nucleic acid and amino acid sequences of human kinases.

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#### SUMMARY OF THE INVENTION

The invention features purified polypeptides, human kinases, referred to collectively as "PKIN" and individually as "PKIN-1," "PKIN-2," "PKIN-3," "PKIN-4," "PKIN-5," "PKIN-6," "PKIN-7," "PKIN-8," "PKIN-9," "PKIN-10," "PKIN-11," "PKIN-12," "PKIN-13," "PKIN-14," "PKIN-15," "PKIN-16," "PKIN-17," "PKIN-18," "PKIN-19," "PKIN-20," "PKIN-21," "PKIN-22," "PKIN-23," "PKIN-25," and "PKIN-26." In one aspect, the invention provides an isolated polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-26. In one alternative, the invention provides an isolated polypeptide comprising the amino acid sequence of SEQ ID NO:1-26.

The invention further provides an isolated polynucleotide encoding a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-26. In one alternative, the polynucleotide encodes a polypeptide selected from the group consisting of SEQ ID NO:1-26. In another alternative, the polynucleotide is selected from the group consisting of SEQ ID NO:27-52.

Additionally, the invention provides a recombinant polynucleotide comprising a promoter sequence operably linked to a polynucleotide encoding a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-26. In one alternative, the invention provides a cell transformed with the recombinant polynucleotide. In another alternative, the invention provides a

transgenic organism comprising the recombinant polynucleotide.

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The invention also provides a method for producing a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-26. The method comprises a) culturing a cell under conditions suitable for expression of the polypeptide, wherein said cell is transformed with a recombinant polynucleotide comprising a promoter sequence operably linked to a polynucleotide encoding the polypeptide, and b) recovering the polypeptide so expressed.

Additionally, the invention provides an isolated antibody which specifically binds to a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-26.

The invention further provides an isolated polynucleotide selected from the group consisting of a) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:27-52, b) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:27-52, c) a polynucleotide complementary to the polynucleotide of a), d) a polynucleotide complementary to the polynucleotide of b), and e) an RNA equivalent of a)-d). In one alternative, the polynucleotide comprises at least 60 contiguous nucleotides.

Additionally, the invention provides a method for detecting a target polynucleotide in a sample, said target polynucleotide having a sequence of a polynucleotide selected from the group consisting of a) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:27-52, b) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:27-52, c) a polynucleotide complementary to the polynucleotide of a), d) a polynucleotide complementary to the polynucleotide of b), and e) an RNA equivalent of a)-d). The method comprises a) hybridizing the sample with a probe comprising at least 20 contiguous nucleotides comprising a sequence complementary to said target polynucleotide in the sample, and which probe specifically hybridizes to

said target polynucleotide, under conditions whereby a hybridization complex is formed between said probe and said target polynucleotide or fragments thereof, and b) detecting the presence or absence of said hybridization complex, and optionally, if present, the amount thereof. In one alternative, the probe comprises at least 60 contiguous nucleotides.

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The invention further provides a method for detecting a target polynucleotide in a sample, said target polynucleotide having a sequence of a polynucleotide selected from the group consisting of a) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:27-52, b) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:27-52, c) a polynucleotide complementary to the polynucleotide of a), d) a polynucleotide complementary to the polynucleotide of b), and e) an RNA equivalent of a)-d). The method comprises a) amplifying said target polynucleotide or fragment thereof using polymerase chain reaction amplification, and b) detecting the presence or absence of said amplified target polynucleotide or fragment thereof, and, optionally, if present, the amount thereof.

The invention further provides a composition comprising an effective amount of a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, and a pharmaceutically acceptable excipient. In one embodiment, the composition comprises an amino acid sequence selected from the group consisting of SEQ ID NO:1-26. The invention additionally provides a method of treating a disease or condition associated with decreased expression of functional PKIN, comprising administering to a patient in need of such treatment the composition.

The invention also provides a method for screening a compound for effectiveness as an agonist of a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-26. The method comprises a) exposing a sample comprising the polypeptide to a compound, and b) detecting agonist activity in the sample. In one alternative, the invention provides a

composition comprising an agonist compound identified by the method and a pharmaceutically acceptable excipient. In another alternative, the invention provides a method of treating a disease or condition associated with decreased expression of functional PKIN, comprising administering to a patient in need of such treatment the composition.

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Additionally, the invention provides a method for screening a compound for effectiveness as an antagonist of a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-26. The method comprises a) exposing a sample comprising the polypeptide to a compound, and b) detecting antagonist activity in the sample. In one alternative, the invention provides a composition comprising an antagonist compound identified by the method and a pharmaceutically acceptable excipient. In another alternative, the invention provides a method of treating a disease or condition associated with overexpression of functional PKIN, comprising administering to a patient in need of such treatment the composition.

The invention further provides a method of screening for a compound that specifically binds to a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-26. The method comprises a) combining the polypeptide with at least one test compound under suitable conditions, and b) detecting binding of the polypeptide to the test compound, thereby identifying a compound that specifically binds to the polypeptide.

The invention further provides a method of screening for a compound that modulates the activity of a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID

NO:1-26. The method comprises a) combining the polypeptide with at least one test compound under conditions permissive for the activity of the polypeptide, b) assessing the activity of the polypeptide in the presence of the test compound, and c) comparing the activity of the polypeptide in the presence of the test compound with the activity of the polypeptide in the absence of the test compound, wherein a change in the activity of the polypeptide in the presence of the test compound is indicative of a compound that modulates the activity of the polypeptide.

The invention further provides a method for screening a compound for effectiveness in altering expression of a target polynucleotide, wherein said target polynucleotide comprises a sequence selected from the group consisting of SEQ ID NO:27-52, the method comprising a) exposing a sample comprising the target polynucleotide to a compound, and b) detecting altered expression of the target polynucleotide.

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The invention further provides a method for assessing toxicity of a test compound, said method comprising a) treating a biological sample containing nucleic acids with the test compound; b) hybridizing the nucleic acids of the treated biological sample with a probe comprising at least 20 contiguous nucleotides of a polynucleotide selected from the group consisting of i) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:27-52, ii) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:27-52, iii) a polynucleotide having a sequence complementary to i), iv) a polynucleotide complementary to the polynucleotide of ii), and v) an RNA equivalent of i)-iv). Hybridization occurs under conditions whereby a specific hybridization complex is formed between said probe and a target polynucleotide in the biological sample, said target polynucleotide selected from the group consisting of i) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:27-52, ii) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:27-52, iii) a polynucleotide complementary to the polynucleotide of i), iv) a polynucleotide complementary to the polynucleotide of ii), and v) an RNA equivalent of i)-iv). Alternatively, the target polynucleotide comprises a fragment of a polynucleotide sequence selected from the group consisting of i)-v) above; c) quantifying the amount of hybridization complex; and d) comparing the amount of hybridization complex in the treated biological sample with the amount of hybridization complex in an untreated biological sample, wherein a difference in the amount of hybridization complex in the treated biological sample is indicative of toxicity of the test compound.

#### BRIEF DESCRIPTION OF THE TABLES

Table 1 summarizes the nomenclature for the full length polynucleotide and polypeptide

sequences of the present invention.

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Table 2 shows the GenBank identification number and annotation of the nearest GenBank homolog for polypeptides of the invention. The probability score for the match between each polypeptide and its GenBank homolog is also shown.

Table 3 shows structural features of polypeptide sequences of the invention, including predicted motifs and domains, along with the methods, algorithms, and searchable databases used for analysis of the polypeptides.

Table 4 lists the cDNA and/or genomic DNA fragments which were used to assemble polynucleotide sequences of the invention, along with selected fragments of the polynucleotide sequences.

Table 5 shows the representative cDNA library for polynucleotides of the invention.

Table 6 provides an appendix which describes the tissues and vectors used for construction of the cDNA libraries shown in Table 5.

Table 7 shows the tools, programs, and algorithms used to analyze the polynucleotides and polypeptides of the invention, along with applicable descriptions, references, and threshold parameters.

#### **DESCRIPTION OF THE INVENTION**

Before the present proteins, nucleotide sequences, and methods are described, it is understood that this invention is not limited to the particular machines, materials and methods described, as these may vary. It is also to be understood that the terminology used herein is for the purpose of describing particular embodiments only, and is not intended to limit the scope of the present invention which will be limited only by the appended claims.

It must be noted that as used herein and in the appended claims, the singular forms "a," "an," and "the" include plural reference unless the context clearly dictates otherwise. Thus, for example, a reference to "a host cell" includes a plurality of such host cells, and a reference to "an antibody" is a reference to one or more antibodies and equivalents thereof known to those skilled in the art, and so forth.

Unless defined otherwise, all technical and scientific terms used herein have the same meanings as commonly understood by one of ordinary skill in the art to which this invention belongs. Although any machines, materials, and methods similar or equivalent to those described herein can be used to practice or test the present invention, the preferred machines, materials and methods are now described. All publications mentioned herein are cited for the purpose of describing and disclosing the cell lines, protocols, reagents and vectors which are reported in the publications and which might be used in connection with the invention. Nothing herein is to be construed as an admission that the invention is

not entitled to antedate such disclosure by virtue of prior invention.

#### **DEFINITIONS**

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"PKIN" refers to the amino acid sequences of substantially purified PKIN obtained from any species, particularly a mammalian species, including bovine, ovine, porcine, murine, equine, and human, and from any source, whether natural, synthetic, semi-synthetic, or recombinant.

The term "agonist" refers to a molecule which intensifies or mimics the biological activity of PKIN. Agonists may include proteins, nucleic acids, carbohydrates, small molecules, or any other compound or composition which modulates the activity of PKIN either by directly interacting with PKIN or by acting on components of the biological pathway in which PKIN participates.

An "allelic variant" is an alternative form of the gene encoding PKIN. Allelic variants may result from at least one mutation in the nucleic acid sequence and may result in altered mRNAs or in polypeptides whose structure or function may or may not be altered. A gene may have none, one, or many allelic variants of its naturally occurring form. Common mutational changes which give rise to allelic variants are generally ascribed to natural deletions, additions, or substitutions of nucleotides. Each of these types of changes may occur alone, or in combination with the others, one or more times in a given sequence.

"Altered" nucleic acid sequences encoding PKIN include those sequences with deletions, insertions, or substitutions of different nucleotides, resulting in a polypeptide the same as PKIN or a polypeptide with at least one functional characteristic of PKIN. Included within this definition are polymorphisms which may or may not be readily detectable using a particular oligonucleotide probe of the polynucleotide encoding PKIN, and improper or unexpected hybridization to allelic variants, with a locus other than the normal chromosomal locus for the polynucleotide sequence encoding PKIN. The encoded protein may also be "altered," and may contain deletions, insertions, or substitutions of amino acid residues which produce a silent change and result in a functionally equivalent PKIN. Deliberate amino acid substitutions may be made on the basis of similarity in polarity, charge, solubility, hydrophobicity, hydrophilicity, and/or the amphipathic nature of the residues, as long as the biological or immunological activity of PKIN is retained. For example, negatively charged amino acids may include aspartic acid and glutamic acid, and positively charged amino acids may include lysine and arginine. Amino acids with uncharged polar side chains having similar hydrophilicity values may include: asparagine and glutamine; and serine and threonine. Amino acids with uncharged side chains having similar hydrophilicity values may include: leucine, isoleucine, and valine; glycine and alanine; and phenylalanine and tyrosine.

The terms "amino acid" and "amino acid sequence" refer to an oligopeptide, peptide, polypeptide, or protein sequence, or a fragment of any of these, and to naturally occurring or synthetic

molecules. Where "amino acid sequence" is recited to refer to a sequence of a naturally occurring protein molecule, "amino acid sequence" and like terms are not meant to limit the amino acid sequence to the complete native amino acid sequence associated with the recited protein molecule.

"Amplification" relates to the production of additional copies of a nucleic acid sequence.

Amplification is generally carried out using polymerase chain reaction (PCR) technologies well known in the art.

The term "antagonist" refers to a molecule which inhibits or attenuates the biological activity of PKIN. Antagonists may include proteins such as antibodies, nucleic acids, carbohydrates, small molecules, or any other compound or composition which modulates the activity of PKIN either by directly interacting with PKIN or by acting on components of the biological pathway in which PKIN participates.

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The term "antibody" refers to intact immunoglobulin molecules as well as to fragments thereof, such as Fab, F(ab')<sub>2</sub>, and Fv fragments, which are capable of binding an epitopic determinant. Antibodies that bind PKIN polypeptides can be prepared using intact polypeptides or using fragments containing small peptides of interest as the immunizing antigen. The polypeptide or oligopeptide used to immunize an animal (e.g., a mouse, a rat, or a rabbit) can be derived from the translation of RNA, or synthesized chemically, and can be conjugated to a carrier protein if desired. Commonly used carriers that are chemically coupled to peptides include bovine serum albumin, thyroglobulin, and keyhole limpet hemocyanin (KLH). The coupled peptide is then used to immunize the animal.

The term "antigenic determinant" refers to that region of a molecule (i.e., an epitope) that makes contact with a particular antibody. When a protein or a fragment of a protein is used to immunize a host animal, numerous regions of the protein may induce the production of antibodies which bind specifically to antigenic determinants (particular regions or three-dimensional structures on the protein). An antigenic determinant may compete with the intact antigen (i.e., the immunogen used to elicit the immune response) for binding to an antibody.

The term "antisense" refers to any composition capable of base-pairing with the "sense" (coding) strand of a specific nucleic acid sequence. Antisense compositions may include DNA; RNA; peptide nucleic acid (PNA); oligonucleotides having modified backbone linkages such as phosphorothioates, methylphosphonates, or benzylphosphonates; oligonucleotides having modified sugar groups such as 2'-methoxyethyl sugars or 2'-methoxyethoxy sugars; or oligonucleotides having modified bases such as 5-methyl cytosine, 2'-deoxyuracil, or 7-deaza-2'-deoxyguanosine. Antisense molecules may be produced by any method including chemical synthesis or transcription. Once introduced into a cell, the complementary antisense molecule base-pairs with a naturally occurring nucleic acid sequence produced by the cell to form duplexes which block either transcription or

translation. The designation "negative" or "minus" can refer to the antisense strand, and the designation "positive" or "plus" can refer to the sense strand of a reference DNA molecule.

The term "biologically active" refers to a protein having structural, regulatory, or biochemical functions of a naturally occurring molecule. Likewise, "immunologically active" or "immunogenic" refers to the capability of the natural, recombinant, or synthetic PKIN, or of any oligopeptide thereof, to induce a specific immune response in appropriate animals or cells and to bind with specific antibodies.

"Complementary" describes the relationship between two single-stranded nucleic acid sequences that anneal by base-pairing. For example, 5'-AGT-3' pairs with its complement, 3'-TCA-5'.

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A "composition comprising a given polynucleotide sequence" and a "composition comprising a given amino acid sequence" refer broadly to any composition containing the given polynucleotide or amino acid sequence. The composition may comprise a dry formulation or an aqueous solution. Compositions comprising polynucleotide sequences encoding PKIN or fragments of PKIN may be employed as hybridization probes. The probes may be stored in freeze-dried form and may be associated with a stabilizing agent such as a carbohydrate. In hybridizations, the probe may be deployed in an aqueous solution containing salts (e.g., NaCl), detergents (e.g., sodium dodecyl sulfate; SDS), and other components (e.g., Denhardt's solution, dry milk, salmon sperm DNA, etc.).

"Consensus sequence" refers to a nucleic acid sequence which has been subjected to repeated DNA sequence analysis to resolve uncalled bases, extended using the XL-PCR kit (Applied Biosystems, Foster City CA) in the 5' and/or the 3' direction, and resequenced, or which has been assembled from one or more overlapping cDNA, EST, or genomic DNA fragments using a computer program for fragment assembly, such as the GELVIEW fragment assembly system (GCG, Madison WI) or Phrap (University of Washington, Seattle WA). Some sequences have been both extended and assembled to produce the consensus sequence.

"Conservative amino acid substitutions" are those substitutions that are predicted to least interfere with the properties of the original protein, i.e., the structure and especially the function of the protein is conserved and not significantly changed by such substitutions. The table below shows amino acids which may be substituted for an original amino acid in a protein and which are regarded as conservative amino acid substitutions.

30	Original Residue	Conservative Substitution
	Ala	Gly, Ser
	Arg	His, Lys
	Asn	Asp, Gln, His
	Asp	Asn, Glu
35	Cys	Ala, Ser
	Gln	Asn, Glu, His

	Glu	Asp, Gln, His
	Gly	Ala
	His	Asn, Arg, Gln, Glu
	Ile	Leu, Val
5	Leu	Ile, Val
	Lys	Arg, Gln, Glu
	Met	Leu, Ile
	Phe	His, Met, Leu, Trp, Tyr
	Ser	Cys, Thr
10	Thr	Ser, Val
	Trp	Phe, Tyr
	Tyr	His, Phe, Trp
	Val	Ile, Leu, Thr

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Conservative amino acid substitutions generally maintain (a) the structure of the polypeptide backbone in the area of the substitution, for example, as a beta sheet or alpha helical conformation, (b) the charge or hydrophobicity of the molecule at the site of the substitution, and/or (c) the bulk of the side chain.

A "deletion" refers to a change in the amino acid or nucleotide sequence that results in the absence of one or more amino acid residues or nucleotides.

The term "derivative" refers to a chemically modified polynucleotide or polypeptide. Chemical modifications of a polynucleotide can include, for example, replacement of hydrogen by an alkyl, acyl, hydroxyl, or amino group. A derivative polynucleotide encodes a polypeptide which retains at least one biological or immunological function of the natural molecule. A derivative polypeptide is one modified by glycosylation, pegylation, or any similar process that retains at least one biological or immunological function of the polypeptide from which it was derived.

A "detectable label" refers to a reporter molecule or enzyme that is capable of generating a measurable signal and is covalently or noncovalently joined to a polynucleotide or polypeptide.

"Differential expression" refers to increased or upregulated; or decreased, downregulated, or absent gene or protein expression, determined by comparing at least two different samples. Such comparisons may be carried out between, for example, a treated and an untreated sample, or a diseased and a normal sample.

A "fragment" is a unique portion of PKIN or the polynucleotide encoding PKIN which is identical in sequence to but shorter in length than the parent sequence. A fragment may comprise up to the entire length of the defined sequence, minus one nucleotide/amino acid residue. For example, a fragment may comprise from 5 to 1000 contiguous nucleotides or amino acid residues. A fragment used as a probe, primer, antigen, therapeutic molecule, or for other purposes, may be at least 5, 10, 15, 16, 20, 25, 30, 40, 50, 60, 75, 100, 150, 250 or at least 500 contiguous nucleotides or amino acid residues in length. Fragments may be preferentially selected from certain regions of a molecule. For

example, a polypeptide fragment may comprise a certain length of contiguous amino acids selected from the first 250 or 500 amino acids (or first 25% or 50%) of a polypeptide as shown in a certain defined sequence. Clearly these lengths are exemplary, and any length that is supported by the specification, including the Sequence Listing, tables, and figures, may be encompassed by the present embodiments.

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A fragment of SEQ ID NO:27-52 comprises a region of unique polynucleotide sequence that specifically identifies SEQ ID NO:27-52, for example, as distinct from any other sequence in the genome from which the fragment was obtained. A fragment of SEQ ID NO:27-52 is useful, for example, in hybridization and amplification technologies and in analogous methods that distinguish SEQ ID NO:27-52 from related polynucleotide sequences. The precise length of a fragment of SEQ ID NO:27-52 and the region of SEQ ID NO:27-52 to which the fragment corresponds are routinely determinable by one of ordinary skill in the art based on the intended purpose for the fragment.

A fragment of SEQ ID NO:1-26 is encoded by a fragment of SEQ ID NO:27-52. A fragment of SEQ ID NO:1-26 comprises a region of unique amino acid sequence that specifically identifies SEQ ID NO:1-26. For example, a fragment of SEQ ID NO:1-26 is useful as an immunogenic peptide for the development of antibodies that specifically recognize SEQ ID NO:1-26. The precise length of a fragment of SEQ ID NO:1-26 and the region of SEQ ID NO:1-26 to which the fragment corresponds are routinely determinable by one of ordinary skill in the art based on the intended purpose for the fragment.

A "full length" polynucleotide sequence is one containing at least a translation initiation codon (e.g., methionine) followed by an open reading frame and a translation termination codon. A "full length" polynucleotide sequence encodes a "full length" polypeptide sequence.

"Homology" refers to sequence similarity or, interchangeably, sequence identity, between two or more polynucleotide sequences or two or more polypeptide sequences.

The terms "percent identity" and "% identity," as applied to polynucleotide sequences, refer to the percentage of residue matches between at least two polynucleotide sequences aligned using a standardized algorithm. Such an algorithm may insert, in a standardized and reproducible way, gaps in the sequences being compared in order to optimize alignment between two sequences, and therefore achieve a more meaningful comparison of the two sequences.

Percent identity between polynucleotide sequences may be determined using the default parameters of the CLUSTAL V algorithm as incorporated into the MEGALIGN version 3.12e sequence alignment program. This program is part of the LASERGENE software package, a suite of molecular biological analysis programs (DNASTAR, Madison WI). CLUSTAL V is described in Higgins, D.G. and P.M. Sharp (1989) CABIOS 5:151-153 and in Higgins, D.G. et al. (1992) CABIOS 8:189-191.

For pairwise alignments of polynucleotide sequences, the default parameters are set as follows: Ktuple=2, gap penalty=5, window=4, and "diagonals saved"=4. The "weighted" residue weight table is selected as the default. Percent identity is reported by CLUSTAL V as the "percent similarity" between aligned polynucleotide sequences.

Alternatively, a suite of commonly used and freely available sequence comparison algorithms is provided by the National Center for Biotechnology Information (NCBI) Basic Local Alignment Search Tool (BLAST) (Altschul, S.F. et al. (1990) J. Mol. Biol. 215:403-410), which is available from several sources, including the NCBI, Bethesda, MD, and on the Internet at http://www.ncbi.nlm.nih.gov/BLAST/. The BLAST software suite includes various sequence analysis programs including "blastn," that is used to align a known polynucleotide sequence with other polynucleotide sequences from a variety of databases. Also available is a tool called "BLAST 2 Sequences" that is used for direct pairwise comparison of two nucleotide sequences. "BLAST 2 Sequences" can be accessed and used interactively at http://www.ncbi.nlm.nih.gov/gorf/bl2.html. The "BLAST 2 Sequences" tool can be used for both blastn and blastp (discussed below). BLAST programs are commonly used with gap and other parameters set to default settings. For example, to compare two nucleotide sequences, one may use blastn with the "BLAST 2 Sequences" tool Version

2.0.12 (April-21-2000) set at default parameters. Such default parameters may be, for example:

Matrix: BLOSUM62

Reward for match: 1

20 Penalty for mismatch: -2

Open Gap: 5 and Extension Gap: 2 penalties

Gap x drop-off: 50

Expect: 10

Word Size: 11

25 Filter: on

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Percent identity may be measured over the length of an entire defined sequence, for example, as defined by a particular SEQ ID number, or may be measured over a shorter length, for example, over the length of a fragment taken from a larger, defined sequence, for instance, a fragment of at least 20, at least 30, at least 40, at least 50, at least 70, at least 100, or at least 200 contiguous nucleotides. Such lengths are exemplary only, and it is understood that any fragment length supported by the sequences shown herein, in the tables, figures, or Sequence Listing, may be used to describe a length over which percentage identity may be measured.

Nucleic acid sequences that do not show a high degree of identity may nevertheless encode similar amino acid sequences due to the degeneracy of the genetic code. It is understood that changes in

a nucleic acid sequence can be made using this degeneracy to produce multiple nucleic acid sequences that all encode substantially the same protein.

The phrases "percent identity" and "% identity," as applied to polypeptide sequences, refer to the percentage of residue matches between at least two polypeptide sequences aligned using a standardized algorithm. Methods of polypeptide sequence alignment are well-known. Some alignment methods take into account conservative amino acid substitutions. Such conservative substitutions, explained in more detail above, generally preserve the charge and hydrophobicity at the site of substitution, thus preserving the structure (and therefore function) of the polypeptide.

Percent identity between polypeptide sequences may be determined using the default parameters of the CLUSTAL V algorithm as incorporated into the MEGALIGN version 3.12e sequence alignment program (described and referenced above). For pairwise alignments of polypeptide sequences using CLUSTAL V, the default parameters are set as follows: Ktuple=1, gap penalty=3, window=5, and "diagonals saved"=5. The PAM250 matrix is selected as the default residue weight table. As with polynucleotide alignments, the percent identity is reported by CLUSTAL V as the "percent similarity" between aligned polypeptide sequence pairs.

Alternatively the NCBI BLAST software suite may be used. For example, for a pairwise comparison of two polypeptide sequences, one may use the "BLAST 2 Sequences" tool Version 2.0.12 (April-21-2000) with blastp set at default parameters. Such default parameters may be, for example:

Matrix: BLOSUM62

Open Gap: 11 and Extension Gap: 1 penalties

Gap x drop-off: 50

Expect: 10
Word Size: 3

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Filter: on

Percent identity may be measured over the length of an entire defined polypeptide sequence, for example, as defined by a particular SEQ ID number, or may be measured over a shorter length, for example, over the length of a fragment taken from a larger, defined polypeptide sequence, for instance, a fragment of at least 15, at least 20, at least 30, at least 40, at least 50, at least 70 or at least 150 contiguous residues. Such lengths are exemplary only, and it is understood that any fragment length supported by the sequences shown herein, in the tables, figures or Sequence Listing, may be used to describe a length over which percentage identity may be measured.

"Human artificial chromosomes" (HACs) are linear microchromosomes which may contain DNA sequences of about 6 kb to 10 Mb in size and which contain all of the elements required for chromosome replication, segregation and maintenance.

The term "humanized antibody" refers to an antibody molecule in which the amino acid sequence in the non-antigen binding regions has been altered so that the antibody more closely resembles a human antibody, and still retains its original binding ability.

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"Hybridization" refers to the process by which a polynucleotide strand anneals with a complementary strand through base pairing under defined hybridization conditions. Specific hybridization is an indication that two nucleic acid sequences share a high degree of complementarity. Specific hybridization complexes form under permissive annealing conditions and remain hybridized after the "washing" step(s). The washing step(s) is particularly important in determining the stringency of the hybridization process, with more stringent conditions allowing less non-specific binding, i.e., binding between pairs of nucleic acid strands that are not perfectly matched. Permissive conditions for annealing of nucleic acid sequences are routinely determinable by one of ordinary skill in the art and may be consistent among hybridization experiments, whereas wash conditions may be varied among experiments to achieve the desired stringency, and therefore hybridization specificity. Permissive annealing conditions occur, for example, at 68°C in the presence of about 6 x SSC, about 1% (w/v) SDS, and about 100 μg/ml sheared, denatured salmon sperm DNA.

Generally, stringency of hybridization is expressed, in part, with reference to the temperature under which the wash step is carried out. Such wash temperatures are typically selected to be about 5°C to 20°C lower than the thermal melting point  $(T_m)$  for the specific sequence at a defined ionic strength and pH. The  $T_m$  is the temperature (under defined ionic strength and pH) at which 50% of the target sequence hybridizes to a perfectly matched probe. An equation for calculating  $T_m$  and conditions for nucleic acid hybridization are well known and can be found in Sambrook, J. et al. (1989) Molecular Cloning: A Laboratory Manual,  $2^{nd}$  ed., vol. 1-3, Cold Spring Harbor Press, Plainview NY; specifically see volume 2, chapter 9.

High stringency conditions for hybridization between polynucleotides of the present invention include wash conditions of 68°C in the presence of about 0.2 x SSC and about 0.1% SDS, for 1 hour. Alternatively, temperatures of about  $65^{\circ}$ C,  $60^{\circ}$ C,  $55^{\circ}$ C, or  $42^{\circ}$ C may be used. SSC concentration may be varied from about 0.1 to 2 x SSC, with SDS being present at about 0.1%. Typically, blocking reagents are used to block non-specific hybridization. Such blocking reagents include, for instance, sheared and denatured salmon sperm DNA at about  $100\text{-}200 \,\mu\text{g/ml}$ . Organic solvent, such as formamide at a concentration of about  $35\text{-}50\% \,\text{v/v}$ , may also be used under particular circumstances, such as for RNA:DNA hybridizations. Useful variations on these wash conditions will be readily apparent to those of ordinary skill in the art. Hybridization, particularly under high stringency conditions, may be suggestive of evolutionary similarity between the nucleotides. Such similarity is strongly indicative of a similar role for the nucleotides and their encoded polypeptides.

The term "hybridization complex" refers to a complex formed between two nucleic acid sequences by virtue of the formation of hydrogen bonds between complementary bases. A hybridization complex may be formed in solution (e.g.,  $C_0$ t or  $R_0$ t analysis) or formed between one nucleic acid sequence present in solution and another nucleic acid sequence immobilized on a solid support (e.g., paper, membranes, filters, chips, pins or glass slides, or any other appropriate substrate to which cells or their nucleic acids have been fixed).

The words "insertion" and "addition" refer to changes in an amino acid or nucleotide sequence resulting in the addition of one or more amino acid residues or nucleotides, respectively.

"Immune response" can refer to conditions associated with inflammation, trauma, immune disorders, or infectious or genetic disease, etc. These conditions can be characterized by expression of various factors, e.g., cytokines, chemokines, and other signaling molecules, which may affect cellular and systemic defense systems.

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An "immunogenic fragment" is a polypeptide or oligopeptide fragment of PKIN which is capable of eliciting an immune response when introduced into a living organism, for example, a mammal. The term "immunogenic fragment" also includes any polypeptide or oligopeptide fragment of PKIN which is useful in any of the antibody production methods disclosed herein or known in the art.

The term "microarray" refers to an arrangement of a plurality of polynucleotides, polypeptides, or other chemical compounds on a substrate.

The terms "element" and "array element" refer to a polynucleotide, polypeptide, or other chemical compound having a unique and defined position on a microarray.

The term "modulate" refers to a change in the activity of PKIN. For example, modulation may cause an increase or a decrease in protein activity, binding characteristics, or any other biological, functional, or immunological properties of PKIN.

The phrases "nucleic acid" and "nucleic acid sequence" refer to a nucleotide, oligonucleotide, polynucleotide, or any fragment thereof. These phrases also refer to DNA or RNA of genomic or synthetic origin which may be single-stranded or double-stranded and may represent the sense or the antisense strand, to peptide nucleic acid (PNA), or to any DNA-like or RNA-like material.

"Operably linked" refers to the situation in which a first nucleic acid sequence is placed in a functional relationship with a second nucleic acid sequence. For instance, a promoter is operably linked to a coding sequence if the promoter affects the transcription or expression of the coding sequence. Operably linked DNA sequences may be in close proximity or contiguous and, where necessary to join two protein coding regions, in the same reading frame.

"Peptide nucleic acid" (PNA) refers to an antisense molecule or anti-gene agent which comprises an oligonucleotide of at least about 5 nucleotides in length linked to a peptide backbone of

amino acid residues ending in lysine. The terminal lysine confers solubility to the composition. PNAs preferentially bind complementary single stranded DNA or RNA and stop transcript elongation, and may be pegylated to extend their lifespan in the cell.

"Post-translational modification" of an PKIN may involve lipidation, glycosylation, phosphorylation, acetylation, racemization, proteolytic cleavage, and other modifications known in the art. These processes may occur synthetically or biochemically. Biochemical modifications will vary by cell type depending on the enzymatic milieu of PKIN.

"Probe" refers to nucleic acid sequences encoding PKIN, their complements, or fragments thereof, which are used to detect identical, allelic or related nucleic acid sequences. Probes are isolated oligonucleotides or polynucleotides attached to a detectable label or reporter molecule. Typical labels include radioactive isotopes, ligands, chemiluminescent agents, and enzymes. "Primers" are short nucleic acids, usually DNA oligonucleotides, which may be annealed to a target polynucleotide by complementary base-pairing. The primer may then be extended along the target DNA strand by a DNA polymerase enzyme. Primer pairs can be used for amplification (and identification) of a nucleic acid sequence, e.g., by the polymerase chain reaction (PCR).

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Probes and primers as used in the present invention typically comprise at least 15 contiguous nucleotides of a known sequence. In order to enhance specificity, longer probes and primers may also be employed, such as probes and primers that comprise at least 20, 25, 30, 40, 50, 60, 70, 80, 90, 100, or at least 150 consecutive nucleotides of the disclosed nucleic acid sequences. Probes and primers may be considerably longer than these examples, and it is understood that any length supported by the specification, including the tables, figures, and Sequence Listing, may be used.

Methods for preparing and using probes and primers are described in the references, for example Sambrook, J. et al. (1989) Molecular Cloning: A Laboratory Manual, 2<sup>nd</sup> ed., vol. 1-3, Cold Spring Harbor Press, Plainview NY; Ausubel, F.M. et al. (1987) Current Protocols in Molecular Biology, Greene Publ. Assoc. & Wiley-Intersciences, New York NY; Innis, M. et al. (1990) PCR Protocols, A Guide to Methods and Applications, Academic Press, San Diego CA. PCR primer pairs can be derived from a known sequence, for example, by using computer programs intended for that purpose such as Primer (Version 0.5, 1991, Whitehead Institute for Biomedical Research, Cambridge MA).

Oligonucleotides for use as primers are selected using software known in the art for such purpose. For example, OLIGO 4.06 software is useful for the selection of PCR primer pairs of up to 100 nucleotides each, and for the analysis of oligonucleotides and larger polynucleotides of up to 5,000 nucleotides from an input polynucleotide sequence of up to 32 kilobases. Similar primer selection programs have incorporated additional features for expanded capabilities. For example, the PrimOU

primer selection program (available to the public from the Genome Center at University of Texas South West Medical Center, Dallas TX) is capable of choosing specific primers from megabase sequences and is thus useful for designing primers on a genome-wide scope. The Primer3 primer selection program (available to the public from the Whitehead Institute/MIT Center for Genome Research, Cambridge MA) allows the user to input a "mispriming library," in which sequences to avoid as primer binding sites are user-specified. Primer3 is useful, in particular, for the selection of oligonucleotides for microarrays. (The source code for the latter two primer selection programs may also be obtained from their respective sources and modified to meet the user's specific needs.) The PrimeGen program (available to the public from the UK Human Genome Mapping Project Resource Centre, Cambridge UK) designs primers based on multiple sequence alignments, thereby allowing selection of primers that hybridize to either the most conserved or least conserved regions of aligned nucleic acid sequences. Hence, this program is useful for identification of both unique and conserved oligonucleotides and polynucleotide fragments. The oligonucleotides and polynucleotide fragments identified by any of the above selection methods are useful in hybridization technologies, for example, as PCR or sequencing primers, microarray elements, or specific probes to identify fully or partially complementary polynucleotides in a sample of nucleic acids. Methods of oligonucleotide selection are not limited to those described above.

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A "recombinant nucleic acid" is a sequence that is not naturally occurring or has a sequence that is made by an artificial combination of two or more otherwise separated segments of sequence. This artificial combination is often accomplished by chemical synthesis or, more commonly, by the artificial manipulation of isolated segments of nucleic acids, e.g., by genetic engineering techniques such as those described in Sambrook, <u>supra</u>. The term recombinant includes nucleic acids that have been altered solely by addition, substitution, or deletion of a portion of the nucleic acid. Frequently, a recombinant nucleic acid may include a nucleic acid sequence operably linked to a promoter sequence. Such a recombinant nucleic acid may be part of a vector that is used, for example, to transform a cell.

Alternatively, such recombinant nucleic acids may be part of a viral vector, e.g., based on a vaccinia virus, that could be use to vaccinate a mammal wherein the recombinant nucleic acid is expressed, inducing a protective immunological response in the mammal.

A "regulatory element" refers to a nucleic acid sequence usually derived from untranslated regions of a gene and includes enhancers, promoters, introns, and 5' and 3' untranslated regions (UTRs). Regulatory elements interact with host or viral proteins which control transcription, translation, or RNA stability.

"Reporter molecules" are chemical or biochemical moieties used for labeling a nucleic acid, amino acid, or antibody. Reporter molecules include radionuclides; enzymes; fluorescent,

chemiluminescent, or chromogenic agents; substrates; cofactors; inhibitors; magnetic particles; and other moieties known in the art.

An "RNA equivalent," in reference to a DNA sequence, is composed of the same linear sequence of nucleotides as the reference DNA sequence with the exception that all occurrences of the nitrogenous base thymine are replaced with uracil, and the sugar backbone is composed of ribose instead of deoxyribose.

The term "sample" is used in its broadest sense. A sample suspected of containing PKIN, nucleic acids encoding PKIN, or fragments thereof may comprise a bodily fluid; an extract from a cell, chromosome, organelle, or membrane isolated from a cell; a cell; genomic DNA, RNA, or cDNA, in solution or bound to a substrate; a tissue; a tissue print; etc.

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The terms "specific binding" and "specifically binding" refer to that interaction between a protein or peptide and an agonist, an antibody, an antagonist, a small molecule, or any natural or synthetic binding composition. The interaction is dependent upon the presence of a particular structure of the protein, e.g., the antigenic determinant or epitope, recognized by the binding molecule. For example, if an antibody is specific for epitope "A," the presence of a polypeptide comprising the epitope A, or the presence of free unlabeled A, in a reaction containing free labeled A and the antibody will reduce the amount of labeled A that binds to the antibody.

The term "substantially purified" refers to nucleic acid or amino acid sequences that are removed from their natural environment and are isolated or separated, and are at least 60% free, preferably at least 75% free, and most preferably at least 90% free from other components with which they are naturally associated.

A "substitution" refers to the replacement of one or more amino acid residues or nucleotides by different amino acid residues or nucleotides, respectively.

"Substrate" refers to any suitable rigid or semi-rigid support including membranes, filters, chips, slides, wafers, fibers, magnetic or nonmagnetic beads, gels, tubing, plates, polymers, microparticles and capillaries. The substrate can have a variety of surface forms, such as wells, trenches, pins, channels and pores, to which polynucleotides or polypeptides are bound.

A "transcript image" refers to the collective pattern of gene expression by a particular cell type or tissue under given conditions at a given time.

"Transformation" describes a process by which exogenous DNA is introduced into a recipient cell. Transformation may occur under natural or artificial conditions according to various methods well known in the art, and may rely on any known method for the insertion of foreign nucleic acid sequences into a prokaryotic or eukaryotic host cell. The method for transformation is selected based on the type of host cell being transformed and may include, but is not limited to, bacteriophage or viral infection,

electroporation, heat shock, lipofection, and particle bombardment. The term "transformed cells" includes stably transformed cells in which the inserted DNA is capable of replication either as an autonomously replicating plasmid or as part of the host chromosome, as well as transiently transformed cells which express the inserted DNA or RNA for limited periods of time.

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A "transgenic organism," as used herein, is any organism, including but not limited to animals and plants, in which one or more of the cells of the organism contains heterologous nucleic acid introduced by way of human intervention, such as by transgenic techniques well known in the art. The nucleic acid is introduced into the cell, directly or indirectly by introduction into a precursor of the cell, by way of deliberate genetic manipulation, such as by microinjection or by infection with a recombinant virus. The term genetic manipulation does not include classical cross-breeding, or <u>in vitro</u> fertilization, but rather is directed to the introduction of a recombinant DNA molecule. The transgenic organisms contemplated in accordance with the present invention include bacteria, cyanobacteria, fungi, plants and animals. The isolated DNA of the present invention can be introduced into the host by methods known in the art, for example infection, transfection, transformation or transconjugation. Techniques for transferring the DNA of the present invention into such organisms are widely known and provided in references such as Sambrook et al. (1989), supra.

A "variant" of a particular nucleic acid sequence is defined as a nucleic acid sequence having at least 40% sequence identity to the particular nucleic acid sequence over a certain length of one of the nucleic acid sequences using blastn with the "BLAST 2 Sequences" tool Version 2.0.9 (May-07-1999) set at default parameters. Such a pair of nucleic acids may show, for example, at least 50%, at least 60%, at least 70%, at least 80%, at least 85%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% or greater sequence identity over a certain defined length. A variant may be described as, for example, an "allelic" (as defined above), "splice," "species," or "polymorphic" variant. A splice variant may have significant identity to a reference molecule, but will generally have a greater or lesser number of polynucleotides due to alternative splicing of exons during mRNA processing. The corresponding polypeptide may possess additional functional domains or lack domains that are present in the reference molecule. Species variants are polynucleotide sequences that vary from one species to another. The resulting polypeptides will generally have significant amino acid identity relative to each other. A polymorphic variant is a variation in the polynucleotide sequence of a particular gene between individuals of a given species. Polymorphic variants also may encompass "single nucleotide polymorphisms" (SNPs) in which the polynucleotide sequence varies by one nucleotide base. The presence of SNPs may be indicative of, for example, a certain population, a disease state, or a propensity for a disease state.

A "variant" of a particular polypeptide sequence is defined as a polypeptide sequence having at least 40% sequence identity to the particular polypeptide sequence over a certain length of one of the polypeptide sequences using blastp with the "BLAST 2 Sequences" tool Version 2.0.9 (May-07-1999) set at default parameters. Such a pair of polypeptides may show, for example, at least 50%, at least 60%, at least 70%, at least 80%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% or greater sequence identity over a certain defined length of one of the polypeptides.

#### THE INVENTION

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The invention is based on the discovery of new human human kinases (PKIN), the polynucleotides encoding PKIN, and the use of these compositions for the diagnosis, treatment, or prevention of cancer, immune disorders, disorders affecting growth and development, cardiovascular diseases, and lipid disorders.

Table 1 summarizes the nomenclature for the full length polynucleotide and polypeptide sequences of the invention. Each polynucleotide and its corresponding polypeptide are correlated to a single Incyte project identification number (Incyte Project ID). Each polypeptide sequence is denoted by both a polypeptide sequence identification number (Polypeptide SEQ ID NO:) and an Incyte polypeptide sequence number (Incyte Polypeptide ID) as shown. Each polynucleotide sequence is denoted by both a polynucleotide sequence identification number (Polynucleotide SEQ ID NO:) and an Incyte polynucleotide consensus sequence number (Incyte Polynucleotide ID) as shown.

Table 2 shows sequences with homology to the polypeptides of the invention as identified by BLAST analysis against the GenBank protein (genpept) database. Columns 1 and 2 show the polypeptide sequence identification number (Polypeptide SEQ ID NO:) and the corresponding Incyte polypeptide sequence number (Incyte Polypeptide ID) for polypeptides of the invention. Column 3 shows the GenBank identification number (Genbank ID NO:) of the nearest GenBank homolog. Column 4 shows the probability score for the match between each polypeptide and its GenBank homolog. Column 5 shows the annotation of the GenBank homolog along with relevant citations where applicable, all of which are expressly incorporated by reference herein.

Table 3 shows various structural features of the polypeptides of the invention. Columns 1 and 2 show the polypeptide sequence identification number (SEQ ID NO:) and the corresponding Incyte polypeptide sequence number (Incyte Polypeptide ID) for each polypeptide of the invention. Column 3 shows the number of amino acid residues in each polypeptide. Column 4 shows potential phosphorylation sites, and column 5 shows potential glycosylation sites, as determined by the MOTIFS program of the GCG sequence analysis software package (Genetics Computer Group, Madison WI).

Column 6 shows amino acid residues comprising signature sequences, domains, and motifs. Column 7 shows analytical methods for protein structure/function analysis and in some cases, searchable databases to which the analytical methods were applied.

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Together, Tables 2 and 3 summarize the properties of polypeptides of the invention, and these properties establish that the claimed polypeptides are human kinases. For example, SEQ ID NO:4 is 94% identical to rat serine/threonine kinase (GenBank ID g2052189) as determined by the Basic Local Alignment Search Tool (BLAST). (See Table 2.) The BLAST probability score is 0.0, which indicates the probability of obtaining the observed polypeptide sequence alignment by chance. SEQ ID NO:4 also contains a protein kinase domain as determined by searching for statistically significant matches in the hidden Markov model (HMM)-based PFAM database of conserved protein family domains. (See Table 3.) Data from BLIMPS, MOTIFS, and PROFILESCAN analyses provide further corroborative evidence that SEQ ID NO:4 is a protein kinase. In an alternate example, SEQ ID NO: 23 is 88% identical to murine protein kinase (GenBank ID g406058) as determined by the Basic Local Alignment Search Tool (BLAST). (See Table 2.) The BLAST probability score is 0.0, which indicates the probability of obtaining the observed polypeptide sequence alignment by chance. SEQ ID NO:23 also contains an eukaryotic protein kinase domain as determined by searching for statistically significant matches in the hidden Markov model (HMM)-based PFAM database of conserved protein family domains. (See Table 3.) Data from BLIMPS, MOTIFS, and PROFILESCAN analyses provide further corroborative evidence that SEQ ID NO:23 is a protein kinase. In an alternate example, SEQ ID NO:6 is 85% identical to rabbit myosin light chain kinase (GenBank ID g165506) as determined by the Basic Local Alignment Search Tool (BLAST). (See Table 2.) The BLAST probability score is 1.5e-272, which indicates the probability of obtaining the observed polypeptide sequence alignment by chance. SEQ ID NO:6 also contains a eukaryotic protein kinase domain as determined by searching for statistically significant matches in the hidden Markov model (HMM)-based PFAM database of conserved protein family domains. (See Table 3.) Data from BLIMPS and MOTIFS analyses provide further corroborative evidence that SEQ ID NO:6 is a myosin light chain kinase. In an alternate example, SEQ ID NO:1 is 64% identical to murine serine/threonine kinase (GenBank ID g404634) as determined by the Basic Local Alignment Search Tool (BLAST). (See Table 2.) The BLAST probability score is 4.5e-60, which indicates the probability of obtaining the observed polypeptide sequence alignment by chance. SEQ ID NO:1 also contains a protein kinase domain as determined by searching for statistically significant matches in the hidden Markov model (HMM)-based PFAM database of conserved protein family domains. (See Table 3.) Data from MOTIFS, BLIMPS and PROFILESCAN analyses provide further corroborative evidence that SEQ ID NO:1 is a protein kinase, notably a serine/threonine kinase. In an alternate example, SEQ ID NO:19 is 49% identical to

human G-protein-coupled receptor kinase GRK4-beta (GenBank ID g992672) as determined by the Basic Local Alignment Search Tool (BLAST). (See Table 2.) The BLAST probability score is 4.3e-129, which indicates the probability of obtaining the observed polypeptide sequence alignment by chance. SEQ ID NO:19 also contains a regulator of G-protein signaling domain as determined by searching for statistically significant matches in the hidden Markov model (HMM)-based PFAM database of conserved protein family domains. (See Table 3.) Data from BLIMPS, MOTIFS, and PROFILESCAN analyses provide further corroborative evidence that SEQ ID NO:19 is a G-protein-coupled receptor kinase. SEQ ID NO:2-3, SEQ ID NO:5, SEQ ID NO:7-18, SEQ ID NO:20-22 and SEQ ID NO:24-26 were analyzed and annotated in a similar manner. The algorithms and parameters for the analysis of SEQ ID NO:1-26 are described in Table 7.

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As shown in Table 4, the full length polynucleotide sequences of the present invention were assembled using cDNA sequences or coding (exon) sequences derived from genomic DNA, or any combination of these two types of sequences. Columns 1 and 2 list the polynucleotide sequence identification number (Polynucleotide SEQ ID NO:) and the corresponding Incyte polynucleotide consensus sequence number (Incyte Polynucleotide ID) for each polynucleotide of the invention. Column 3 shows the length of each polynucleotide sequence in basepairs. Column 4 lists fragments of the polynucleotide sequences which are useful, for example, in hybridization or amplification technologies that identify SEQ ID NO:27-52 or that distinguish between SEQ ID NO:27-52 and related polynucleotide sequences. Column 5 shows identification numbers corresponding to cDNA sequences, coding sequences (exons) predicted from genomic DNA, and/or sequence assemblages comprised of both cDNA and genomic DNA. These sequences were used to assemble the full length polynucleotide sequences of the invention. Columns 6 and 7 of Table 4 show the nucleotide start (5') and stop (3') positions of the cDNA and/or genomic sequences in column 5 relative to their respective full length sequences.

The identification numbers in Column 5 of Table 4 may refer specifically, for example, to Incyte cDNAs along with their corresponding cDNA libraries. For example, 6829315H1 is the identification number of an Incyte cDNA sequence, and SINTNOR01 is the cDNA library from which it is derived. Incyte cDNAs for which cDNA libraries are not indicated were derived from pooled cDNA libraries (e.g., 55057226H1). Alternatively, the identification numbers in column 5 may refer to GenBank cDNAs or ESTs (e.g., g2954208) which contributed to the assembly of the full length polynucleotide sequences. In addition, the identification numbers in column 5 may identify sequences derived from the ENSEMBL (The Sanger Centre, Cambridge, UK) database (*i.e.*, those sequences including the designation "ENST"). Alternatively, the identification numbers in column 5 may be derived from the NCBI RefSeq Nucleotide Sequence Records Database (*i.e.*, those sequences including

the designation "NM" or "NT") or the NCBI RefSeq Protein Sequence Records (i.e., those sequences including the designation "NP"). Alternatively, the identification numbers in column 5 may refer to assemblages of both cDNA and Genscan-predicted exons brought together by an "exon stitching" algorithm. For example,  $FL_XXXXXX_N_1_N_2_YYYYY_N_3_N_4$  represents a "stitched" sequence in which XXXXXX is the identification number of the cluster of sequences to which the algorithm was applied, and YYYYY is the number of the prediction generated by the algorithm, and  $N_{1,2,3,...}$ , if present, represent specific exons that may have been manually edited during analysis (See Example V). Alternatively, the identification numbers in column 5 may refer to assemblages of exons brought together by an "exon-stretching" algorithm. For example, FLXXXXXX\_gAAAAA\_gBBBBB\_1\_N is the identification number of a "stretched" sequence, with XXXXXX being the Incyte project identification number, gAAAAA being the GenBank identification number of the human genomic sequence to which the "exon-stretching" algorithm was applied, gBBBBB being the GenBank identification number or NCBI RefSeq identification number of the nearest GenBank protein homolog, and N referring to specific exons (See Example V). In instances where a RefSeq sequence was used as a protein homolog for the "exon-stretching" algorithm, a RefSeq identifier (denoted by "NM," "NP," or "NT") may be used in place of the GenBank identifier (i.e., gBBBBB).

Alternatively, a prefix identifies component sequences that were hand-edited, predicted from genomic DNA sequences, or derived from a combination of sequence analysis methods. The following Table lists examples of component sequence prefixes and corresponding sequence analysis methods associated with the prefixes (see Example IV and Example V).

Prefix	Type of analysis and/or examples of programs	
GNN, GFG,	Exon prediction from genomic sequences using, for example,	
ENST	GENSCAN (Stanford University, CA, USA) or FGENES	
	(Computer Genomics Group, The Sanger Centre, Cambridge, UK).	
GBI	Hand-edited analysis of genomic sequences.	
FL	Stitched or stretched genomic sequences (see Example V).	

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In some cases, Incyte cDNA coverage redundant with the sequence coverage shown in column 5 was obtained to confirm the final consensus polynucleotide sequence, but the relevant Incyte cDNA identification numbers are not shown.

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Table 5 shows the representative cDNA libraries for those full length polynucleotide sequences which were assembled using Incyte cDNA sequences. The representative cDNA library is the Incyte cDNA library which is most frequently represented by the Incyte cDNA sequences which were used to

assemble and confirm the above polynucleotide sequences. The tissues and vectors which were used to construct the cDNA libraries shown in Table 5 are described in Table 6.

The invention also encompasses PKIN variants. A preferred PKIN variant is one which has at least about 80%, or alternatively at least about 90%, or even at least about 95% amino acid sequence identity to the PKIN amino acid sequence, and which contains at least one functional or structural characteristic of PKIN.

The invention also encompasses polynucleotides which encode PKIN. In a particular embodiment, the invention encompasses a polynucleotide sequence comprising a sequence selected from the group consisting of SEQ ID NO:27-52, which encodes PKIN. The polynucleotide sequences of SEQ ID NO:27-52, as presented in the Sequence Listing, embrace the equivalent RNA sequences, wherein occurrences of the nitrogenous base thymine are replaced with uracil, and the sugar backbone is composed of ribose instead of deoxyribose.

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The invention also encompasses a variant of a polynucleotide sequence encoding PKIN. In particular, such a variant polynucleotide sequence will have at least about 70%, or alternatively at least about 85%, or even at least about 95% polynucleotide sequence identity to the polynucleotide sequence encoding PKIN. A particular aspect of the invention encompasses a variant of a polynucleotide sequence comprising a sequence selected from the group consisting of SEQ ID NO:27-52 which has at least about 70%, or alternatively at least about 85%, or even at least about 95% polynucleotide sequence identity to a nucleic acid sequence selected from the group consisting of SEQ ID NO:27-52. Any one of the polynucleotide variants described above can encode an amino acid sequence which contains at least one functional or structural characteristic of PKIN.

It will be appreciated by those skilled in the art that as a result of the degeneracy of the genetic code, a multitude of polynucleotide sequences encoding PKIN, some bearing minimal similarity to the polynucleotide sequences of any known and naturally occurring gene, may be produced. Thus, the invention contemplates each and every possible variation of polynucleotide sequence that could be made by selecting combinations based on possible codon choices. These combinations are made in accordance with the standard triplet genetic code as applied to the polynucleotide sequence of naturally occurring PKIN, and all such variations are to be considered as being specifically disclosed.

Although nucleotide sequences which encode PKIN and its variants are generally capable of hybridizing to the nucleotide sequence of the naturally occurring PKIN under appropriately selected conditions of stringency, it may be advantageous to produce nucleotide sequences encoding PKIN or its derivatives possessing a substantially different codon usage, e.g., inclusion of non-naturally occurring codons. Codons may be selected to increase the rate at which expression of the peptide occurs in a particular prokaryotic or eukaryotic host in accordance with the frequency with which particular codons

are utilized by the host. Other reasons for substantially altering the nucleotide sequence encoding PKIN and its derivatives without altering the encoded amino acid sequences include the production of RNA transcripts having more desirable properties, such as a greater half-life, than transcripts produced from the naturally occurring sequence.

The invention also encompasses production of DNA sequences which encode PKIN and PKIN derivatives, or fragments thereof, entirely by synthetic chemistry. After production, the synthetic sequence may be inserted into any of the many available expression vectors and cell systems using reagents well known in the art. Moreover, synthetic chemistry may be used to introduce mutations into a sequence encoding PKIN or any fragment thereof.

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Also encompassed by the invention are polynucleotide sequences that are capable of hybridizing to the claimed polynucleotide sequences, and, in particular, to those shown in SEQ ID NO:27-52 and fragments thereof under various conditions of stringency. (See, e.g., Wahl, G.M. and S.L. Berger (1987) Methods Enzymol. 152:399-407; Kimmel, A.R. (1987) Methods Enzymol. 152:507-511.) Hybridization conditions, including annealing and wash conditions, are described in "Definitions."

Methods for DNA sequencing are well known in the art and may be used to practice any of the embodiments of the invention. The methods may employ such enzymes as the Klenow fragment of DNA polymerase I, SEQUENASE (US Biochemical, Cleveland OH), Taq polymerase (Applied Biosystems), thermostable T7 polymerase (Amersham Pharmacia Biotech, Piscataway NJ), or combinations of polymerases and proofreading exonucleases such as those found in the ELONGASE amplification system (Life Technologies, Gaithersburg MD). Preferably, sequence preparation is automated with machines such as the MICROLAB 2200 liquid transfer system (Hamilton, Reno NV), PTC200 thermal cycler (MJ Research, Watertown MA) and ABI CATALYST 800 thermal cycler (Applied Biosystems). Sequencing is then carried out using either the ABI 373 or 377 DNA sequencing system (Applied Biosystems), the MEGABACE 1000 DNA sequencing system (Molecular Dynamics, Sunnyvale CA), or other systems known in the art. The resulting sequences are analyzed using a variety of algorithms which are well known in the art. (See, e.g., Ausubel, F.M. (1997) Short Protocols in Molecular Biology, John Wiley & Sons, New York NY, unit 7.7; Meyers, R.A. (1995) Molecular Biology and Biotechnology, Wiley VCH, New York NY, pp. 856-853.)

The nucleic acid sequences encoding PKIN may be extended utilizing a partial nucleotide sequence and employing various PCR-based methods known in the art to detect upstream sequences, such as promoters and regulatory elements. For example, one method which may be employed, restriction-site PCR, uses universal and nested primers to amplify unknown sequence from genomic DNA within a cloning vector. (See, e.g., Sarkar, G. (1993) PCR Methods Applic. 2:318-322.)

Another method, inverse PCR, uses primers that extend in divergent directions to amplify unknown sequence from a circularized template. The template is derived from restriction fragments comprising a known genomic locus and surrounding sequences. (See, e.g., Triglia, T. et al. (1988) Nucleic Acids Res. 16:8186.) A third method, capture PCR, involves PCR amplification of DNA fragments adjacent to known sequences in human and yeast artificial chromosome DNA. (See, e.g., Lagerstrom, M. et al. (1991) PCR Methods Applic. 1:111-119.) In this method, multiple restriction enzyme digestions and ligations may be used to insert an engineered double-stranded sequence into a region of unknown sequence before performing PCR. Other methods which may be used to retrieve unknown sequences are known in the art. (See, e.g., Parker, J.D. et al. (1991) Nucleic Acids Res. 19:3055-3060). Additionally, one may use PCR, nested primers, and PROMOTERFINDER libraries (Clontech, Palo Alto CA) to walk genomic DNA. This procedure avoids the need to screen libraries and is useful in finding intron/exon junctions. For all PCR-based methods, primers may be designed using commercially available software, such as OLIGO 4.06 primer analysis software (National Biosciences, Plymouth MN) or another appropriate program, to be about 22 to 30 nucleotides in length, to have a GC content of about 50% or more, and to anneal to the template at temperatures of about 68°C to 72°C.

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When screening for full length cDNAs, it is preferable to use libraries that have been size-selected to include larger cDNAs. In addition, random-primed libraries, which often include sequences containing the 5' regions of genes, are preferable for situations in which an oligo d(T) library does not yield a full-length cDNA. Genomic libraries may be useful for extension of sequence into 5' non-transcribed regulatory regions.

Capillary electrophoresis systems which are commercially available may be used to analyze the size or confirm the nucleotide sequence of sequencing or PCR products. In particular, capillary sequencing may employ flowable polymers for electrophoretic separation, four different nucleotide-specific, laser-stimulated fluorescent dyes, and a charge coupled device camera for detection of the emitted wavelengths. Output/light intensity may be converted to electrical signal using appropriate software (e.g., GENOTYPER and SEQUENCE NAVIGATOR, Applied Biosystems), and the entire process from loading of samples to computer analysis and electronic data display may be computer controlled. Capillary electrophoresis is especially preferable for sequencing small DNA fragments which may be present in limited amounts in a particular sample.

In another embodiment of the invention, polynucleotide sequences or fragments thereof which encode PKIN may be cloned in recombinant DNA molecules that direct expression of PKIN, or fragments or functional equivalents thereof, in appropriate host cells. Due to the inherent degeneracy of

the genetic code, other DNA sequences which encode substantially the same or a functionally equivalent amino acid sequence may be produced and used to express PKIN.

The nucleotide sequences of the present invention can be engineered using methods generally known in the art in order to alter PKIN-encoding sequences for a variety of purposes including, but not limited to, modification of the cloning, processing, and/or expression of the gene product. DNA shuffling by random fragmentation and PCR reassembly of gene fragments and synthetic oligonucleotides may be used to engineer the nucleotide sequences. For example, oligonucleotide-mediated site-directed mutagenesis may be used to introduce mutations that create new restriction sites, alter glycosylation patterns, change codon preference, produce splice variants, and so forth.

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The nucleotides of the present invention may be subjected to DNA shuffling techniques such as MOLECULARBREEDING (Maxygen Inc., Santa Clara CA; described in U.S. Patent Number 5,837,458; Chang, C.-C. et al. (1999) Nat. Biotechnol. 17:793-797; Christians, F.C. et al. (1999) Nat. Biotechnol. 17:259-264; and Crameri, A. et al. (1996) Nat. Biotechnol. 14:315-319) to alter or improve the biological properties of PKIN, such as its biological or enzymatic activity or its ability to bind to other molecules or compounds. DNA shuffling is a process by which a library of gene variants is produced using PCR-mediated recombination of gene fragments. The library is then subjected to selection or screening procedures that identify those gene variants with the desired properties. These preferred variants may then be pooled and further subjected to recursive rounds of DNA shuffling and selection/screening. Thus, genetic diversity is created through "artificial" breeding and rapid molecular evolution. For example, fragments of a single gene containing random point mutations may be recombined, screened, and then reshuffled until the desired properties are optimized. Alternatively, fragments of a given gene may be recombined with fragments of homologous genes in the same gene family, either from the same or different species, thereby maximizing the genetic diversity of multiple naturally occurring genes in a directed and controllable manner.

In another embodiment, sequences encoding PKIN may be synthesized, in whole or in part, using chemical methods well known in the art. (See, e.g., Caruthers, M.H. et al. (1980) Nucleic Acids Symp. Ser. 7:215-223; and Horn, T. et al. (1980) Nucleic Acids Symp. Ser. 7:225-232.) Alternatively, PKIN itself or a fragment thereof may be synthesized using chemical methods. For example, peptide synthesis can be performed using various solution-phase or solid-phase techniques. (See, e.g., Creighton, T. (1984) Proteins, Structures and Molecular Properties, WH Freeman, New York NY, pp. 55-60; and Roberge, J.Y. et al. (1995) Science 269:202-204.) Automated synthesis may be achieved using the ABI 431A peptide synthesizer (Applied Biosystems). Additionally, the amino acid sequence of PKIN, or any part thereof, may be altered during direct synthesis and/or combined with sequences

from other proteins, or any part thereof, to produce a variant polypeptide or a polypeptide having a sequence of a naturally occurring polypeptide.

The peptide may be substantially purified by preparative high performance liquid chromatography. (See, e.g., Chiez, R.M. and F.Z. Regnier (1990) Methods Enzymol. 182:392-421.) The composition of the synthetic peptides may be confirmed by amino acid analysis or by sequencing. (See, e.g., Creighton, supra, pp. 28-53.)

In order to express a biologically active PKIN, the nucleotide sequences encoding PKIN or derivatives thereof may be inserted into an appropriate expression vector, i.e., a vector which contains the necessary elements for transcriptional and translational control of the inserted coding sequence in a suitable host. These elements include regulatory sequences, such as enhancers, constitutive and inducible promoters, and 5' and 3' untranslated regions in the vector and in polynucleotide sequences encoding PKIN. Such elements may vary in their strength and specificity. Specific initiation signals may also be used to achieve more efficient translation of sequences encoding PKIN. Such signals include the ATG initiation codon and adjacent sequences, e.g. the Kozak sequence. In cases where sequences encoding PKIN and its initiation codon and upstream regulatory sequences are inserted into the appropriate expression vector, no additional transcriptional or translational control signals may be needed. However, in cases where only coding sequence, or a fragment thereof, is inserted, exogenous translational control signals including an in-frame ATG initiation codon should be provided by the vector. Exogenous translational elements and initiation codons may be of various origins, both natural and synthetic. The efficiency of expression may be enhanced by the inclusion of enhancers appropriate for the particular host cell system used. (See, e.g., Scharf, D. et al. (1994) Results Probl. Cell Differ. 20:125-162.)

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Methods which are well known to those skilled in the art may be used to construct expression vectors containing sequences encoding PKIN and appropriate transcriptional and translational control elements. These methods include <u>in vitro</u> recombinant DNA techniques, synthetic techniques, and <u>in vivo</u> genetic recombination. (See, e.g., Sambrook, J. et al. (1989) <u>Molecular Cloning, A Laboratory Manual</u>, Cold Spring Harbor Press, Plainview NY, ch. 4, 8, and 16-17; Ausubel, F.M. et al. (1995) <u>Current Protocols in Molecular Biology</u>, John Wiley & Sons, New York NY, ch. 9, 13, and 16.)

A variety of expression vector/host systems may be utilized to contain and express sequences encoding PKIN. These include, but are not limited to, microorganisms such as bacteria transformed with recombinant bacteriophage, plasmid, or cosmid DNA expression vectors; yeast transformed with yeast expression vectors; insect cell systems infected with viral expression vectors (e.g., baculovirus); plant cell systems transformed with viral expression vectors (e.g., cauliflower mosaic virus, CaMV, or tobacco mosaic virus, TMV) or with bacterial expression vectors (e.g., Ti or pBR322 plasmids); or

animal cell systems. (See, e.g., Sambrook, supra; Ausubel, supra; Van Heeke, G. and S.M. Schuster (1989) J. Biol. Chem. 264:5503-5509; Engelhard, E.K. et al. (1994) Proc. Natl. Acad. Sci. USA 91:3224-3227; Sandig, V. et al. (1996) Hum. Gene Ther. 7:1937-1945; Takamatsu, N. (1987) EMBO J. 6:307-311; The McGraw Hill Yearbook of Science and Technology (1992) McGraw Hill, New York NY, pp. 191-196; Logan, J. and T. Shenk (1984) Proc. Natl. Acad. Sci. USA 81:3655-3659; and Harrington, J.J. et al. (1997) Nat. Genet. 15:345-355.) Expression vectors derived from retroviruses, adenoviruses, or herpes or vaccinia viruses, or from various bacterial plasmids, may be used for delivery of nucleotide sequences to the targeted organ, tissue, or cell population. (See, e.g., Di Nicola, M. et al. (1998) Cancer Gen. Ther. 5(6):350-356; Yu, M. et al. (1993) Proc. Natl. Acad. Sci. USA 90(13):6340-6344; Buller, R.M. et al. (1985) Nature 317(6040):813-815; McGregor, D.P. et al. (1994) Mol. Immunol. 31(3):219-226; and Verma, I.M. and N. Somia (1997) Nature 389:239-242.) The invention is not limited by the host cell employed.

In bacterial systems, a number of cloning and expression vectors may be selected depending upon the use intended for polynucleotide sequences encoding PKIN. For example, routine cloning, subcloning, and propagation of polynucleotide sequences encoding PKIN can be achieved using a multifunctional E. coli vector such as PBLUESCRIPT (Stratagene, La Jolla CA) or PSPORT1 plasmid (Life Technologies). Ligation of sequences encoding PKIN into the vector's multiple cloning site disrupts the *lacZ* gene, allowing a colorimetric screening procedure for identification of transformed bacteria containing recombinant molecules. In addition, these vectors may be useful for <u>in vitro</u> transcription, dideoxy sequencing, single strand rescue with helper phage, and creation of nested deletions in the cloned sequence. (See, e.g., Van Heeke, G. and S.M. Schuster (1989) J. Biol. Chem. 264:5503-5509.) When large quantities of PKIN are needed, e.g. for the production of antibodies, vectors which direct high level expression of PKIN may be used. For example, vectors containing the strong, inducible SP6 or T7 bacteriophage promoter may be used.

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Yeast expression systems may be used for production of PKIN. A number of vectors containing constitutive or inducible promoters, such as alpha factor, alcohol oxidase, and PGH promoters, may be used in the yeast <u>Saccharomyces cerevisiae</u> or <u>Pichia pastoris</u>. In addition, such vectors direct either the secretion or intracellular retention of expressed proteins and enable integration of foreign sequences into the host genome for stable propagation. (See, e.g., Ausubel, 1995, <u>supra</u>; Bitter, G.A. et al. (1987) Methods Enzymol. 153:516-544; and Scorer, C.A. et al. (1994) Bio/Technology 12:181-184.)

Plant systems may also be used for expression of PKIN. Transcription of sequences encoding PKIN may be driven by viral promoters, e.g., the 35S and 19S promoters of CaMV used alone or in combination with the omega leader sequence from TMV (Takamatsu, N. (1987) EMBO J. 6:307-311).

Alternatively, plant promoters such as the small subunit of RUBISCO or heat shock promoters may be used. (See, e.g., Coruzzi, G. et al. (1984) EMBO J. 3:1671-1680; Broglie, R. et al. (1984) Science 224:838-843; and Winter, J. et al. (1991) Results Probl. Cell Differ. 17:85-105.) These constructs can be introduced into plant cells by direct DNA transformation or pathogen-mediated transfection. (See, e.g., The McGraw Hill Yearbook of Science and Technology (1992) McGraw Hill, New York NY, pp. 191-196.)

In mammalian cells, a number of viral-based expression systems may be utilized. In cases where an adenovirus is used as an expression vector, sequences encoding PKIN may be ligated into an adenovirus transcription/translation complex consisting of the late promoter and tripartite leader sequence. Insertion in a non-essential E1 or E3 region of the viral genome may be used to obtain infective virus which expresses PKIN in host cells. (See, e.g., Logan, J. and T. Shenk (1984) Proc. Natl. Acad. Sci. USA 81:3655-3659.) In addition, transcription enhancers, such as the Rous sarcoma virus (RSV) enhancer, may be used to increase expression in mammalian host cells. SV40 or EBV-based vectors may also be used for high-level protein expression.

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Human artificial chromosomes (HACs) may also be employed to deliver larger fragments of DNA than can be contained in and expressed from a plasmid. HACs of about 6 kb to 10 Mb are constructed and delivered via conventional delivery methods (liposomes, polycationic amino polymers, or vesicles) for therapeutic purposes. (See, e.g., Harrington, J.J. et al. (1997) Nat. Genet. 15:345-355.)

For long term production of recombinant proteins in mammalian systems, stable expression of PKIN in cell lines is preferred. For example, sequences encoding PKIN can be transformed into cell lines using expression vectors which may contain viral origins of replication and/or endogenous expression elements and a selectable marker gene on the same or on a separate vector. Following the introduction of the vector, cells may be allowed to grow for about 1 to 2 days in enriched media before being switched to selective media. The purpose of the selectable marker is to confer resistance to a selective agent, and its presence allows growth and recovery of cells which successfully express the introduced sequences. Resistant clones of stably transformed cells may be propagated using tissue culture techniques appropriate to the cell type.

Any number of selection systems may be used to recover transformed cell lines. These include, but are not limited to, the herpes simplex virus thymidine kinase and adenine phosphoribosyltransferase genes, for use in  $tk^-$  and  $apr^-$  cells, respectively. (See, e.g., Wigler, M. et al. (1977) Cell 11:223-232; Lowy, I. et al. (1980) Cell 22:817-823.) Also, antimetabolite, antibiotic, or herbicide resistance can be used as the basis for selection. For example, dhfr confers resistance to methotrexate; neo confers resistance to the aminoglycosides neomycin and G-418; and als and pat confer resistance to chlorsulfuron and phosphinotricin acetyltransferase, respectively. (See, e.g., Wigler, M. et al. (1980)

Proc. Natl. Acad. Sci. USA 77:3567-3570; Colbere-Garapin, F. et al. (1981) J. Mol. Biol. 150:1-14.) Additional selectable genes have been described, e.g., *trpB* and *hisD*, which alter cellular requirements for metabolites. (See, e.g., Hartman, S.C. and R.C. Mulligan (1988) Proc. Natl. Acad. Sci. USA 85:8047-8051.) Visible markers, e.g., anthocyanins, green fluorescent proteins (GFP; Clontech), β glucuronidase and its substrate β-glucuronide, or luciferase and its substrate luciferin may be used. These markers can be used not only to identify transformants, but also to quantify the amount of transient or stable protein expression attributable to a specific vector system. (See, e.g., Rhodes, C.A. (1995) Methods Mol. Biol. 55:121-131.)

Although the presence/absence of marker gene expression suggests that the gene of interest is also present, the presence and expression of the gene may need to be confirmed. For example, if the sequence encoding PKIN is inserted within a marker gene sequence, transformed cells containing sequences encoding PKIN can be identified by the absence of marker gene function. Alternatively, a marker gene can be placed in tandem with a sequence encoding PKIN under the control of a single promoter. Expression of the marker gene in response to induction or selection usually indicates expression of the tandem gene as well.

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In general, host cells that contain the nucleic acid sequence encoding PKIN and that express PKIN may be identified by a variety of procedures known to those of skill in the art. These procedures include, but are not limited to, DNA-DNA or DNA-RNA hybridizations, PCR amplification, and protein bioassay or immunoassay techniques which include membrane, solution, or chip based technologies for the detection and/or quantification of nucleic acid or protein sequences.

Immunological methods for detecting and measuring the expression of PKIN using either specific polyclonal or monoclonal antibodies are known in the art. Examples of such techniques include enzyme-linked immunosorbent assays (ELISAs), radioimmunoassays (RIAs), and fluorescence activated cell sorting (FACS). A two-site, monoclonal-based immunoassay utilizing monoclonal antibodies reactive to two non-interfering epitopes on PKIN is preferred, but a competitive binding assay may be employed. These and other assays are well known in the art. (See, e.g., Hampton, R. et al. (1990) Serological Methods, a Laboratory Manual, APS Press, St. Paul MN, Sect. IV; Coligan, J.E. et al. (1997) Current Protocols in Immunology, Greene Pub. Associates and Wiley-Interscience, New York NY; and Pound, J.D. (1998) Immunochemical Protocols, Humana Press, Totowa NJ.)

A wide variety of labels and conjugation techniques are known by those skilled in the art and may be used in various nucleic acid and amino acid assays. Means for producing labeled hybridization or PCR probes for detecting sequences related to polynucleotides encoding PKIN include oligolabeling, nick translation, end-labeling, or PCR amplification using a labeled nucleotide. Alternatively, the sequences encoding PKIN, or any fragments thereof, may be cloned into a vector for the production of

an mRNA probe. Such vectors are known in the art, are commercially available, and may be used to synthesize RNA probes in vitro by addition of an appropriate RNA polymerase such as T7, T3, or SP6 and labeled nucleotides. These procedures may be conducted using a variety of commercially available kits, such as those provided by Amersham Pharmacia Biotech, Promega (Madison WI), and US Biochemical. Suitable reporter molecules or labels which may be used for ease of detection include radionuclides, enzymes, fluorescent, chemiluminescent, or chromogenic agents, as well as substrates, cofactors, inhibitors, magnetic particles, and the like.

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Host cells transformed with nucleotide sequences encoding PKIN may be cultured under conditions suitable for the expression and recovery of the protein from cell culture. The protein produced by a transformed cell may be secreted or retained intracellularly depending on the sequence and/or the vector used. As will be understood by those of skill in the art, expression vectors containing polynucleotides which encode PKIN may be designed to contain signal sequences which direct secretion of PKIN through a prokaryotic or eukaryotic cell membrane.

In addition, a host cell strain may be chosen for its ability to modulate expression of the inserted sequences or to process the expressed protein in the desired fashion. Such modifications of the polypeptide include, but are not limited to, acetylation, carboxylation, glycosylation, phosphorylation, lipidation, and acylation. Post-translational processing which cleaves a "prepro" or "pro" form of the protein may also be used to specify protein targeting, folding, and/or activity. Different host cells which have specific cellular machinery and characteristic mechanisms for post-translational activities (e.g., CHO, HeLa, MDCK, HEK293, and WI38) are available from the American Type Culture Collection (ATCC, Manassas VA) and may be chosen to ensure the correct modification and processing of the foreign protein.

In another embodiment of the invention, natural, modified, or recombinant nucleic acid sequences encoding PKIN may be ligated to a heterologous sequence resulting in translation of a fusion protein in any of the aforementioned host systems. For example, a chimeric PKIN protein containing a heterologous moiety that can be recognized by a commercially available antibody may facilitate the screening of peptide libraries for inhibitors of PKIN activity. Heterologous protein and peptide moieties may also facilitate purification of fusion proteins using commercially available affinity matrices. Such moieties include, but are not limited to, glutathione S-transferase (GST), maltose binding protein (MBP), thioredoxin (Trx), calmodulin binding peptide (CBP), 6-His, FLAG, *c-myc*, and hemagglutinin (HA). GST, MBP, Trx, CBP, and 6-His enable purification of their cognate fusion proteins on immobilized glutathione, maltose, phenylarsine oxide, calmodulin, and metal-chelate resins, respectively. FLAG, *c-myc*, and hemagglutinin (HA) enable immunoaffinity purification of fusion proteins using commercially available monoclonal and polyclonal antibodies that specifically recognize

these epitope tags. A fusion protein may also be engineered to contain a proteolytic cleavage site located between the PKIN encoding sequence and the heterologous protein sequence, so that PKIN may be cleaved away from the heterologous moiety following purification. Methods for fusion protein expression and purification are discussed in Ausubel (1995, <u>supra</u>, ch. 10). A variety of commercially available kits may also be used to facilitate expression and purification of fusion proteins.

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In a further embodiment of the invention, synthesis of radiolabeled PKIN may be achieved <u>in vitro</u> using the TNT rabbit reticulocyte lysate or wheat germ extract system (Promega). These systems couple transcription and translation of protein-coding sequences operably associated with the T7, T3, or SP6 promoters. Translation takes place in the presence of a radiolabeled amino acid precursor, for example, <sup>35</sup>S-methionine.

PKIN of the present invention or fragments thereof may be used to screen for compounds that specifically bind to PKIN. At least one and up to a plurality of test compounds may be screened for specific binding to PKIN. Examples of test compounds include antibodies, oligonucleotides, proteins (e.g., receptors), or small molecules.

In one embodiment, the compound thus identified is closely related to the natural ligand of PKIN, e.g., a ligand or fragment thereof, a natural substrate, a structural or functional mimetic, or a natural binding partner. (See, e.g., Coligan, J.E. et al. (1991) <u>Current Protocols in Immunology</u> 1(2): Chapter 5.) Similarly, the compound can be closely related to the natural receptor to which PKIN binds, or to at least a fragment of the receptor, e.g., the ligand binding site. In either case, the compound can be rationally designed using known techniques. In one embodiment, screening for these compounds involves producing appropriate cells which express PKIN, either as a secreted protein or on the cell membrane. Preferred cells include cells from mammals, yeast, <u>Drosophila</u>, or <u>E. coli</u>. Cells expressing PKIN or cell membrane fractions which contain PKIN are then contacted with a test compound and binding, stimulation, or inhibition of activity of either PKIN or the compound is analyzed.

An assay may simply test binding of a test compound to the polypeptide, wherein binding is detected by a fluorophore, radioisotope, enzyme conjugate, or other detectable label. For example, the assay may comprise the steps of combining at least one test compound with PKIN, either in solution or affixed to a solid support, and detecting the binding of PKIN to the compound. Alternatively, the assay may detect or measure binding of a test compound in the presence of a labeled competitor. Additionally, the assay may be carried out using cell-free preparations, chemical libraries, or natural product mixtures, and the test compound(s) may be free in solution or affixed to a solid support.

PKIN of the present invention or fragments thereof may be used to screen for compounds that modulate the activity of PKIN. Such compounds may include agonists, antagonists, or partial or

inverse agonists. In one embodiment, an assay is performed under conditions permissive for PKIN activity, wherein PKIN is combined with at least one test compound, and the activity of PKIN in the presence of a test compound is compared with the activity of PKIN in the absence of the test compound. A change in the activity of PKIN in the presence of the test compound is indicative of a compound that modulates the activity of PKIN. Alternatively, a test compound is combined with an <u>in vitro</u> or cell-free system comprising PKIN under conditions suitable for PKIN activity, and the assay is performed. In either of these assays, a test compound which modulates the activity of PKIN may do so indirectly and need not come in direct contact with the test compound. At least one and up to a plurality of test compounds may be screened.

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In another embodiment, polynucleotides encoding PKIN or their mammalian homologs may be "knocked out" in an animal model system using homologous recombination in embryonic stem (ES) cells. Such techniques are well known in the art and are useful for the generation of animal models of human disease. (See, e.g., U.S. Patent Number 5,175,383 and U.S. Patent Number 5,767,337.) For example, mouse ES cells, such as the mouse 129/SvJ cell line, are derived from the early mouse embryo and grown in culture. The ES cells are transformed with a vector containing the gene of interest disrupted by a marker gene, e.g., the neomycin phosphotransferase gene (neo; Capecchi, M.R. (1989) Science 244:1288-1292). The vector integrates into the corresponding region of the host genome by homologous recombination. Alternatively, homologous recombination takes place using the Cre-loxP system to knockout a gene of interest in a tissue- or developmental stage-specific manner (Marth, J.D. (1996) Clin. Invest. 97:1999-2002; Wagner, K.U. et al. (1997) Nucleic Acids Res. 25:4323-4330). Transformed ES cells are identified and microinjected into mouse cell blastocysts such as those from the C57BL/6 mouse strain. The blastocysts are surgically transferred to pseudopregnant dams, and the resulting chimeric progeny are genotyped and bred to produce heterozygous or homozygous strains. Transgenic animals thus generated may be tested with potential therapeutic or toxic agents.

Polynucleotides encoding PKIN may also be manipulated <u>in vitro</u> in ES cells derived from human blastocysts. Human ES cells have the potential to differentiate into at least eight separate cell lineages including endoderm, mesoderm, and ectodermal cell types. These cell lineages differentiate into, for example, neural cells, hematopoietic lineages, and cardiomyocytes (Thomson, J.A. et al. (1998) Science 282:1145-1147).

Polynucleotides encoding PKIN can also be used to create "knockin" humanized animals (pigs) or transgenic animals (mice or rats) to model human disease. With knockin technology, a region of a polynucleotide encoding PKIN is injected into animal ES cells, and the injected sequence integrates into the animal cell genome. Transformed cells are injected into blastulae, and the blastulae are implanted as described above. Transgenic progeny or inbred lines are studied and treated with potential

pharmaceutical agents to obtain information on treatment of a human disease. Alternatively, a mammal inbred to overexpress PKIN, e.g., by secreting PKIN in its milk, may also serve as a convenient source of that protein (Janne, J. et al. (1998) Biotechnol. Annu. Rev. 4:55-74).

#### **THERAPEUTICS**

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Chemical and structural similarity, e.g., in the context of sequences and motifs, exists between regions of PKIN and human kinases. In addition, the expression of PKIN is closely associated with lipid disorders, pancreatic islet cells, liver disease, leukocytes, umbilical endothelial cells, cancer, as well as, normal and diseased brain, renal, reproductive, bladder tumor, posterior hippocampus, kidney, small intestine, colon, and digestive tissues. Therefore, PKIN appears to play a role in cancer, immune disorders, disorders affecting growth and development, cardiovascular diseases, and lipid disorders. In the treatment of disorders associated with increased PKIN expression or activity, it is desirable to decrease the expression or activity of PKIN. In the treatment of disorders associated with decreased PKIN expression or activity, it is desirable to increase the expression or activity of PKIN.

Therefore, in one embodiment, PKIN or a fragment or derivative thereof may be administered to a subject to treat or prevent a disorder associated with decreased expression or activity of PKIN. Examples of such disorders include, but are not limited to, a cancer, such as adenocarcinoma, leukemia, lymphoma, melanoma, myeloma, sarcoma, teratocarcinoma, and, in particular, cancers of the adrenal gland, bladder, bone, bone marrow, brain, breast, cervix, gall bladder, ganglia, gastrointestinal tract, heart, kidney, liver, lung, muscle, ovary, pancreas, parathyroid, penis, prostate, salivary glands, skin, spleen, testis, thymus, thyroid, and uterus, leukemias such as multiple myeloma and lymphomas such as Hodgkin's disease; an immune disorder, such as acquired immunodeficiency syndrome (AIDS), Addison's disease, adult respiratory distress syndrome, allergies, ankylosing spondylitis, amyloidosis, anemia, asthma, atherosclerosis, autoimmune hemolytic anemia, autoimmune thyroiditis, autoimmune polyendocrinopathy-candidiasis-ectodermal dystrophy (APECED), bronchitis, cholecystitis, contact dermatitis, Crohn's disease, atopic dermatitis, dermatomyositis, diabetes mellitus, emphysema, episodic lymphopenia with lymphocytotoxins, erythroblastosis fetalis, erythema nodosum, atrophic gastritis, glomerulonephritis, Goodpasture's syndrome, gout, Graves' disease, Hashimoto's thyroiditis, hypereosinophilia, irritable bowel syndrome, multiple sclerosis, myasthenia gravis, myocardial or pericardial inflammation, osteoarthritis, osteoporosis, pancreatitis, polymyositis, psoriasis, Reiter's syndrome, rheumatoid arthritis, scleroderma, Sjögren's syndrome, systemic anaphylaxis, systemic lupus erythematosus, systemic sclerosis, thrombocytopenic purpura, ulcerative colitis, uveitis, Werner syndrome, complications of cancer, hemodialysis, and extracorporeal circulation, viral, bacterial, fungal, parasitic, protozoal, and helminthic infections, and trauma; a growth and developmental

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disorder, such as actinic keratosis, arteriosclerosis, atherosclerosis, bursitis, cirrhosis, hepatitis, mixed connective tissue disease (MCTD), myelofibrosis, paroxysmal nocturnal hemoglobinuria, polycythemia vera, psoriasis, primary thrombocythemia, and cancers including adenocarcinoma, leukemia, lymphoma, melanoma, myeloma, sarcoma, teratocarcinoma, and, in particular, cancers of the adrenal gland, bladder, bone, bone marrow, brain, breast, cervix, gall bladder, ganglia, gastrointestinal tract, heart, kidney, liver, lung, muscle, ovary, pancreas, parathyroid, penis, prostate, salivary glands, skin, spleen, testis, thymus, thyroid, and uterus, renal tubular acidosis, anemia, Cushing's syndrome, achondroplastic dwarfism, Duchenne and Becker muscular dystrophy, epilepsy, gonadal dysgenesis, WAGR syndrome (Wilms' tumor, aniridia, genitourinary abnormalities, and mental retardation), Smith-Magenis syndrome, myelodysplastic syndrome, hereditary mucoepithelial dysplasia, hereditary keratodermas, hereditary neuropathies such as Charcot-Marie-Tooth disease and neurofibromatosis, hypothyroidism, hydrocephalus, seizure disorders such as Syndenham's chorea and cerebral palsy, spina bifida, anencephaly, craniorachischisis, congenital glaucoma, cataract, and sensorineural hearing loss; a cardiovascular disease, such as arteriovenous fistula, atherosclerosis, hypertension, vasculitis, Raynaud's disease, aneurysms, arterial dissections, varicose veins, thrombophlebitis and phlebothrombosis, vascular tumors, and complications of thrombolysis, balloon angioplasty, vascular replacement, and coronary artery bypass graft surgery, congestive heart failure, ischemic heart disease, angina pectoris, myocardial infarction, hypertensive heart disease, degenerative valvular heart disease, calcific aortic valve stenosis, congenitally bicuspid aortic valve, mitral annular calcification, mitral valve prolapse, rheumatic fever and rheumatic heart disease, infective endocarditis, nonbacterial thrombotic endocarditis, endocarditis of systemic lupus erythematosus, carcinoid heart disease, cardiomyopathy, myocarditis, pericarditis, neoplastic heart disease, congenital heart disease, and complications of cardiac transplantation, congenital lung anomalies, atelectasis, pulmonary congestion and edema, pulmonary embolism, pulmonary hemorrhage, pulmonary infarction, pulmonary hypertension, vascular sclerosis, obstructive pulmonary disease, restrictive pulmonary disease, chronic obstructive pulmonary disease, emphysema, chronic bronchitis, bronchial asthma, bronchiectasis, bacterial pneumonia, viral and mycoplasmal pneumonia, lung abscess, pulmonary tuberculosis, diffuse interstitial diseases, pneumoconioses, sarcoidosis, idiopathic pulmonary fibrosis, desquamative interstitial pneumonitis, hypersensitivity pneumonitis, pulmonary eosinophilia bronchiolitis obliterans-organizing pneumonia, diffuse pulmonary hemorrhage syndromes, Goodpasture's syndromes, idiopathic pulmonary hemosiderosis, pulmonary involvement in collagen-vascular disorders, pulmonary alveolar proteinosis, lung tumors, inflammatory and noninflammatory pleural effusions, pneumothorax, pleural tumors, drug-induced lung disease, radiation-induced lung disease, and complications of lung transplantation; and a lipid disorder, such as fatty liver, cholestasis, primary biliary cirrhosis, carnitine

deficiency, carnitine palmitoyltransferase deficiency, myoadenylate deaminase deficiency, hypertriglyceridemia, lipid storage disorders such Fabry's disease, Gaucher's disease, Niemann-Pick's disease, metachromatic leukodystrophy, adrenoleukodystrophy, GM<sub>2</sub> gangliosidosis, and ceroid lipofuscinosis, abetalipoproteinemia, Tangier disease, hyperlipoproteinemia, diabetes mellitus,

5 lipodystrophy, lipomatoses, acute panniculitis, disseminated fat necrosis, adiposis dolorosa, lipoid adrenal hyperplasia, minimal change disease, lipomas, atherosclerosis, hypercholesterolemia, hypercholesterolemia, hypercholesterolemia, primary hypoalphalipoproteinemia, hypothyroidism, renal disease, liver disease, lecithin:cholesterol acyltransferase deficiency, cerebrotendinous xanthomatosis, sitosterolemia, hypocholesterolemia, Tay-Sachs disease, Sandhoff's disease, hyperlipidemia, hyperlipemia, lipid myopathies, and obesity.

In another embodiment, a vector capable of expressing PKIN or a fragment or derivative thereof may be administered to a subject to treat or prevent a disorder associated with decreased expression or activity of PKIN including, but not limited to, those described above.

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In a further embodiment, a composition comprising a substantially purified PKIN in conjunction with a suitable pharmaceutical carrier may be administered to a subject to treat or prevent a disorder associated with decreased expression or activity of PKIN including, but not limited to, those provided above.

In still another embodiment, an agonist which modulates the activity of PKIN may be administered to a subject to treat or prevent a disorder associated with decreased expression or activity of PKIN including, but not limited to, those listed above.

In a further embodiment, an antagonist of PKIN may be administered to a subject to treat or prevent a disorder associated with increased expression or activity of PKIN. Examples of such disorders include, but are not limited to, those cancer, immune disorders, disorders affecting growth and development, cardiovascular diseases, and lipid disorders described above. In one aspect, an antibody which specifically binds PKIN may be used directly as an antagonist or indirectly as a targeting or delivery mechanism for bringing a pharmaceutical agent to cells or tissues which express PKIN.

In an additional embodiment, a vector expressing the complement of the polynucleotide encoding PKIN may be administered to a subject to treat or prevent a disorder associated with increased expression or activity of PKIN including, but not limited to, those described above.

In other embodiments, any of the proteins, antagonists, antibodies, agonists, complementary sequences, or vectors of the invention may be administered in combination with other appropriate therapeutic agents. Selection of the appropriate agents for use in combination therapy may be made by one of ordinary skill in the art, according to conventional pharmaceutical principles. The combination of therapeutic agents may act synergistically to effect the treatment or prevention of the various

disorders described above. Using this approach, one may be able to achieve therapeutic efficacy with lower dosages of each agent, thus reducing the potential for adverse side effects.

An antagonist of PKIN may be produced using methods which are generally known in the art. In particular, purified PKIN may be used to produce antibodies or to screen libraries of pharmaceutical agents to identify those which specifically bind PKIN. Antibodies to PKIN may also be generated using methods that are well known in the art. Such antibodies may include, but are not limited to, polyclonal, monoclonal, chimeric, and single chain antibodies, Fab fragments, and fragments produced by a Fab expression library. Neutralizing antibodies (i.e., those which inhibit dimer formation) are generally preferred for therapeutic use.

For the production of antibodies, various hosts including goats, rabbits, rats, mice, humans, and others may be immunized by injection with PKIN or with any fragment or oligopeptide thereof which has immunogenic properties. Depending on the host species, various adjuvants may be used to increase immunological response. Such adjuvants include, but are not limited to, Freund's, mineral gels such as aluminum hydroxide, and surface active substances such as lysolecithin, pluronic polyols, polyanions, peptides, oil emulsions, KLH, and dinitrophenol. Among adjuvants used in humans, BCG (bacilli Calmette-Guerin) and Corynebacterium parvum are especially preferable.

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It is preferred that the oligopeptides, peptides, or fragments used to induce antibodies to PKIN have an amino acid sequence consisting of at least about 5 amino acids, and generally will consist of at least about 10 amino acids. It is also preferable that these oligopeptides, peptides, or fragments are identical to a portion of the amino acid sequence of the natural protein. Short stretches of PKIN amino acids may be fused with those of another protein, such as KLH, and antibodies to the chimeric molecule may be produced.

Monoclonal antibodies to PKIN may be prepared using any technique which provides for the production of antibody molecules by continuous cell lines in culture. These include, but are not limited to, the hybridoma technique, the human B-cell hybridoma technique, and the EBV-hybridoma technique. (See, e.g., Kohler, G. et al. (1975) Nature 256:495-497; Kozbor, D. et al. (1985) J. Immunol. Methods 81:31-42; Cote, R.J. et al. (1983) Proc. Natl. Acad. Sci. USA 80:2026-2030; and Cole, S.P. et al. (1984) Mol. Cell Biol. 62:109-120.)

In addition, techniques developed for the production of "chimeric antibodies," such as the splicing of mouse antibody genes to human antibody genes to obtain a molecule with appropriate antigen specificity and biological activity, can be used. (See, e.g., Morrison, S.L. et al. (1984) Proc. Natl. Acad. Sci. USA 81:6851-6855; Neuberger, M.S. et al. (1984) Nature 312:604-608; and Takeda, S. et al. (1985) Nature 314:452-454.) Alternatively, techniques described for the production of single chain antibodies may be adapted, using methods known in the art, to produce PKIN-specific single

chain antibodies. Antibodies with related specificity, but of distinct idiotypic composition, may be generated by chain shuffling from random combinatorial immunoglobulin libraries. (See, e.g., Burton, D.R. (1991) Proc. Natl. Acad. Sci. USA 88:10134-10137.)

Antibodies may also be produced by inducing <u>in vivo</u> production in the lymphocyte population or by screening immunoglobulin libraries or panels of highly specific binding reagents as disclosed in the literature. (See, e.g., Orlandi, R. et al. (1989) Proc. Natl. Acad. Sci. USA 86:3833-3837; Winter, G. et al. (1991) Nature 349:293-299.)

Antibody fragments which contain specific binding sites for PKIN may also be generated. For example, such fragments include, but are not limited to,  $F(ab)_2$  fragments produced by pepsin digestion of the antibody molecule and Fab fragments generated by reducing the disulfide bridges of the  $F(ab)_2$  fragments. Alternatively, Fab expression libraries may be constructed to allow rapid and easy identification of monoclonal Fab fragments with the desired specificity. (See, e.g., Huse, W.D. et al. (1989) Science 246:1275-1281.)

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Various immunoassays may be used for screening to identify antibodies having the desired specificity. Numerous protocols for competitive binding or immunoradiometric assays using either polyclonal or monoclonal antibodies with established specificities are well known in the art. Such immunoassays typically involve the measurement of complex formation between PKIN and its specific antibody. A two-site, monoclonal-based immunoassay utilizing monoclonal antibodies reactive to two non-interfering PKIN epitopes is generally used, but a competitive binding assay may also be employed (Pound, supra).

Various methods such as Scatchard analysis in conjunction with radioimmunoassay techniques may be used to assess the affinity of antibodies for PKIN. Affinity is expressed as an association constant,  $K_a$ , which is defined as the molar concentration of PKIN-antibody complex divided by the molar concentrations of free antigen and free antibody under equilibrium conditions. The  $K_a$  determined for a preparation of polyclonal antibodies, which are heterogeneous in their affinities for multiple PKIN epitopes, represents the average affinity, or avidity, of the antibodies for PKIN. The  $K_a$  determined for a preparation of monoclonal antibodies, which are monospecific for a particular PKIN epitope, represents a true measure of affinity. High-affinity antibody preparations with  $K_a$  ranging from about  $10^9$  to  $10^{12}$  L/mole are preferred for use in immunoassays in which the PKIN-antibody complex must withstand rigorous manipulations. Low-affinity antibody preparations with  $K_a$  ranging from about  $10^6$  to  $10^7$  L/mole are preferred for use in immunopurification and similar procedures which ultimately require dissociation of PKIN, preferably in active form, from the antibody (Catty, D. (1988) Antibodies, Volume I: A Practical Approach, IRL Press, Washington DC; Liddell, J.E. and A. Cryer (1991) A Practical Guide to Monoclonal Antibodies, John Wiley & Sons, New York NY).

The titer and avidity of polyclonal antibody preparations may be further evaluated to determine the quality and suitability of such preparations for certain downstream applications. For example, a polyclonal antibody preparation containing at least 1-2 mg specific antibody/ml, preferably 5-10 mg specific antibody/ml, is generally employed in procedures requiring precipitation of PKIN-antibody complexes. Procedures for evaluating antibody specificity, titer, and avidity, and guidelines for antibody quality and usage in various applications, are generally available. (See, e.g., Catty, <u>supra</u>, and Coligan et al. <u>supra</u>.)

In another embodiment of the invention, the polynucleotides encoding PKIN, or any fragment or complement thereof, may be used for therapeutic purposes. In one aspect, modifications of gene expression can be achieved by designing complementary sequences or antisense molecules (DNA, RNA, PNA, or modified oligonucleotides) to the coding or regulatory regions of the gene encoding PKIN. Such technology is well known in the art, and antisense oligonucleotides or larger fragments can be designed from various locations along the coding or control regions of sequences encoding PKIN. (See, e.g., Agrawal, S., ed. (1996) Antisense Therapeutics, Humana Press Inc., Totawa NJ.)

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In therapeutic use, any gene delivery system suitable for introduction of the antisense sequences into appropriate target cells can be used. Antisense sequences can be delivered intracellularly in the form of an expression plasmid which, upon transcription, produces a sequence complementary to at least a portion of the cellular sequence encoding the target protein. (See, e.g., Slater, J.E. et al. (1998) J. Allergy Cli. Immunol. 102(3):469-475; and Scanlon, K.J. et al. (1995) 9(13):1288-1296.) Antisense sequences can also be introduced intracellularly through the use of viral vectors, such as retrovirus and adeno-associated virus vectors. (See, e.g., Miller, A.D. (1990) Blood 76:271; Ausubel, supra; Uckert, W. and W. Walther (1994) Pharmacol. Ther. 63(3):323-347.) Other gene delivery mechanisms include liposome-derived systems, artificial viral envelopes, and other systems known in the art. (See, e.g., Rossi, J.J. (1995) Br. Med. Bull. 51(1):217-225; Boado, R.J. et al. (1998) J. Pharm. Sci. 87(11):1308-1315; and Morris, M.C. et al. (1997) Nucleic Acids Res. 25(14):2730-2736.)

In another embodiment of the invention, polynucleotides encoding PKIN may be used for somatic or germline gene therapy. Gene therapy may be performed to (i) correct a genetic deficiency (e.g., in the cases of severe combined immunodeficiency (SCID)-X1 disease characterized by X-linked inheritance (Cavazzana-Calvo, M. et al. (2000) Science 288:669-672), severe combined immunodeficiency syndrome associated with an inherited adenosine deaminase (ADA) deficiency (Blaese, R.M. et al. (1995) Science 270:475-480; Bordignon, C. et al. (1995) Science 270:470-475), cystic fibrosis (Zabner, J. et al. (1993) Cell 75:207-216; Crystal, R.G. et al. (1995) Hum. Gene Therapy 6:643-666; Crystal, R.G. et al. (1995) Hum. Gene Therapy 6:667-703), thalassamias, familial

hypercholesterolemia, and hemophilia resulting from Factor VIII or Factor IX deficiencies (Crystal, R.G. (1995) Science 270:404-410; Verma, I.M. and N. Somia (1997) Nature 389:239-242)), (ii) express a conditionally lethal gene product (e.g., in the case of cancers which result from unregulated cell proliferation), or (iii) express a protein which affords protection against intracellular parasites (e.g., against human retroviruses, such as human immunodeficiency virus (HIV) (Baltimore, D. (1988) Nature 335:395-396; Poeschla, E. et al. (1996) Proc. Natl. Acad. Sci. USA. 93:11395-11399), hepatitis B or C virus (HBV, HCV); fungal parasites, such as Candida albicans and Paracoccidioides brasiliensis; and protozoan parasites such as Plasmodium falciparum and Trypanosoma cruzi). In the case where a genetic deficiency in PKIN expression or regulation causes disease, the expression of PKIN from an appropriate population of transduced cells may alleviate the clinical manifestations caused by the genetic deficiency.

In a further embodiment of the invention, diseases or disorders caused by deficiencies in PKIN are treated by constructing mammalian expression vectors encoding PKIN and introducing these vectors by mechanical means into PKIN-deficient cells. Mechanical transfer technologies for use with cells <u>in vivo</u> or <u>ex vitro</u> include (i) direct DNA microinjection into individual cells, (ii) ballistic gold particle delivery, (iii) liposome-mediated transfection, (iv) receptor-mediated gene transfer, and (v) the use of DNA transposons (Morgan, R.A. and W.F. Anderson (1993) Annu. Rev. Biochem. 62:191-217; Ivics, Z. (1997) Cell 91:501-510; Boulay, J-L. and H. Récipon (1998) Curr. Opin. Biotechnol. 9:445-450).

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Expression vectors that may be effective for the expression of PKIN include, but are not limited to, the PCDNA 3.1, EPITAG, PRCCMV2, PREP, PVAX vectors (Invitrogen, Carlsbad CA), PCMV-SCRIPT, PCMV-TAG, PEGSH/PERV (Stratagene, La Jolla CA), and PTET-OFF, PTET-ON, PTRE2, PTRE2-LUC, PTK-HYG (Clontech, Palo Alto CA). PKIN may be expressed using (i) a constitutively active promoter, (e.g., from cytomegalovirus (CMV), Rous sarcoma virus (RSV), SV40 virus, thymidine kinase (TK), or β-actin genes), (ii) an inducible promoter (e.g., the tetracycline-regulated promoter (Gossen, M. and H. Bujard (1992) Proc. Natl. Acad. Sci. USA 89:5547-5551; Gossen, M. et al. (1995) Science 268:1766-1769; Rossi, F.M.V. and H.M. Blau (1998) Curr. Opin. Biotechnol. 9:451-456), commercially available in the T-REX plasmid (Invitrogen)); the ecdysone-inducible promoter (available in the plasmids PVGRXR and PIND; Invitrogen); the FK506/rapamycin inducible promoter; or the RU486/mifepristone inducible promoter (Rossi, F.M.V. and Blau, H.M. supra)), or (iii) a tissue-specific promoter or the native promoter of the endogenous gene encoding PKIN from a normal individual.

Commercially available liposome transformation kits (e.g., the PERFECT LIPID TRANSFECTION KIT, available from Invitrogen) allow one with ordinary skill in the art to deliver polynucleotides to target cells in culture and require minimal effort to optimize experimental

parameters. In the alternative, transformation is performed using the calcium phosphate method (Graham, F.L. and A.J. Eb (1973) Virology 52:456-467), or by electroporation (Neumann, E. et al. (1982) EMBO J. 1:841-845). The introduction of DNA to primary cells requires modification of these standardized mammalian transfection protocols.

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In another embodiment of the invention, diseases or disorders caused by genetic defects with respect to PKIN expression are treated by constructing a retrovirus vector consisting of (i) the polynucleotide encoding PKIN under the control of an independent promoter or the retrovirus long terminal repeat (LTR) promoter, (ii) appropriate RNA packaging signals, and (iii) a Rev-responsive element (RRE) along with additional retrovirus cis-acting RNA sequences and coding sequences required for efficient vector propagation. Retrovirus vectors (e.g., PFB and PFBNEO) are commercially available (Stratagene) and are based on published data (Riviere, I. et al. (1995) Proc. Natl. Acad. Sci. USA 92:6733-6737), incorporated by reference herein. The vector is propagated in an appropriate vector producing cell line (VPCL) that expresses an envelope gene with a tropism for receptors on the target cells or a promiscuous envelope protein such as VSVg (Armentano, D. et al. (1987) J. Virol. 61:1647-1650; Bender, M.A. et al. (1987) J. Virol. 61:1639-1646; Adam, M.A. and A.D. Miller (1988) J. Virol. 62:3802-3806; Dull, T. et al. (1998) J. Virol. 72:8463-8471; Zufferey, R. et al. (1998) J. Virol. 72:9873-9880). U.S. Patent Number 5,910,434 to Rigg ("Method for obtaining retrovirus packaging cell lines producing high transducing efficiency retroviral supernatant") discloses a method for obtaining retrovirus packaging cell lines and is hereby incorporated by reference. Propagation of retrovirus vectors, transduction of a population of cells (e.g., CD4<sup>+</sup> T-cells), and the return of transduced cells to a patient are procedures well known to persons skilled in the art of gene therapy and have been well documented (Ranga, U. et al. (1997) J. Virol. 71:7020-7029; Bauer, G. et al. (1997) Blood 89:2259-2267; Bonyhadi, M.L. (1997) J. Virol. 71:4707-4716; Ranga, U. et al.

In the alternative, an adenovirus-based gene therapy delivery system is used to deliver polynucleotides encoding PKIN to cells which have one or more genetic abnormalities with respect to the expression of PKIN. The construction and packaging of adenovirus-based vectors are well known to those with ordinary skill in the art. Replication defective adenovirus vectors have proven to be versatile for importing genes encoding immunoregulatory proteins into intact islets in the pancreas (Csete, M.E. et al. (1995) Transplantation 27:263-268). Potentially useful adenoviral vectors are described in U.S. Patent Number 5,707,618 to Armentano ("Adenovirus vectors for gene therapy"), hereby incorporated by reference. For adenoviral vectors, see also Antinozzi, P.A. et al. (1999) Annu. Rev. Nutr. 19:511-544 and Verma, I.M. and N. Somia (1997) Nature 18:389:239-242, both incorporated by reference herein.

(1998) Proc. Natl. Acad. Sci. USA 95:1201-1206; Su, L. (1997) Blood 89:2283-2290).

In another alternative, a herpes-based, gene therapy delivery system is used to deliver polynucleotides encoding PKIN to target cells which have one or more genetic abnormalities with respect to the expression of PKIN. The use of herpes simplex virus (HSV)-based vectors may be especially valuable for introducing PKIN to cells of the central nervous system, for which HSV has a tropism. The construction and packaging of herpes-based vectors are well known to those with ordinary skill in the art. A replication-competent herpes simplex virus (HSV) type 1-based vector has been used to deliver a reporter gene to the eyes of primates (Liu, X. et al. (1999) Exp. Eye Res. 169:385-395). The construction of a HSV-1 virus vector has also been disclosed in detail in U.S. Patent Number 5,804,413 to DeLuca ("Herpes simplex virus strains for gene transfer"), which is hereby incorporated by reference. U.S. Patent Number 5,804,413 teaches the use of recombinant HSV d92 which consists of a genome containing at least one exogenous gene to be transferred to a cell under the control of the appropriate promoter for purposes including human gene therapy. Also taught by this patent are the construction and use of recombinant HSV strains deleted for ICP4, ICP27 and ICP22. For HSV vectors, see also Goins, W.F. et al. (1999) J. Virol. 73:519-532 and Xu, H. et al. (1994) Dev. Biol. 163:152-161, hereby incorporated by reference. The manipulation of cloned herpesvirus sequences, the generation of recombinant virus following the transfection of multiple plasmids containing different segments of the large herpesvirus genomes, the growth and propagation of herpesvirus, and the infection of cells with herpesvirus are techniques well known to those of ordinary skill in the art.

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In another alternative, an alphavirus (positive, single-stranded RNA virus) vector is used to deliver polynucleotides encoding PKIN to target cells. The biology of the prototypic alphavirus, Semliki Forest Virus (SFV), has been studied extensively and gene transfer vectors have been based on the SFV genome (Garoff, H. and K.-J. Li (1998) Curr. Opin. Biotechnol. 9:464-469). During alphavirus RNA replication, a subgenomic RNA is generated that normally encodes the viral capsid proteins. This subgenomic RNA replicates to higher levels than the full length genomic RNA, resulting in the overproduction of capsid proteins relative to the viral proteins with enzymatic activity (e.g., protease and polymerase). Similarly, inserting the coding sequence for PKIN into the alphavirus genome in place of the capsid-coding region results in the production of a large number of PKIN-coding RNAs and the synthesis of high levels of PKIN in vector transduced cells. While alphavirus infection is typically associated with cell lysis within a few days, the ability to establish a persistent infection in hamster normal kidney cells (BHK-21) with a variant of Sindbis virus (SIN) indicates that the lytic replication of alphaviruses can be altered to suit the needs of the gene therapy application (Dryga, S.A. et al. (1997) Virology 228:74-83). The wide host range of alphaviruses will allow the introduction of PKIN into a variety of cell types. The specific transduction of a subset of cells in a population may

require the sorting of cells prior to transduction. The methods of manipulating infectious cDNA clones of alphaviruses, performing alphavirus cDNA and RNA transfections, and performing alphavirus infections, are well known to those with ordinary skill in the art.

Oligonucleotides derived from the transcription initiation site, e.g., between about positions -10 and +10 from the start site, may also be employed to inhibit gene expression. Similarly, inhibition can be achieved using triple helix base-pairing methodology. Triple helix pairing is useful because it causes inhibition of the ability of the double helix to open sufficiently for the binding of polymerases, transcription factors, or regulatory molecules. Recent therapeutic advances using triplex DNA have been described in the literature. (See, e.g., Gee, J.E. et al. (1994) in Huber, B.E. and B.I. Carr, Molecular and Immunologic Approaches, Futura Publishing, Mt. Kisco NY, pp. 163-177.) A complementary sequence or antisense molecule may also be designed to block translation of mRNA by preventing the transcript from binding to ribosomes.

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Ribozymes, enzymatic RNA molecules, may also be used to catalyze the specific cleavage of RNA. The mechanism of ribozyme action involves sequence-specific hybridization of the ribozyme molecule to complementary target RNA, followed by endonucleolytic cleavage. For example, engineered hammerhead motif ribozyme molecules may specifically and efficiently catalyze endonucleolytic cleavage of sequences encoding PKIN.

Specific ribozyme cleavage sites within any potential RNA target are initially identified by scanning the target molecule for ribozyme cleavage sites, including the following sequences: GUA, GUU, and GUC. Once identified, short RNA sequences of between 15 and 20 ribonucleotides, corresponding to the region of the target gene containing the cleavage site, may be evaluated for secondary structural features which may render the oligonucleotide inoperable. The suitability of candidate targets may also be evaluated by testing accessibility to hybridization with complementary oligonucleotides using ribonuclease protection assays.

Complementary ribonucleic acid molecules and ribozymes of the invention may be prepared by any method known in the art for the synthesis of nucleic acid molecules. These include techniques for chemically synthesizing oligonucleotides such as solid phase phosphoramidite chemical synthesis.

Alternatively, RNA molecules may be generated by <u>in vitro</u> and <u>in vivo</u> transcription of DNA sequences encoding PKIN. Such DNA sequences may be incorporated into a wide variety of vectors with suitable RNA polymerase promoters such as T7 or SP6. Alternatively, these cDNA constructs that synthesize complementary RNA, constitutively or inducibly, can be introduced into cell lines, cells, or tissues.

RNA molecules may be modified to increase intracellular stability and half-life. Possible modifications include, but are not limited to, the addition of flanking sequences at the 5' and/or 3' ends of the molecule, or the use of phosphorothioate or 2' O-methyl rather than phosphodiesterase linkages

within the backbone of the molecule. This concept is inherent in the production of PNAs and can be extended in all of these molecules by the inclusion of nontraditional bases such as inosine, queosine, and wybutosine, as well as acetyl-, methyl-, thio-, and similarly modified forms of adenine, cytidine, guanine, thymine, and uridine which are not as easily recognized by endogenous endonucleases.

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An additional embodiment of the invention encompasses a method for screening for a compound which is effective in altering expression of a polynucleotide encoding PKIN. Compounds which may be effective in altering expression of a specific polynucleotide may include, but are not limited to, oligonucleotides, antisense oligonucleotides, triple helix-forming oligonucleotides, transcription factors and other polypeptide transcriptional regulators, and non-macromolecular chemical entities which are capable of interacting with specific polynucleotide sequences. Effective compounds may alter polynucleotide expression by acting as either inhibitors or promoters of polynucleotide expression. Thus, in the treatment of disorders associated with increased PKIN expression or activity, a compound which specifically inhibits expression of the polynucleotide encoding PKIN may be therapeutically useful, and in the treatment of disorders associated with decreased PKIN expression or activity, a compound which specifically promotes expression of the polynucleotide encoding PKIN may be therapeutically useful.

At least one, and up to a plurality, of test compounds may be screened for effectiveness in altering expression of a specific polynucleotide. A test compound may be obtained by any method commonly known in the art, including chemical modification of a compound known to be effective in altering polynucleotide expression; selection from an existing, commercially-available or proprietary library of naturally-occurring or non-natural chemical compounds; rational design of a compound based on chemical and/or structural properties of the target polynucleotide; and selection from a library of chemical compounds created combinatorially or randomly. A sample comprising a polynucleotide encoding PKIN is exposed to at least one test compound thus obtained. The sample may comprise, for example, an intact or permeabilized cell, or an in vitro cell-free or reconstituted biochemical system. Alterations in the expression of a polynucleotide encoding PKIN are assayed by any method commonly known in the art. Typically, the expression of a specific nucleotide is detected by hybridization with a probe having a nucleotide sequence complementary to the sequence of the polynucleotide encoding PKIN. The amount of hybridization may be quantified, thus forming the basis for a comparison of the expression of the polynucleotide both with and without exposure to one or more test compounds. Detection of a change in the expression of a polynucleotide exposed to a test compound indicates that the test compound is effective in altering the expression of the polynucleotide. A screen for a compound effective in altering expression of a specific polynucleotide can be carried out, for example, using a Schizosaccharomyces pombe gene expression system (Atkins, D. et al. (1999) U.S. Patent No. 5,932,435; Arndt, G.M. et al. (2000) Nucleic Acids Res. 28:E15) or a

human cell line such as HeLa cell (Clarke, M.L. et al. (2000) Biochem. Biophys. Res. Commun. 268:8-13). A particular embodiment of the present invention involves screening a combinatorial library of oligonucleotides (such as deoxyribonucleotides, ribonucleotides, peptide nucleic acids, and modified oligonucleotides) for antisense activity against a specific polynucleotide sequence (Bruice, T.W. et al. (1997) U.S. Patent No. 5,686,242; Bruice, T.W. et al. (2000) U.S. Patent No. 6,022,691).

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Many methods for introducing vectors into cells or tissues are available and equally suitable for use <u>in vivo</u>, <u>in vitro</u>, and <u>ex vivo</u>. For <u>ex vivo</u> therapy, vectors may be introduced into stem cells taken from the patient and clonally propagated for autologous transplant back into that same patient. Delivery by transfection, by liposome injections, or by polycationic amino polymers may be achieved using methods which are well known in the art. (See, e.g., Goldman, C.K. et al. (1997) Nat. Biotechnol. 15:462-466.)

Any of the therapeutic methods described above may be applied to any subject in need of such therapy, including, for example, mammals such as humans, dogs, cats, cows, horses, rabbits, and monkeys.

An additional embodiment of the invention relates to the administration of a composition which generally comprises an active ingredient formulated with a pharmaceutically acceptable excipient. Excipients may include, for example, sugars, starches, celluloses, gums, and proteins. Various formulations are commonly known and are thoroughly discussed in the latest edition of Remington's Pharmaceutical Sciences (Maack Publishing, Easton PA). Such compositions may consist of PKIN, antibodies to PKIN, and mimetics, agonists, antagonists, or inhibitors of PKIN.

The compositions utilized in this invention may be administered by any number of routes including, but not limited to, oral, intravenous, intramuscular, intra-arterial, intramedullary, intrathecal, intraventricular, pulmonary, transdermal, subcutaneous, intraperitoneal, intranasal, enteral, topical, sublingual, or rectal means.

Compositions for pulmonary administration may be prepared in liquid or dry powder form. These compositions are generally aerosolized immediately prior to inhalation by the patient. In the case of small molecules (e.g. traditional low molecular weight organic drugs), aerosol delivery of fast-acting formulations is well-known in the art. In the case of macromolecules (e.g. larger peptides and proteins), recent developments in the field of pulmonary delivery via the alveolar region of the lung have enabled the practical delivery of drugs such as insulin to blood circulation (see, e.g., Patton, J.S. et al., U.S. Patent No. 5,997,848). Pulmonary delivery has the advantage of administration without needle injection, and obviates the need for potentially toxic penetration enhancers.

Compositions suitable for use in the invention include compositions wherein the active ingredients are contained in an effective amount to achieve the intended purpose. The determination of an effective dose is well within the capability of those skilled in the art.

Specialized forms of compositions may be prepared for direct intracellular delivery of macromolecules comprising PKIN or fragments thereof. For example, liposome preparations containing a cell-impermeable macromolecule may promote cell fusion and intracellular delivery of the macromolecule. Alternatively, PKIN or a fragment thereof may be joined to a short cationic N-terminal portion from the HIV Tat-1 protein. Fusion proteins thus generated have been found to transduce into the cells of all tissues, including the brain, in a mouse model system (Schwarze, S.R. et al. (1999) Science 285:1569-1572).

For any compound, the therapeutically effective dose can be estimated initially either in cell culture assays, e.g., of neoplastic cells, or in animal models such as mice, rats, rabbits, dogs, monkeys, or pigs. An animal model may also be used to determine the appropriate concentration range and route of administration. Such information can then be used to determine useful doses and routes for administration in humans.

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A therapeutically effective dose refers to that amount of active ingredient, for example PKIN or fragments thereof, antibodies of PKIN, and agonists, antagonists or inhibitors of PKIN, which ameliorates the symptoms or condition. Therapeutic efficacy and toxicity may be determined by standard pharmaceutical procedures in cell cultures or with experimental animals, such as by calculating the  $ED_{50}$  (the dose therapeutically effective in 50% of the population) or  $LD_{50}$  (the dose lethal to 50% of the population) statistics. The dose ratio of toxic to therapeutic effects is the therapeutic index, which can be expressed as the  $LD_{50}/ED_{50}$  ratio. Compositions which exhibit large therapeutic indices are preferred. The data obtained from cell culture assays and animal studies are used to formulate a range of dosage for human use. The dosage contained in such compositions is preferably within a range of circulating concentrations that includes the  $ED_{50}$  with little or no toxicity. The dosage varies within this range depending upon the dosage form employed, the sensitivity of the patient, and the route of administration.

The exact dosage will be determined by the practitioner, in light of factors related to the subject requiring treatment. Dosage and administration are adjusted to provide sufficient levels of the active moiety or to maintain the desired effect. Factors which may be taken into account include the severity of the disease state, the general health of the subject, the age, weight, and gender of the subject, time and frequency of administration, drug combination(s), reaction sensitivities, and response to therapy. Long-acting compositions may be administered every 3 to 4 days, every week, or biweekly depending on the half-life and clearance rate of the particular formulation.

Normal dosage amounts may vary from about  $0.1~\mu g$  to  $100,000~\mu g$ , up to a total dose of about 1 gram, depending upon the route of administration. Guidance as to particular dosages and methods of delivery is provided in the literature and generally available to practitioners in the art. Those skilled in the art will employ different formulations for nucleotides than for proteins or their inhibitors. Similarly, delivery of polynucleotides or polypeptides will be specific to particular cells, conditions, locations, etc.

### DIAGNOSTICS

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In another embodiment, antibodies which specifically bind PKIN may be used for the diagnosis of disorders characterized by expression of PKIN, or in assays to monitor patients being treated with PKIN or agonists, antagonists, or inhibitors of PKIN. Antibodies useful for diagnostic purposes may be prepared in the same manner as described above for therapeutics. Diagnostic assays for PKIN include methods which utilize the antibody and a label to detect PKIN in human body fluids or in extracts of cells or tissues. The antibodies may be used with or without modification, and may be labeled by covalent or non-covalent attachment of a reporter molecule. A wide variety of reporter molecules, several of which are described above, are known in the art and may be used.

A variety of protocols for measuring PKIN, including ELISAs, RIAs, and FACS, are known in the art and provide a basis for diagnosing altered or abnormal levels of PKIN expression. Normal or standard values for PKIN expression are established by combining body fluids or cell extracts taken from normal mammalian subjects, for example, human subjects, with antibodies to PKIN under conditions suitable for complex formation. The amount of standard complex formation may be quantitated by various methods, such as photometric means. Quantities of PKIN expressed in subject, control, and disease samples from biopsied tissues are compared with the standard values. Deviation between standard and subject values establishes the parameters for diagnosing disease.

In another embodiment of the invention, the polynucleotides encoding PKIN may be used for diagnostic purposes. The polynucleotides which may be used include oligonucleotide sequences, complementary RNA and DNA molecules, and PNAs. The polynucleotides may be used to detect and quantify gene expression in biopsied tissues in which expression of PKIN may be correlated with disease. The diagnostic assay may be used to determine absence, presence, and excess expression of PKIN, and to monitor regulation of PKIN levels during therapeutic intervention.

In one aspect, hybridization with PCR probes which are capable of detecting polynucleotide sequences, including genomic sequences, encoding PKIN or closely related molecules may be used to identify nucleic acid sequences which encode PKIN. The specificity of the probe, whether it is made from a highly specific region, e.g., the 5' regulatory region, or from a less specific region, e.g., a conserved motif, and the stringency of the hybridization or amplification will determine whether the

probe identifies only naturally occurring sequences encoding PKIN, allelic variants, or related sequences.

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Probes may also be used for the detection of related sequences, and may have at least 50% sequence identity to any of the PKIN encoding sequences. The hybridization probes of the subject invention may be DNA or RNA and may be derived from the sequence of SEQ ID NO:27-52 or from genomic sequences including promoters, enhancers, and introns of the PKIN gene.

Means for producing specific hybridization probes for DNAs encoding PKIN include the cloning of polynucleotide sequences encoding PKIN or PKIN derivatives into vectors for the production of mRNA probes. Such vectors are known in the art, are commercially available, and may be used to synthesize RNA probes <u>in vitro</u> by means of the addition of the appropriate RNA polymerases and the appropriate labeled nucleotides. Hybridization probes may be labeled by a variety of reporter groups, for example, by radionuclides such as <sup>32</sup>P or <sup>35</sup>S, or by enzymatic labels, such as alkaline phosphatase coupled to the probe via avidin/biotin coupling systems, and the like.

Polynucleotide sequences encoding PKIN may be used for the diagnosis of disorders associated with expression of PKIN. Examples of such disorders include, but are not limited to, a cancer, such as adenocarcinoma, leukemia, lymphoma, melanoma, myeloma, sarcoma, teratocarcinoma, and, in particular, cancers of the adrenal gland, bladder, bone, bone marrow, brain, breast, cervix, gall bladder, ganglia, gastrointestinal tract, heart, kidney, liver, lung, muscle, ovary, pancreas, parathyroid, penis, prostate, salivary glands, skin, spleen, testis, thymus, thyroid, and uterus, leukemias such as multiple myeloma and lymphomas such as Hodgkin's disease; an immune disorder, such as acquired immunodeficiency syndrome (AIDS), Addison's disease, adult respiratory distress syndrome, allergies, ankylosing spondylitis, amyloidosis, anemia, asthma, atherosclerosis, autoimmune hemolytic anemia, autoimmune thyroiditis, autoimmune polyendocrinopathy-candidiasis-ectodermal dystrophy (APECED), bronchitis, cholecystitis, contact dermatitis, Crohn's disease, atopic dermatitis, dermatomyositis, diabetes mellitus, emphysema, episodic lymphopenia with lymphocytotoxins, erythroblastosis fetalis, erythema nodosum, atrophic gastritis, glomerulonephritis, Goodpasture's syndrome, gout, Graves' disease, Hashimoto's thyroiditis, hypereosinophilia, irritable bowel syndrome, multiple sclerosis, myasthenia gravis, myocardial or pericardial inflammation, osteoarthritis, osteoporosis, pancreatitis, polymyositis, psoriasis, Reiter's syndrome, rheumatoid arthritis, scleroderma, Sjögren's syndrome, systemic anaphylaxis, systemic lupus erythematosus, systemic sclerosis, thrombocytopenic purpura, ulcerative colitis, uveitis, Werner syndrome, complications of cancer, hemodialysis, and extracorporeal circulation, viral, bacterial, fungal, parasitic, protozoal, and helminthic infections, and trauma; a growth and developmental disorder, such as actinic keratosis, arteriosclerosis, atherosclerosis, bursitis, cirrhosis, hepatitis, mixed connective tissue disease

(MCTD), myelofibrosis, paroxysmal nocturnal hemoglobinuria, polycythemia vera, psoriasis, primary thrombocythemia, and cancers including adenocarcinoma, leukemia, lymphoma, melanoma, myeloma, sarcoma, teratocarcinoma, and, in particular, cancers of the adrenal gland, bladder, bone, bone marrow, brain, breast, cervix, gall bladder, ganglia, gastrointestinal tract, heart, kidney, liver, lung, muscle, ovary, pancreas, parathyroid, penis, prostate, salivary glands, skin, spleen, testis, thymus, thyroid, and uterus, renal tubular acidosis, anemia, Cushing's syndrome, achondroplastic dwarfism, Duchenne and Becker muscular dystrophy, epilepsy, gonadal dysgenesis, WAGR syndrome (Wilms' tumor, aniridia, genitourinary abnormalities, and mental retardation), Smith-Magenis syndrome, myelodysplastic syndrome, hereditary mucoepithelial dysplasia, hereditary keratodermas, hereditary neuropathies such as Charcot-Marie-Tooth disease and neurofibromatosis, hypothyroidism, hydrocephalus, seizure disorders such as Syndenham's chorea and cerebral palsy, spina bifida, anencephaly, craniorachischisis, congenital glaucoma, cataract, and sensorineural hearing loss; a cardiovascular disease, such as arteriovenous fistula, atherosclerosis, hypertension, vasculitis, Raynaud's disease, aneurysms, arterial dissections, varicose veins, thrombophlebitis and phlebothrombosis, vascular tumors, and complications of thrombolysis, balloon angioplasty, vascular replacement, and coronary artery bypass graft surgery, congestive heart failure, ischemic heart disease, angina pectoris, myocardial infarction, hypertensive heart disease, degenerative valvular heart disease, calcific aortic valve stenosis, congenitally bicuspid aortic valve, mitral annular calcification, mitral valve prolapse, rheumatic fever and rheumatic heart disease, infective endocarditis, nonbacterial thrombotic endocarditis, endocarditis of systemic lupus erythematosus, carcinoid heart disease, cardiomyopathy, myocarditis, pericarditis, neoplastic heart disease, congenital heart disease, and complications of cardiac transplantation, congenital lung anomalies, atelectasis, pulmonary congestion and edema, pulmonary embolism, pulmonary hemorrhage, pulmonary infarction, pulmonary hypertension, vascular sclerosis, obstructive pulmonary disease, restrictive pulmonary disease, chronic obstructive pulmonary disease, emphysema, chronic bronchitis, bronchial asthma, bronchiectasis, bacterial pneumonia, viral and mycoplasmal pneumonia, lung abscess, pulmonary tuberculosis, diffuse interstitial diseases, pneumoconioses, sarcoidosis, idiopathic pulmonary fibrosis, desquamative interstitial pneumonitis, hypersensitivity pneumonitis, pulmonary eosinophilia bronchiolitis obliterans-organizing pneumonia, diffuse pulmonary hemorrhage syndromes, Goodpasture's syndromes, idiopathic pulmonary hemosiderosis, pulmonary involvement in collagen-vascular disorders, pulmonary alveolar proteinosis, lung tumors, inflammatory and noninflammatory pleural effusions, pneumothorax, pleural tumors, drug-induced lung disease, radiation-induced lung disease, and complications of lung transplantation; and a lipid disorder, such as fatty liver, cholestasis, primary biliary cirrhosis, carnitine deficiency, carnitine palmitoyltransferase deficiency, myoadenylate deaminase deficiency,

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hypertriglyceridemia, lipid storage disorders such Fabry's disease, Gaucher's disease, Niemann-Pick's disease, metachromatic leukodystrophy, adrenoleukodystrophy, GM2 gangliosidosis, and ceroid lipofuscinosis, abetalipoproteinemia, Tangier disease, hyperlipoproteinemia, diabetes mellitus, lipodystrophy, lipomatoses, acute panniculitis, disseminated fat necrosis, adiposis dolorosa, lipoid adrenal hyperplasia, minimal change disease, lipomas, atherosclerosis, hypercholesterolemia, hypercholesterolemia with hypertriglyceridemia, primary hypoalphalipoproteinemia, hypothyroidism, renal disease, liver disease, lecithin:cholesterol acyltransferase deficiency, cerebrotendinous xanthomatosis, sitosterolemia, hypocholesterolemia, Tay-Sachs disease, Sandhoff's disease, hyperlipidemia, hyperlipemia, lipid myopathies, and obesity. The polynucleotide sequences encoding PKIN may be used in Southern or northern analysis, dot blot, or other membrane-based technologies; in PCR technologies; in dipstick, pin, and multiformat ELISA-like assays; and in microarrays utilizing fluids or tissues from patients to detect altered PKIN expression. Such qualitative or quantitative methods are well known in the art.

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In a particular aspect, the nucleotide sequences encoding PKIN may be useful in assays that detect the presence of associated disorders, particularly those mentioned above. The nucleotide sequences encoding PKIN may be labeled by standard methods and added to a fluid or tissue sample from a patient under conditions suitable for the formation of hybridization complexes. After a suitable incubation period, the sample is washed and the signal is quantified and compared with a standard value. If the amount of signal in the patient sample is significantly altered in comparison to a control sample then the presence of altered levels of nucleotide sequences encoding PKIN in the sample indicates the presence of the associated disorder. Such assays may also be used to evaluate the efficacy of a particular therapeutic treatment regimen in animal studies, in clinical trials, or to monitor the treatment of an individual patient.

In order to provide a basis for the diagnosis of a disorder associated with expression of PKIN, a normal or standard profile for expression is established. This may be accomplished by combining body fluids or cell extracts taken from normal subjects, either animal or human, with a sequence, or a fragment thereof, encoding PKIN, under conditions suitable for hybridization or amplification.

Standard hybridization may be quantified by comparing the values obtained from normal subjects with values from an experiment in which a known amount of a substantially purified polynucleotide is used. Standard values obtained in this manner may be compared with values obtained from samples from patients who are symptomatic for a disorder. Deviation from standard values is used to establish the presence of a disorder.

Once the presence of a disorder is established and a treatment protocol is initiated, hybridization assays may be repeated on a regular basis to determine if the level of expression in the

patient begins to approximate that which is observed in the normal subject. The results obtained from successive assays may be used to show the efficacy of treatment over a period ranging from several days to months.

With respect to cancer, the presence of an abnormal amount of transcript (either under- or overexpressed) in biopsied tissue from an individual may indicate a predisposition for the development of the disease, or may provide a means for detecting the disease prior to the appearance of actual clinical symptoms. A more definitive diagnosis of this type may allow health professionals to employ preventative measures or aggressive treatment earlier thereby preventing the development or further progression of the cancer.

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Additional diagnostic uses for oligonucleotides designed from the sequences encoding PKIN may involve the use of PCR. These oligomers may be chemically synthesized, generated enzymatically, or produced <u>in vitro</u>. Oligomers will preferably contain a fragment of a polynucleotide encoding PKIN, or a fragment of a polynucleotide complementary to the polynucleotide encoding PKIN, and will be employed under optimized conditions for identification of a specific gene or condition. Oligomers may also be employed under less stringent conditions for detection or quantification of closely related DNA or RNA sequences.

In a particular aspect, oligonucleotide primers derived from the polynucleotide sequences encoding PKIN may be used to detect single nucleotide polymorphisms (SNPs). SNPs are substitutions, insertions and deletions that are a frequent cause of inherited or acquired genetic disease in humans. Methods of SNP detection include, but are not limited to, single-stranded conformation polymorphism (SSCP) and fluorescent SSCP (fSSCP) methods. In SSCP, oligonucleotide primers derived from the polynucleotide sequences encoding PKIN are used to amplify DNA using the polymerase chain reaction (PCR). The DNA may be derived, for example, from diseased or normal tissue, biopsy samples, bodily fluids, and the like. SNPs in the DNA cause differences in the secondary and tertiary structures of PCR products in single-stranded form, and these differences are detectable using gel electrophoresis in non-denaturing gels. In fSCCP, the oligonucleotide primers are fluorescently labeled, which allows detection of the amplimers in high-throughput equipment such as DNA sequencing machines. Additionally, sequence database analysis methods, termed in silico SNP (isSNP), are capable of identifying polymorphisms by comparing the sequence of individual overlapping DNA fragments which assemble into a common consensus sequence. These computerbased methods filter out sequence variations due to laboratory preparation of DNA and sequencing errors using statistical models and automated analyses of DNA sequence chromatograms. In the alternative, SNPs may be detected and characterized by mass spectrometry using, for example, the high throughput MASSARRAY system (Sequenom, Inc., San Diego CA).

Methods which may also be used to quantify the expression of PKIN include radiolabeling or biotinylating nucleotides, coamplification of a control nucleic acid, and interpolating results from standard curves. (See, e.g., Melby, P.C. et al. (1993) J. Immunol. Methods 159:235-244; Duplaa, C. et al. (1993) Anal. Biochem. 212:229-236.) The speed of quantitation of multiple samples may be accelerated by running the assay in a high-throughput format where the oligomer or polynucleotide of interest is presented in various dilutions and a spectrophotometric or colorimetric response gives rapid quantitation.

In further embodiments, oligonucleotides or longer fragments derived from any of the polynucleotide sequences described herein may be used as elements on a microarray. The microarray can be used in transcript imaging techniques which monitor the relative expression levels of large numbers of genes simultaneously as described below. The microarray may also be used to identify genetic variants, mutations, and polymorphisms. This information may be used to determine gene function, to understand the genetic basis of a disorder, to diagnose a disorder, to monitor progression/regression of disease as a function of gene expression, and to develop and monitor the activities of therapeutic agents in the treatment of disease. In particular, this information may be used to develop a pharmacogenomic profile of a patient in order to select the most appropriate and effective treatment regimen for that patient. For example, therapeutic agents which are highly effective and display the fewest side effects may be selected for a patient based on his/her pharmacogenomic profile.

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In another embodiment, PKIN, fragments of PKIN, or antibodies specific for PKIN may be used as elements on a microarray. The microarray may be used to monitor or measure protein-protein interactions, drug-target interactions, and gene expression profiles, as described above.

A particular embodiment relates to the use of the polynucleotides of the present invention to generate a transcript image of a tissue or cell type. A transcript image represents the global pattern of gene expression by a particular tissue or cell type. Global gene expression patterns are analyzed by quantifying the number of expressed genes and their relative abundance under given conditions and at a given time. (See Seilhamer et al., "Comparative Gene Transcript Analysis," U.S. Patent Number 5,840,484, expressly incorporated by reference herein.) Thus a transcript image may be generated by hybridizing the polynucleotides of the present invention or their complements to the totality of transcripts or reverse transcripts of a particular tissue or cell type. In one embodiment, the hybridization takes place in high-throughput format, wherein the polynucleotides of the present invention or their complements comprise a subset of a plurality of elements on a microarray. The resultant transcript image would provide a profile of gene activity.

Transcript images may be generated using transcripts isolated from tissues, cell lines, biopsies, or other biological samples. The transcript image may thus reflect gene expression <u>in vivo</u>, as in the case of a tissue or biopsy sample, or <u>in vitro</u>, as in the case of a cell line.

Transcript images which profile the expression of the polynucleotides of the present invention may also be used in conjunction with in vitro model systems and preclinical evaluation of pharmaceuticals, as well as toxicological testing of industrial and naturally-occurring environmental compounds. All compounds induce characteristic gene expression patterns, frequently termed molecular fingerprints or toxicant signatures, which are indicative of mechanisms of action and toxicity (Nuwaysir, E.F. et al. (1999) Mol. Carcinog. 24:153-159; Steiner, S. and N.L. Anderson (2000) Toxicol. Lett. 112-113:467-471, expressly incorporated by reference herein). If a test compound has a signature similar to that of a compound with known toxicity, it is likely to share those toxic properties. These fingerprints or signatures are most useful and refined when they contain expression information from a large number of genes and gene families. Ideally, a genome-wide measurement of expression provides the highest quality signature. Even genes whose expression is not altered by any tested compounds are important as well, as the levels of expression of these genes are used to normalize the rest of the expression data. The normalization procedure is useful for comparison of expression data after treatment with different compounds. While the assignment of gene function to elements of a toxicant signature aids in interpretation of toxicity mechanisms, knowledge of gene function is not necessary for the statistical matching of signatures which leads to prediction of toxicity. (See, for example, Press Release 00-02 from the National Institute of Environmental Health Sciences, released February 29, 2000, available at http://www.niehs.nih.gov/oc/news/toxchip.htm.) Therefore, it is important and desirable in toxicological screening using toxicant signatures to include all expressed gene sequences.

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In one embodiment, the toxicity of a test compound is assessed by treating a biological sample containing nucleic acids with the test compound. Nucleic acids that are expressed in the treated biological sample are hybridized with one or more probes specific to the polynucleotides of the present invention, so that transcript levels corresponding to the polynucleotides of the present invention may be quantified. The transcript levels in the treated biological sample are compared with levels in an untreated biological sample. Differences in the transcript levels between the two samples are indicative of a toxic response caused by the test compound in the treated sample.

Another particular embodiment relates to the use of the polypeptide sequences of the present invention to analyze the proteome of a tissue or cell type. The term proteome refers to the global pattern of protein expression in a particular tissue or cell type. Each protein component of a proteome can be subjected individually to further analysis. Proteome expression patterns, or profiles, are

analyzed by quantifying the number of expressed proteins and their relative abundance under given conditions and at a given time. A profile of a cell's proteome may thus be generated by separating and analyzing the polypeptides of a particular tissue or cell type. In one embodiment, the separation is achieved using two-dimensional gel electrophoresis, in which proteins from a sample are separated by isoelectric focusing in the first dimension, and then according to molecular weight by sodium dodecyl sulfate slab gel electrophoresis in the second dimension (Steiner and Anderson, supra). The proteins are visualized in the gel as discrete and uniquely positioned spots, typically by staining the gel with an agent such as Coomassie Blue or silver or fluorescent stains. The optical density of each protein spot is generally proportional to the level of the protein in the sample. The optical densities of equivalently positioned protein spots from different samples, for example, from biological samples either treated or untreated with a test compound or therapeutic agent, are compared to identify any changes in protein spot density related to the treatment. The proteins in the spots are partially sequenced using, for example, standard methods employing chemical or enzymatic cleavage followed by mass spectrometry. The identity of the protein in a spot may be determined by comparing its partial sequence, preferably of at least 5 contiguous amino acid residues, to the polypeptide sequences of the present invention. In some cases, further sequence data may be obtained for definitive protein identification.

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A proteomic profile may also be generated using antibodies specific for PKIN to quantify the levels of PKIN expression. In one embodiment, the antibodies are used as elements on a microarray, and protein expression levels are quantified by exposing the microarray to the sample and detecting the levels of protein bound to each array element (Lueking, A. et al. (1999) Anal. Biochem. 270:103-111; Mendoze, L.G. et al. (1999) Biotechniques 27:778-788). Detection may be performed by a variety of methods known in the art, for example, by reacting the proteins in the sample with a thiol- or aminoreactive fluorescent compound and detecting the amount of fluorescence bound at each array element.

Toxicant signatures at the proteome level are also useful for toxicological screening, and should be analyzed in parallel with toxicant signatures at the transcript level. There is a poor correlation between transcript and protein abundances for some proteins in some tissues (Anderson, N.L. and J. Seilhamer (1997) Electrophoresis 18:533-537), so proteome toxicant signatures may be useful in the analysis of compounds which do not significantly affect the transcript image, but which alter the proteomic profile. In addition, the analysis of transcripts in body fluids is difficult, due to rapid degradation of mRNA, so proteomic profiling may be more reliable and informative in such cases.

In another embodiment, the toxicity of a test compound is assessed by treating a biological sample containing proteins with the test compound. Proteins that are expressed in the treated biological sample are separated so that the amount of each protein can be quantified. The amount of each protein is compared to the amount of the corresponding protein in an untreated biological sample. A difference

in the amount of protein between the two samples is indicative of a toxic response to the test compound in the treated sample. Individual proteins are identified by sequencing the amino acid residues of the individual proteins and comparing these partial sequences to the polypeptides of the present invention.

In another embodiment, the toxicity of a test compound is assessed by treating a biological sample containing proteins with the test compound. Proteins from the biological sample are incubated with antibodies specific to the polypeptides of the present invention. The amount of protein recognized by the antibodies is quantified. The amount of protein in the treated biological sample is compared with the amount in an untreated biological sample. A difference in the amount of protein between the two samples is indicative of a toxic response to the test compound in the treated sample.

Microarrays may be prepared, used, and analyzed using methods known in the art. (See, e.g., Brennan, T.M. et al. (1995) U.S. Patent No. 5,474,796; Schena, M. et al. (1996) Proc. Natl. Acad. Sci. USA 93:10614-10619; Baldeschweiler et al. (1995) PCT application WO95/251116; Shalon, D. et al. (1995) PCT application WO95/35505; Heller, R.A. et al. (1997) Proc. Natl. Acad. Sci. USA 94:2150-2155; and Heller, M.J. et al. (1997) U.S. Patent No. 5,605,662.) Various types of microarrays are well known and thoroughly described in <u>DNA Microarrays: A Practical Approach</u>, M. Schena, ed. (1999) Oxford University Press, London, hereby expressly incorporated by reference.

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In another embodiment of the invention, nucleic acid sequences encoding PKIN may be used to generate hybridization probes useful in mapping the naturally occurring genomic sequence. Either coding or noncoding sequences may be used, and in some instances, noncoding sequences may be preferable over coding sequences. For example, conservation of a coding sequence among members of a multi-gene family may potentially cause undesired cross hybridization during chromosomal mapping. The sequences may be mapped to a particular chromosome, to a specific region of a chromosome, or to artificial chromosome constructions, e.g., human artificial chromosomes (HACs), yeast artificial chromosomes (YACs), bacterial artificial chromosomes (BACs), bacterial P1 constructions, or single chromosome cDNA libraries. (See, e.g., Harrington, J.J. et al. (1997) Nat. Genet. 15:345-355; Price, C.M. (1993) Blood Rev. 7:127-134; and Trask, B.J. (1991) Trends Genet. 7:149-154.) Once mapped, the nucleic acid sequences of the invention may be used to develop genetic linkage maps, for example, which correlate the inheritance of a disease state with the inheritance of a particular chromosome region or restriction fragment length polymorphism (RFLP). (See, for example, Lander, E.S. and D. Botstein (1986) Proc. Natl. Acad. Sci. USA 83:7353-7357.)

Fluorescent <u>in situ</u> hybridization (FISH) may be correlated with other physical and genetic map data. (See, e.g., Heinz-Ulrich, et al. (1995) in Meyers, <u>supra</u>, pp. 965-968.) Examples of genetic map data can be found in various scientific journals or at the Online Mendelian Inheritance in Man (OMIM) World Wide Web site. Correlation between the location of the gene encoding PKIN on a physical map

and a specific disorder, or a predisposition to a specific disorder, may help define the region of DNA associated with that disorder and thus may further positional cloning efforts.

In situ hybridization of chromosomal preparations and physical mapping techniques, such as linkage analysis using established chromosomal markers, may be used for extending genetic maps. Often the placement of a gene on the chromosome of another mammalian species, such as mouse, may reveal associated markers even if the exact chromosomal locus is not known. This information is valuable to investigators searching for disease genes using positional cloning or other gene discovery techniques. Once the gene or genes responsible for a disease or syndrome have been crudely localized by genetic linkage to a particular genomic region, e.g., ataxia-telangiectasia to 11q22-23, any sequences mapping to that area may represent associated or regulatory genes for further investigation. (See, e.g., Gatti, R.A. et al. (1988) Nature 336:577-580.) The nucleotide sequence of the instant invention may also be used to detect differences in the chromosomal location due to translocation, inversion, etc., among normal, carrier, or affected individuals.

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In another embodiment of the invention, PKIN, its catalytic or immunogenic fragments, or oligopeptides thereof can be used for screening libraries of compounds in any of a variety of drug screening techniques. The fragment employed in such screening may be free in solution, affixed to a solid support, borne on a cell surface, or located intracellularly. The formation of binding complexes between PKIN and the agent being tested may be measured.

Another technique for drug screening provides for high throughput screening of compounds having suitable binding affinity to the protein of interest. (See, e.g., Geysen, et al. (1984) PCT application WO84/03564.) In this method, large numbers of different small test compounds are synthesized on a solid substrate. The test compounds are reacted with PKIN, or fragments thereof, and washed. Bound PKIN is then detected by methods well known in the art. Purified PKIN can also be coated directly onto plates for use in the aforementioned drug screening techniques. Alternatively, non-neutralizing antibodies can be used to capture the peptide and immobilize it on a solid support.

In another embodiment, one may use competitive drug screening assays in which neutralizing antibodies capable of binding PKIN specifically compete with a test compound for binding PKIN. In this manner, antibodies can be used to detect the presence of any peptide which shares one or more antigenic determinants with PKIN.

In additional embodiments, the nucleotide sequences which encode PKIN may be used in any molecular biology techniques that have yet to be developed, provided the new techniques rely on properties of nucleotide sequences that are currently known, including, but not limited to, such properties as the triplet genetic code and specific base pair interactions.

Without further elaboration, it is believed that one skilled in the art can, using the preceding description, utilize the present invention to its fullest extent. The following embodiments are, therefore, to be construed as merely illustrative, and not limitative of the remainder of the disclosure in any way whatsoever.

The disclosures of all patents, applications and publications, mentioned above and below, including U.S. Ser. No. 60/212,073, U.S. Ser. No. 60/213,467, U.S. Ser. No. 60/215,651, U.S. Ser. No. 60/216,605, U.S. Ser. No. 60/218,372, and U.S. Ser. No. 60/228,056 are expressly incorporated by reference herein.

10 EXAMPLES

## I. Construction of cDNA Libraries

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Incyte cDNAs were derived from cDNA libraries described in the LIFESEQ GOLD database (Incyte Genomics, Palo Alto CA) and shown in Table 4, column 5. Some tissues were homogenized and lysed in guanidinium isothiocyanate, while others were homogenized and lysed in phenol or in a suitable mixture of denaturants, such as TRIZOL (Life Technologies), a monophasic solution of phenol and guanidine isothiocyanate. The resulting lysates were centrifuged over CsCl cushions or extracted with chloroform. RNA was precipitated from the lysates with either isopropanol or sodium acetate and ethanol, or by other routine methods.

Phenol extraction and precipitation of RNA were repeated as necessary to increase RNA purity. In some cases, RNA was treated with DNase. For most libraries, poly(A)+ RNA was isolated using oligo d(T)-coupled paramagnetic particles (Promega), OLIGOTEX latex particles (QIAGEN, Chatsworth CA), or an OLIGOTEX mRNA purification kit (QIAGEN). Alternatively, RNA was isolated directly from tissue lysates using other RNA isolation kits, e.g., the POLY(A)PURE mRNA purification kit (Ambion, Austin TX).

In some cases, Stratagene was provided with RNA and constructed the corresponding cDNA libraries. Otherwise, cDNA was synthesized and cDNA libraries were constructed with the UNIZAP vector system (Stratagene) or SUPERSCRIPT plasmid system (Life Technologies), using the recommended procedures or similar methods known in the art. (See, e.g., Ausubel, 1997, <a href="supra">supra</a>, units 5.1-6.6.) Reverse transcription was initiated using oligo d(T) or random primers. Synthetic oligonucleotide adapters were ligated to double stranded cDNA, and the cDNA was digested with the appropriate restriction enzyme or enzymes. For most libraries, the cDNA was size-selected (300-1000 bp) using SEPHACRYL S1000, SEPHAROSE CL2B, or SEPHAROSE CL4B column chromatography (Amersham Pharmacia Biotech) or preparative agarose gel electrophoresis. cDNAs were ligated into compatible restriction enzyme sites of the polylinker of a suitable plasmid, e.g.,

PBLUESCRIPT plasmid (Stratagene), PSPORT1 plasmid (Life Technologies), PCDNA2.1 plasmid (Invitrogen, Carlsbad CA), PBK-CMV plasmid (Stratagene), or pINCY (Incyte Genomics, Palo Alto CA), or derivatives thereof. Recombinant plasmids were transformed into competent <u>E. coli</u> cells including XL1-Blue, XL1-BlueMRF, or SOLR from Stratagene or DH5α, DH10B, or ElectroMAX DH10B from Life Technologies.

### II. Isolation of cDNA Clones

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Plasmids obtained as described in Example I were recovered from host cells by <u>in vivo</u> excision using the UNIZAP vector system (Stratagene) or by cell lysis. Plasmids were purified using at least one of the following: a Magic or WIZARD Minipreps DNA purification system (Promega); an AGTC Miniprep purification kit (Edge Biosystems, Gaithersburg MD); and QIAWELL 8 Plasmid, QIAWELL 8 Plasmid, QIAWELL 8 Ultra Plasmid purification systems or the R.E.A.L. PREP 96 plasmid purification kit from QIAGEN. Following precipitation, plasmids were resuspended in 0.1 ml of distilled water and stored, with or without lyophilization, at 4°C.

Alternatively, plasmid DNA was amplified from host cell lysates using direct link PCR in a high-throughput format (Rao, V.B. (1994) Anal. Biochem. 216:1-14). Host cell lysis and thermal cycling steps were carried out in a single reaction mixture. Samples were processed and stored in 384-well plates, and the concentration of amplified plasmid DNA was quantified fluorometrically using PICOGREEN dye (Molecular Probes, Eugene OR) and a FLUOROSKAN II fluorescence scanner (Labsystems Oy, Helsinki, Finland).

# 20 III. Sequencing and Analysis

Incyte cDNA recovered in plasmids as described in Example II were sequenced as follows. Sequencing reactions were processed using standard methods or high-throughput instrumentation such as the ABI CATALYST 800 (Applied Biosystems) thermal cycler or the PTC-200 thermal cycler (MJ Research) in conjunction with the HYDRA microdispenser (Robbins Scientific) or the 25 MICROLAB 2200 (Hamilton) liquid transfer system. cDNA sequencing reactions were prepared using reagents provided by Amersham Pharmacia Biotech or supplied in ABI sequencing kits such as the ABI PRISM BIGDYE Terminator cycle sequencing ready reaction kit (Applied Biosystems). Electrophoretic separation of cDNA sequencing reactions and detection of labeled polynucleotides were carried out using the MEGABACE 1000 DNA sequencing system (Molecular Dynamics); the ABI PRISM 373 or 377 sequencing system (Applied Biosystems) in conjunction with standard ABI 30 protocols and base calling software; or other sequence analysis systems known in the art. Reading frames within the cDNA sequences were identified using standard methods (reviewed in Ausubel, 1997, supra, unit 7.7). Some of the cDNA sequences were selected for extension using the techniques disclosed in Example VIII.

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The polynucleotide sequences derived from Incyte cDNAs were validated by removing vector, linker, and poly(A) sequences and by masking ambiguous bases, using algorithms and programs based on BLAST, dynamic programming, and dinucleotide nearest neighbor analysis. The Incyte cDNA sequences or translations thereof were then queried against a selection of public databases such as the GenBank primate, rodent, mammalian, vertebrate, and eukaryote databases, and BLOCKS, PRINTS, DOMO, PRODOM, and hidden Markov model (HMM)-based protein family databases such as PFAM. (HMM is a probabilistic approach which analyzes consensus primary structures of gene families. See, for example, Eddy, S.R. (1996) Curr. Opin. Struct. Biol. 6:361-365.) The queries were performed using programs based on BLAST, FASTA, BLIMPS, and HMMER. The Incyte cDNA 10 sequences were assembled to produce full length polynucleotide sequences. Alternatively, GenBank cDNAs, GenBank ESTs, stitched sequences, stretched sequences, or Genscan-predicted coding sequences (see Examples IV and V) were used to extend Incyte cDNA assemblages to full length. Assembly was performed using programs based on Phred, Phrap, and Consed, and cDNA assemblages were screened for open reading frames using programs based on GeneMark, BLAST, and FASTA. The full length polynucleotide sequences were translated to derive the corresponding full length polypeptide sequences. Alternatively, a polypeptide of the invention may begin at any of the methionine residues of the full length translated polypeptide. Full length polypeptide sequences were subsequently analyzed by querying against databases such as the GenBank protein databases (genpept), SwissProt, BLOCKS, PRINTS, DOMO, PRODOM, Prosite, and hidden Markov model (HMM)-based protein 20 family databases such as PFAM. Full length polynucleotide sequences are also analyzed using MACDNASIS PRO software (Hitachi Software Engineering, South San Francisco CA) and LASERGENE software (DNASTAR). Polynucleotide and polypeptide sequence alignments are generated using default parameters specified by the CLUSTAL algorithm as incorporated into the MEGALIGN multisequence alignment program (DNASTAR), which also calculates the percent identity between aligned sequences.

Table 7 summarizes the tools, programs, and algorithms used for the analysis and assembly of Incyte cDNA and full length sequences and provides applicable descriptions, references, and threshold parameters. The first column of Table 7 shows the tools, programs, and algorithms used, the second column provides brief descriptions thereof, the third column presents appropriate references, all of which are incorporated by reference herein in their entirety, and the fourth column presents, where applicable, the scores, probability values, and other parameters used to evaluate the strength of a match between two sequences (the higher the score or the lower the probability value, the greater the identity between two sequences).

The programs described above for the assembly and analysis of full length polynucleotide and polypeptide sequences were also used to identify polynucleotide sequence fragments from SEQ ID NO:27-52. Fragments from about 20 to about 4000 nucleotides which are useful in hybridization and amplification technologies are described in Table 4, column 4.

### 5 IV. Identification and Editing of Coding Sequences from Genomic DNA

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Putative human kinases were initially identified by running the Genscan gene identification program against public genomic sequence databases (e.g., gbpri and gbhtg). Genscan is a generalpurpose gene identification program which analyzes genomic DNA sequences from a variety of organisms (See Burge, C. and S. Karlin (1997) J. Mol. Biol. 268:78-94, and Burge, C. and S. Karlin (1998) Curr. Opin. Struct. Biol. 8;346-354). The program concatenates predicted exons to form an assembled cDNA sequence extending from a methionine to a stop codon. The output of Genscan is a FASTA database of polynucleotide and polypeptide sequences. The maximum range of sequence for Genscan to analyze at once was set to 30 kb. To determine which of these Genscan predicted cDNA sequences encode human kinases, the encoded polypeptides were analyzed by querying against PFAM models for human kinases. Potential human kinases were also identified by homology to Incyte cDNA sequences that had been annotated as human kinases. These selected Genscan-predicted sequences were then compared by BLAST analysis to the genpept and gbpri public databases. Where necessary, the Genscan-predicted sequences were then edited by comparison to the top BLAST hit from genpept to correct errors in the sequence predicted by Genscan, such as extra or omitted exons. BLAST analysis was also used to find any Incyte cDNA or public cDNA coverage of the Genscan-predicted sequences, thus providing evidence for transcription. When Incyte cDNA coverage was available, this information was used to correct or confirm the Genscan predicted sequence. Full length polynucleotide sequences were obtained by assembling Genscan-predicted coding sequences with Incyte cDNA sequences and/or public cDNA sequences using the assembly process described in Example III. Alternatively, full length polynucleotide sequences were derived entirely from edited or unedited Genscan-predicted coding sequences.

### V. Assembly of Genomic Sequence Data with cDNA Sequence Data "Stitched" Sequences

Partial cDNA sequences were extended with exons predicted by the Genscan gene identification program described in Example IV. Partial cDNAs assembled as described in Example III were mapped to genomic DNA and parsed into clusters containing related cDNAs and Genscan exon predictions from one or more genomic sequences. Each cluster was analyzed using an algorithm based on graph theory and dynamic programming to integrate cDNA and genomic information, generating possible splice variants that were subsequently confirmed, edited, or extended to create a full length sequence.

Sequence intervals in which the entire length of the interval was present on more than one sequence in the cluster were identified, and intervals thus identified were considered to be equivalent by transitivity. For example, if an interval was present on a cDNA and two genomic sequences, then all three intervals were considered to be equivalent. This process allows unrelated but consecutive genomic sequences to be brought together, bridged by cDNA sequence. Intervals thus identified were then "stitched" together by the stitching algorithm in the order that they appear along their parent sequences to generate the longest possible sequence, as well as sequence variants. Linkages between intervals which proceed along one type of parent sequence (cDNA to cDNA or genomic sequence to genomic sequence) were given preference over linkages which change parent type (cDNA to genomic sequence). The resultant stitched sequences were translated and compared by BLAST analysis to the genpept and gbpri public databases. Incorrect exons predicted by Genscan were corrected by comparison to the top BLAST hit from genpept. Sequences were further extended with additional cDNA sequences, or by inspection of genomic DNA, when necessary.

### "Stretched" Sequences

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Partial DNA sequences were extended to full length with an algorithm based on BLAST analysis. First, partial cDNAs assembled as described in Example III were queried against public databases such as the GenBank primate, rodent, mammalian, vertebrate, and eukaryote databases using the BLAST program. The nearest GenBank protein homolog was then compared by BLAST analysis to either Incyte cDNA sequences or GenScan exon predicted sequences described in Example IV. A chimeric protein was generated by using the resultant high-scoring segment pairs (HSPs) to map the translated sequences onto the GenBank protein homolog. Insertions or deletions may occur in the chimeric protein with respect to the original GenBank protein homolog. The GenBank protein homolog, the chimeric protein, or both were used as probes to search for homologous genomic sequences from the public human genome databases. Partial DNA sequences were therefore "stretched" or extended by the addition of homologous genomic sequences. The resultant stretched sequences were examined to determine whether it contained a complete gene.

### VI. Chromosomal Mapping of PKIN Encoding Polynucleotides

The sequences which were used to assemble SEQ ID NO:27-52 were compared with sequences from the Incyte LIFESEQ database and public domain databases using BLAST and other implementations of the Smith-Waterman algorithm. Sequences from these databases that matched SEQ ID NO:27-52 were assembled into clusters of contiguous and overlapping sequences using assembly algorithms such as Phrap (Table 7). Radiation hybrid and genetic mapping data available from public resources such as the Stanford Human Genome Center (SHGC), Whitehead Institute for Genome Research (WIGR), and Généthon were used to determine if any of the clustered sequences

had been previously mapped. Inclusion of a mapped sequence in a cluster resulted in the assignment of all sequences of that cluster, including its particular SEQ ID NO:, to that map location.

Map locations are represented by ranges, or intervals, of human chromosomes. The map position of an interval, in centiMorgans, is measured relative to the terminus of the chromosome's p-arm. (The centiMorgan (cM) is a unit of measurement based on recombination frequencies between chromosomal markers. On average, 1 cM is roughly equivalent to 1 megabase (Mb) of DNA in humans, although this can vary widely due to hot and cold spots of recombination.) The cM distances are based on genetic markers mapped by Généthon which provide boundaries for radiation hybrid markers whose sequences were included in each of the clusters. Human genome maps and other resources available to the public, such as the NCBI "GeneMap'99" World Wide Web site (http://www.ncbi.nlm.nih.gov/genemap/), can be employed to determine if previously identified disease genes map within or in proximity to the intervals indicated above.

In this manner, SEQ ID NO:27 was mapped to chromosome 19 and SEQ ID NO:35 was mapped to chromosome 15 within the interval from 72.30 to 77.30 centiMorgans. SEQ ID NO:48 was mapped to chromosome 10 within the interval from 93.80 to 96.90 centiMorgans. SEQ ID NO:49 was mapped to chromosome 13 within the interval from 11.60 to 22.80 centiMorgans, to chromosome 17 within the interval from 0.60 to 14.80 centiMorgans, and to chromosome 20 within the interval from 57.70 to 64.10 centiMorgans. More than one map location is reported for SEQ ID NO:49, indicating that sequences having different map locations were assembled into a single cluster. This situation occurs, for example, when sequences having strong similarity, but not complete identity, are assembled into a single cluster.

### VII. Analysis of Polynucleotide Expression

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Northern analysis is a laboratory technique used to detect the presence of a transcript of a gene and involves the hybridization of a labeled nucleotide sequence to a membrane on which RNAs from a particular cell type or tissue have been bound. (See, e.g., Sambrook, <u>supra</u>, ch. 7; Ausubel (1995) <u>supra</u>, ch. 4 and 16.)

Analogous computer techniques applying BLAST were used to search for identical or related molecules in cDNA databases such as GenBank or LIFESEQ (Incyte Genomics). This analysis is much faster than multiple membrane-based hybridizations. In addition, the sensitivity of the computer search can be modified to determine whether any particular match is categorized as exact or similar. The basis of the search is the product score, which is defined as:

### BLAST Score x Percent Identity 5 x minimum {length(Seq. 1), length(Seq. 2)}

The product score takes into account both the degree of similarity between two sequences and the length of the sequence match. The product score is a normalized value between 0 and 100, and is calculated as follows: the BLAST score is multiplied by the percent nucleotide identity and the product is divided by (5 times the length of the shorter of the two sequences). The BLAST score is calculated by assigning a score of +5 for every base that matches in a high-scoring segment pair (HSP), and -4 for every mismatch. Two sequences may share more than one HSP (separated by gaps). If there is more than one HSP, then the pair with the highest BLAST score is used to calculate the product score. The product score represents a balance between fractional overlap and quality in a BLAST alignment. For example, a product score of 100 is produced only for 100% identity over the entire length of the shorter of the two sequences being compared. A product score of 70 is produced either by 100% identity and 70% overlap at one end, or by 88% identity and 100% overlap at the other. A product score of 50 is produced either by 100% identity and 50% overlap at one end, or 79% identity and 100% overlap.

Alternatively, polynucleotide sequences encoding PKIN are analyzed with respect to the tissue sources from which they were derived. For example, some full length sequences are assembled, at least in part, with overlapping Incyte cDNA sequences (see Example III). Each cDNA sequence is derived from a cDNA library constructed from a human tissue. Each human tissue is classified into one of the following organ/tissue categories: cardiovascular system; connective tissue; digestive system; embryonic structures; endocrine system; exocrine glands; genitalia, female; genitalia, male; germ cells; hemic and immune system; liver; musculoskeletal system; nervous system; pancreas; respiratory system; sense organs; skin; stomatognathic system; unclassified/mixed; or urinary tract. The number of libraries in each category is counted and divided by the total number of libraries across all categories. Similarly, each human tissue is classified into one of the following disease/condition categories: cancer, cell line, developmental, inflammation, neurological, trauma, cardiovascular, pooled, and other, and the number of libraries in each category is counted and divided by the total number of libraries across all categories. The resulting percentages reflect the tissue- and disease-specific expression of cDNA encoding PKIN. cDNA sequences and cDNA library/tissue information are found in the LIFESEQ GOLD database (Incyte Genomics, Palo Alto CA).

### VIII. Extension of PKIN Encoding Polynucleotides

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Full length polynucleotide sequences were also produced by extension of an appropriate fragment of the full length molecule using oligonucleotide primers designed from this fragment. One primer was synthesized to initiate 5' extension of the known fragment, and the other primer was

synthesized to initiate 3' extension of the known fragment. The initial primers were designed using OLIGO 4.06 software (National Biosciences), or another appropriate program, to be about 22 to 30 nucleotides in length, to have a GC content of about 50% or more, and to anneal to the target sequence at temperatures of about 68°C to about 72°C. Any stretch of nucleotides which would result in hairpin structures and primer-primer dimerizations was avoided.

Selected human cDNA libraries were used to extend the sequence. If more than one extension was necessary or desired, additional or nested sets of primers were designed.

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High fidelity amplification was obtained by PCR using methods well known in the art. PCR was performed in 96-well plates using the PTC-200 thermal cycler (MJ Research, Inc.). The reaction mix contained DNA template, 200 nmol of each primer, reaction buffer containing Mg<sup>2+</sup>, (NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub>, and 2-mercaptoethanol, Taq DNA polymerase (Amersham Pharmacia Biotech), ELONGASE enzyme (Life Technologies), and Pfu DNA polymerase (Stratagene), with the following parameters for primer pair PCI A and PCI B: Step 1: 94°C, 3 min; Step 2: 94°C, 15 sec; Step 3: 60°C, 1 min; Step 4: 68°C, 2 min; Step 5: Steps 2, 3, and 4 repeated 20 times; Step 6: 68°C, 5 min; Step 7: storage at 4°C. In the alternative, the parameters for primer pair T7 and SK+ were as follows: Step 1: 94°C, 3 min; Step 2: 94°C, 15 sec; Step 3: 57°C, 1 min; Step 4: 68°C, 2 min; Step 5: Steps 2, 3, and 4 repeated 20 times; Step 6: 68°C, 5 min; Step 7: storage at 4°C.

The concentration of DNA in each well was determined by dispensing 100  $\mu$ l PICOGREEN quantitation reagent (0.25% (v/v) PICOGREEN; Molecular Probes, Eugene OR) dissolved in 1X TE and 0.5  $\mu$ l of undiluted PCR product into each well of an opaque fluorimeter plate (Corning Costar, Acton MA), allowing the DNA to bind to the reagent. The plate was scanned in a Fluoroskan II (Labsystems Oy, Helsinki, Finland) to measure the fluorescence of the sample and to quantify the concentration of DNA. A 5  $\mu$ l to 10  $\mu$ l aliquot of the reaction mixture was analyzed by electrophoresis on a 1% agarose gel to determine which reactions were successful in extending the sequence.

The extended nucleotides were desalted and concentrated, transferred to 384-well plates, digested with CviJI cholera virus endonuclease (Molecular Biology Research, Madison WI), and sonicated or sheared prior to religation into pUC 18 vector (Amersham Pharmacia Biotech). For shotgun sequencing, the digested nucleotides were separated on low concentration (0.6 to 0.8%) agarose gels, fragments were excised, and agar digested with Agar ACE (Promega). Extended clones were religated using T4 ligase (New England Biolabs, Beverly MA) into pUC 18 vector (Amersham Pharmacia Biotech), treated with Pfu DNA polymerase (Stratagene) to fill-in restriction site overhangs, and transfected into competent <u>E. coli</u> cells. Transformed cells were selected on antibiotic-containing media, and individual colonies were picked and cultured overnight at 37°C in 384-well plates in LB/2x carb liquid media.

The cells were lysed, and DNA was amplified by PCR using Taq DNA polymerase (Amersham Pharmacia Biotech) and Pfu DNA polymerase (Stratagene) with the following parameters: Step 1: 94°C, 3 min; Step 2: 94°C, 15 sec; Step 3: 60°C, 1 min; Step 4: 72°C, 2 min; Step 5: steps 2, 3, and 4 repeated 29 times; Step 6: 72°C, 5 min; Step 7: storage at 4°C. DNA was quantified by PICOGREEN reagent (Molecular Probes) as described above. Samples with low DNA recoveries were reamplified using the same conditions as described above. Samples were diluted with 20% dimethysulfoxide (1:2, v/v), and sequenced using DYENAMIC energy transfer sequencing primers and the DYENAMIC DIRECT kit (Amersham Pharmacia Biotech) or the ABI PRISM BIGDYE Terminator cycle sequencing ready reaction kit (Applied Biosystems).

In like manner, full length polynucleotide sequences are verified using the above procedure or are used to obtain 5' regulatory sequences using the above procedure along with oligonucleotides designed for such extension, and an appropriate genomic library.

### Labeling and Use of Individual Hybridization Probes IX.

Hybridization probes derived from SEQ ID NO:27-52 are employed to screen cDNAs, genomic DNAs, or mRNAs. Although the labeling of oligonucleotides, consisting of about 20 base pairs, is specifically described, essentially the same procedure is used with larger nucleotide fragments. Oligonucleotides are designed using state-of-the-art software such as OLIGO 4.06 software (National Biosciences) and labeled by combining 50 pmol of each oligomer, 250  $\mu$ Ci of [ $\gamma$ -32P] adenosine triphosphate (Amersham Pharmacia Biotech), and T4 polynucleotide kinase (DuPont NEN, Boston MA). The labeled oligonucleotides are substantially purified using a SEPHADEX G-25 superfine size 20 exclusion dextran bead column (Amersham Pharmacia Biotech). An aliquot containing 10<sup>7</sup> counts per minute of the labeled probe is used in a typical membrane-based hybridization analysis of human genomic DNA digested with one of the following endonucleases: Ase I, Bgl II, Eco RI, Pst I, Xba I, or Pvu II (DuPont NEN).

The DNA from each digest is fractionated on a 0.7% agarose gel and transferred to nylon membranes (Nytran Plus, Schleicher & Schuell, Durham NH). Hybridization is carried out for 16 hours at 40°C. To remove nonspecific signals, blots are sequentially washed at room temperature under conditions of up to, for example, 0.1 x saline sodium citrate and 0.5% sodium dodecyl sulfate. Hybridization patterns are visualized using autoradiography or an alternative imaging means and compared.

### X. Microarrays

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The linkage or synthesis of array elements upon a microarray can be achieved utilizing photolithography, piezoelectric printing (ink-jet printing, See, e.g., Baldeschweiler, supra.), mechanical microspotting technologies, and derivatives thereof. The substrate in each of the aforementioned

technologies should be uniform and solid with a non-porous surface (Schena (1999), <u>supra</u>). Suggested substrates include silicon, silica, glass slides, glass chips, and silicon wafers. Alternatively, a procedure analogous to a dot or slot blot may also be used to arrange and link elements to the surface of a substrate using thermal, UV, chemical, or mechanical bonding procedures. A typical array may be produced using available methods and machines well known to those of ordinary skill in the art and may contain any appropriate number of elements. (See, e.g., Schena, M. et al. (1995) Science 270:467-470; Shalon, D. et al. (1996) Genome Res. 6:639-645; Marshall, A. and J. Hodgson (1998) Nat. Biotechnol. 16:27-31.)

Full length cDNAs, Expressed Sequence Tags (ESTs), or fragments or oligomers thereof may comprise the elements of the microarray. Fragments or oligomers suitable for hybridization can be selected using software well known in the art such as LASERGENE software (DNASTAR). The array elements are hybridized with polynucleotides in a biological sample. The polynucleotides in the biological sample are conjugated to a fluorescent label or other molecular tag for ease of detection. After hybridization, nonhybridized nucleotides from the biological sample are removed, and a fluorescence scanner is used to detect hybridization at each array element. Alternatively, laser desorbtion and mass spectrometry may be used for detection of hybridization. The degree of complementarity and the relative abundance of each polynucleotide which hybridizes to an element on the microarray may be assessed. In one embodiment, microarray preparation and usage is described in detail below.

### 20 <u>Tissue or Cell Sample Preparation</u>

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Total RNA is isolated from tissue samples using the guanidinium thiocyanate method and poly(A)<sup>+</sup> RNA is purified using the oligo-(dT) cellulose method. Each poly(A)<sup>+</sup> RNA sample is reverse transcribed using MMLV reverse-transcriptase, 0.05 pg/µl oligo-(dT) primer (21mer), 1X first strand buffer, 0.03 units/µl RNase inhibitor, 500 µM dATP, 500 µM dGTP, 500 µM dTTP, 40 µM dCTP-Cy3 (BDS) or dCTP-Cy5 (Amersham Pharmacia Biotech). The reverse transcription reaction is performed in a 25 ml volume containing 200 ng poly(A)<sup>+</sup> RNA with GEMBRIGHT kits (Incyte). Specific control poly(A)<sup>+</sup> RNAs are synthesized by in vitro transcription from non-coding yeast genomic DNA. After incubation at 37°C for 2 hr, each reaction sample (one with Cy3 and another with Cy5 labeling) is treated with 2.5 ml of 0.5M sodium hydroxide and incubated for 20 minutes at 85°C to the stop the reaction and degrade the RNA. Samples are purified using two successive CHROMA SPIN 30 gel filtration spin columns (CLONTECH Laboratories, Inc. (CLONTECH), Palo Alto CA) and after combining, both reaction samples are ethanol precipitated using 1 ml of glycogen (1 mg/ml), 60 ml sodium acetate, and 300 ml of 100% ethanol. The sample is

then dried to completion using a SpeedVAC (Savant Instruments Inc., Holbrook NY) and resuspended in 14 µl 5X SSC/0.2% SDS.

### **Microarray Preparation**

Sequences of the present invention are used to generate array elements. Each array element is amplified from bacterial cells containing vectors with cloned cDNA inserts. PCR amplification uses primers complementary to the vector sequences flanking the cDNA insert. Array elements are amplified in thirty cycles of PCR from an initial quantity of 1-2 ng to a final quantity greater than 5 µg. Amplified array elements are then purified using SEPHACRYL-400 (Amersham Pharmacia Biotech).

Purified array elements are immobilized on polymer-coated glass slides. Glass microscope slides (Corning) are cleaned by ultrasound in 0.1% SDS and acetone, with extensive distilled water washes between and after treatments. Glass slides are etched in 4% hydrofluoric acid (VWR Scientific Products Corporation (VWR), West Chester PA), washed extensively in distilled water, and coated with 0.05% aminopropyl silane (Sigma) in 95% ethanol. Coated slides are cured in a 110°C oven.

Array elements are applied to the coated glass substrate using a procedure described in US Patent No. 5,807,522, incorporated herein by reference. 1  $\mu$ l of the array element DNA, at an average concentration of 100 ng/ $\mu$ l, is loaded into the open capillary printing element by a high-speed robotic apparatus. The apparatus then deposits about 5 nl of array element sample per slide.

Microarrays are UV-crosslinked using a STRATALINKER UV-crosslinker (Stratagene). Microarrays are washed at room temperature once in 0.2% SDS and three times in distilled water. Non-specific binding sites are blocked by incubation of microarrays in 0.2% casein in phosphate buffered saline (PBS) (Tropix, Inc., Bedford MA) for 30 minutes at 60°C followed by washes in 0.2% SDS and distilled water as before.

### Hybridization

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Hybridization reactions contain 9  $\mu$ l of sample mixture consisting of 0.2  $\mu$ g each of Cy3 and Cy5 labeled cDNA synthesis products in 5X SSC, 0.2% SDS hybridization buffer. The sample mixture is heated to 65°C for 5 minutes and is aliquoted onto the microarray surface and covered with an 1.8 cm² coverslip. The arrays are transferred to a waterproof chamber having a cavity just slightly larger than a microscope slide. The chamber is kept at 100% humidity internally by the addition of 140  $\mu$ l of 5X SSC in a corner of the chamber. The chamber containing the arrays is incubated for about 6.5 hours at 60°C. The arrays are washed for 10 min at 45°C in a first wash buffer (1X SSC, 0.1% SDS), three times for 10 minutes each at 45°C in a second wash buffer (0.1X SSC), and dried.

**Detection** 

Reporter-labeled hybridization complexes are detected with a microscope equipped with an Innova 70 mixed gas 10 W laser (Coherent, Inc., Santa Clara CA) capable of generating spectral lines at 488 nm for excitation of Cy3 and at 632 nm for excitation of Cy5. The excitation laser light is focused on the array using a 20X microscope objective (Nikon, Inc., Melville NY). The slide containing the array is placed on a computer-controlled X-Y stage on the microscope and raster-scanned past the objective. The 1.8 cm x 1.8 cm array used in the present example is scanned with a resolution of 20 micrometers.

In two separate scans, a mixed gas multiline laser excites the two fluorophores sequentially. Emitted light is split, based on wavelength, into two photomultiplier tube detectors (PMT R1477, Hamamatsu Photonics Systems, Bridgewater NJ) corresponding to the two fluorophores. Appropriate filters positioned between the array and the photomultiplier tubes are used to filter the signals. The emission maxima of the fluorophores used are 565 nm for Cy3 and 650 nm for Cy5. Each array is typically scanned twice, one scan per fluorophore using the appropriate filters at the laser source, although the apparatus is capable of recording the spectra from both fluorophores simultaneously.

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The sensitivity of the scans is typically calibrated using the signal intensity generated by a cDNA control species added to the sample mixture at a known concentration. A specific location on the array contains a complementary DNA sequence, allowing the intensity of the signal at that location to be correlated with a weight ratio of hybridizing species of 1:100,000. When two samples from different sources (e.g., representing test and control cells), each labeled with a different fluorophore, are hybridized to a single array for the purpose of identifying genes that are differentially expressed, the calibration is done by labeling samples of the calibrating cDNA with the two fluorophores and adding identical amounts of each to the hybridization mixture.

The output of the photomultiplier tube is digitized using a 12-bit RTI-835H analog-to-digital (A/D) conversion board (Analog Devices, Inc., Norwood MA) installed in an IBM-compatible PC computer. The digitized data are displayed as an image where the signal intensity is mapped using a linear 20-color transformation to a pseudocolor scale ranging from blue (low signal) to red (high signal). The data is also analyzed quantitatively. Where two different fluorophores are excited and measured simultaneously, the data are first corrected for optical crosstalk (due to overlapping emission spectra) between the fluorophores using each fluorophore's emission spectrum.

A grid is superimposed over the fluorescence signal image such that the signal from each spot is centered in each element of the grid. The fluorescence signal within each element is then integrated to obtain a numerical value corresponding to the average intensity of the signal. The software used for signal analysis is the GEMTOOLS gene expression analysis program (Incyte).

### XI. Complementary Polynucleotides

Sequences complementary to the PKIN-encoding sequences, or any parts thereof, are used to detect, decrease, or inhibit expression of naturally occurring PKIN. Although use of oligonucleotides comprising from about 15 to 30 base pairs is described, essentially the same procedure is used with smaller or with larger sequence fragments. Appropriate oligonucleotides are designed using OLIGO 4.06 software (National Biosciences) and the coding sequence of PKIN. To inhibit transcription, a complementary oligonucleotide is designed from the most unique 5' sequence and used to prevent promoter binding to the coding sequence. To inhibit translation, a complementary oligonucleotide is designed to prevent ribosomal binding to the PKIN-encoding transcript.

### 10 XII. Expression of PKIN

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Expression and purification of PKIN is achieved using bacterial or virus-based expression systems. For expression of PKIN in bacteria, cDNA is subcloned into an appropriate vector containing an antibiotic resistance gene and an inducible promoter that directs high levels of cDNA transcription. Examples of such promoters include, but are not limited to, the trp-lac (tac) hybrid promoter and the T5 or T7 bacteriophage promoter in conjunction with the *lac* operator regulatory element. Recombinant vectors are transformed into suitable bacterial hosts, e.g., BL21(DE3). Antibiotic resistant bacteria express PKIN upon induction with isopropyl beta-D-thiogalactopyranoside (IPTG). Expression of PKIN in eukaryotic cells is achieved by infecting insect or mammalian cell lines with recombinant Autographica californica nuclear polyhedrosis virus (AcMNPV), commonly known as baculovirus. The nonessential polyhedrin gene of baculovirus is replaced with cDNA encoding PKIN by either homologous recombination or bacterial-mediated transposition involving transfer plasmid intermediates. Viral infectivity is maintained and the strong polyhedrin promoter drives high levels of cDNA transcription. Recombinant baculovirus is used to infect Spodoptera frugiperda (Sf9) insect cells in most cases, or human hepatocytes, in some cases. Infection of the latter requires additional genetic modifications to baculovirus. (See Engelhard, E.K. et al. (1994) Proc. Natl. Acad. Sci. USA 91:3224-3227; Sandig, V. et al. (1996) Hum. Gene Ther. 7:1937-1945.)

In most expression systems, PKIN is synthesized as a fusion protein with, e.g., glutathione S-transferase (GST) or a peptide epitope tag, such as FLAG or 6-His, permitting rapid, single-step, affinity-based purification of recombinant fusion protein from crude cell lysates. GST, a 26-kilodalton enzyme from Schistosoma japonicum, enables the purification of fusion proteins on immobilized glutathione under conditions that maintain protein activity and antigenicity (Amersham Pharmacia Biotech). Following purification, the GST moiety can be proteolytically cleaved from PKIN at specifically engineered sites. FLAG, an 8-amino acid peptide, enables immunoaffinity purification using commercially available monoclonal and polyclonal anti-FLAG antibodies (Eastman Kodak). 6-

His, a stretch of six consecutive histidine residues, enables purification on metal-chelate resins (QIAGEN). Methods for protein expression and purification are discussed in Ausubel (1995, <u>supra</u>, ch. 10 and 16). Purified PKIN obtained by these methods can be used directly in the assays shown in Examples XVI, XVII, and XVIII where applicable.

### 5 XIII. Functional Assays

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PKIN function is assessed by expressing the sequences encoding PKIN at physiologically elevated levels in mammalian cell culture systems. cDNA is subcloned into a mammalian expression vector containing a strong promoter that drives high levels of cDNA expression. Vectors of choice include PCMV SPORT (Life Technologies) and PCR3.1 (Invitrogen, Carlsbad CA), both of which contain the cytomegalovirus promoter.  $5-10 \mu g$  of recombinant vector are transiently transfected into a human cell line, for example, an endothelial or hematopoietic cell line, using either liposome formulations or electroporation. 1-2  $\mu$ g of an additional plasmid containing sequences encoding a marker protein are co-transfected. Expression of a marker protein provides a means to distinguish transfected cells from nontransfected cells and is a reliable predictor of cDNA expression from the recombinant vector. Marker proteins of choice include, e.g., Green Fluorescent Protein (GFP; Clontech), CD64, or a CD64-GFP fusion protein. Flow cytometry (FCM), an automated, laser opticsbased technique, is used to identify transfected cells expressing GFP or CD64-GFP and to evaluate the apoptotic state of the cells and other cellular properties. FCM detects and quantifies the uptake of fluorescent molecules that diagnose events preceding or coincident with cell death. These events include changes in nuclear DNA content as measured by staining of DNA with propidium iodide; changes in cell size and granularity as measured by forward light scatter and 90 degree side light scatter; downregulation of DNA synthesis as measured by decrease in bromodeoxyuridine uptake; alterations in expression of cell surface and intracellular proteins as measured by reactivity with specific antibodies; and alterations in plasma membrane composition as measured by the binding of fluorescein-conjugated Annexin V protein to the cell surface. Methods in flow cytometry are discussed in Ormerod, M.G. (1994) Flow Cytometry, Oxford, New York NY.

The influence of PKIN on gene expression can be assessed using highly purified populations of cells transfected with sequences encoding PKIN and either CD64 or CD64-GFP. CD64 and CD64-GFP are expressed on the surface of transfected cells and bind to conserved regions of human immunoglobulin G (IgG). Transfected cells are efficiently separated from nontransfected cells using magnetic beads coated with either human IgG or antibody against CD64 (DYNAL, Lake Success NY). mRNA can be purified from the cells using methods well known by those of skill in the art. Expression of mRNA encoding PKIN and other genes of interest can be analyzed by northern analysis or microarray techniques.

### XIV. Production of PKIN Specific Antibodies

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PKIN substantially purified using polyacrylamide gel electrophoresis (PAGE; see, e.g., Harrington, M.G. (1990) Methods Enzymol. 182:488-495), or other purification techniques, is used to immunize rabbits and to produce antibodies using standard protocols.

Alternatively, the PKIN amino acid sequence is analyzed using LASERGENE software (DNASTAR) to determine regions of high immunogenicity, and a corresponding oligopeptide is synthesized and used to raise antibodies by means known to those of skill in the art. Methods for selection of appropriate epitopes, such as those near the C-terminus or in hydrophilic regions are well described in the art. (See, e.g., Ausubel, 1995, supra, ch. 11.)

Typically, oligopeptides of about 15 residues in length are synthesized using an ABI 431A peptide synthesizer (Applied Biosystems) using FMOC chemistry and coupled to KLH (Sigma-Aldrich, St. Louis MO) by reaction with N-maleimidobenzoyl-N-hydroxysuccinimide ester (MBS) to increase immunogenicity. (See, e.g., Ausubel, 1995, <a href="supra">supra</a>.) Rabbits are immunized with the oligopeptide-KLH complex in complete Freund's adjuvant. Resulting antisera are tested for antipeptide and anti-PKIN activity by, for example, binding the peptide or PKIN to a substrate, blocking with 1% BSA, reacting with rabbit antisera, washing, and reacting with radio-iodinated goat anti-rabbit IgG.

### XV. Purification of Naturally Occurring PKIN Using Specific Antibodies

Naturally occurring or recombinant PKIN is substantially purified by immunoaffinity chromatography using antibodies specific for PKIN. An immunoaffinity column is constructed by covalently coupling anti-PKIN antibody to an activated chromatographic resin, such as CNBr-activated SEPHAROSE (Amersham Pharmacia Biotech). After the coupling, the resin is blocked and washed according to the manufacturer's instructions.

Media containing PKIN are passed over the immunoaffinity column, and the column is washed under conditions that allow the preferential absorbance of PKIN (e.g., high ionic strength buffers in the presence of detergent). The column is eluted under conditions that disrupt antibody/PKIN binding (e.g., a buffer of pH 2 to pH 3, or a high concentration of a chaotrope, such as urea or thiocyanate ion), and PKIN is collected.

### XVI. Identification of Molecules Which Interact with PKIN

PKIN, or biologically active fragments thereof, are labeled with <sup>125</sup>I Bolton-Hunter reagent. (See, e.g., Bolton A.E. and W.M. Hunter (1973) Biochem. J. 133:529-539.) Candidate molecules previously arrayed in the wells of a multi-well plate are incubated with the labeled PKIN, washed, and any wells with labeled PKIN complex are assayed. Data obtained using different concentrations of PKIN are used to calculate values for the number, affinity, and association of PKIN with the candidate molecules.

Alternatively, molecules interacting with PKIN are analyzed using the yeast two-hybrid system as described in Fields, S. and O. Song (1989) Nature 340:245-246, or using commercially available kits based on the two-hybrid system, such as the MATCHMAKER system (Clontech).

PKIN may also be used in the PATHCALLING process (CuraGen Corp., New Haven CT) which employs the yeast two-hybrid system in a high-throughput manner to determine all interactions between the proteins encoded by two large libraries of genes (Nandabalan, K. et al. (2000) U.S. Patent No. 6,057,101).

### XVII. Demonstration of PKIN Activity

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Generally, protein kinase activity is measured by quantifying the phosphorylation of a protein substrate by PKIN in the presence of gamma-labeled <sup>32</sup>P-ATP. PKIN is incubated with the protein substrate, <sup>32</sup>P-ATP, and an appropriate kinase buffer. The <sup>32</sup>P incorporated into the substrate is separated from free <sup>32</sup>P-ATP by electrophoresis and the incorporated <sup>32</sup>P is counted using a radioisotope counter. The amount of incorporated <sup>32</sup>P is proportional to the activity of PKIN. A determination of the specific amino acid residue phosphorylated is made by phosphoamino acid analysis of the hydrolyzed protein.

In one alternative, protein kinase activity is measured by quantifying the transfer of gamma phosphate from adenosine triphosphate (ATP) to a serine, threonine or tyrosine residue in a protein substrate. The reaction occurs between a protein kinase sample with a biotinylated peptide substrate and gamma <sup>32</sup>P-ATP. Following the reaction, free avidin in solution is added for binding to the biotinylated <sup>32</sup>P-peptide product. The binding sample then undergoes a centrifugal ultrafiltration process with a membrane which will retain the product-avidin complex and allow passage of free gamma <sup>32</sup>P-ATP. The reservoir of the centrifuged unit containing the <sup>32</sup>P-peptide product as retentate is then counted in a scintillation counter. This procedure allows assay of any type of protein kinase sample, depending on the peptide substrate and kinase reaction buffer selected. This assay is provided in kit form (ASUA, Affinity Ultrafiltration Separation Assay, Transbio Corporation, Baltimore MD, U.S. Patent No. 5,869,275). Suggested substrates and their respective enzymes are as follows: Histone H1 (Sigma) and p34<sup>ede2</sup>kinase, Annexin I, Angiotensin (Sigma) and EGF receptor kinase, Annexin II and *src* kinase, ERK1 & ERK2 substrates and MEK, and myelin basic protein and ERK (Pearson, J.D. et al. (1991) Methods Enzymol. 200:62-81).

In another alternative, protein kinase activity of PKIN is demonstrated <u>in vitro</u> in an assay containing PKIN, 50µl of kinase buffer, 1µg substrate, such as myelin basic protein (MBP) or synthetic peptide substrates, 1 mM DTT, 10 µg ATP, and  $0.5\mu\text{Ci}$  [ $\gamma$ - $^{33}$ P]ATP. The reaction is incubated at 30°C for 30 minutes and stopped by pipetting onto P81 paper. The unincorporated [ $\gamma$ - $^{33}$ P]ATP is removed by washing and the incorporated radioactivity is measured using a radioactivity scintillation

counter. Alternatively, the reaction is stopped by heating to 100°C in the presence of SDS loading buffer and visualized on a 12% SDS polyacrylamide gel by autoradiography. Incorporated radioactivity is corrected for reactions carried out in the absence of PKIN or in the presence of the inactive kinase, K38A.

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In yet another alternative, adenylate kinase or guanylate kinase activity may be measured by the incorporation of <sup>32</sup>P from gamma-labeled <sup>32</sup>P -ATP into ADP or GDP using a gamma radioisotope counter. The enzyme, in a kinase buffer, is incubated together with the appropriate nucleotide mono-phosphate substrate (AMP or GMP) and <sup>32</sup>P-labeled ATP as the phosphate donor. The reaction is incubated at 37°C and terminated by addition of trichloroacetic acid. The acid extract is neutralized and subjected to gel electrophoresis to separate the mono-, di-, and triphosphonucleotide fractions. The diphosphonucleotide fraction is cut out and counted. The radioactivity recovered is proportional to the enzyme activity.

In yet another alternative, other assays for PKIN include scintillation proximity assays (SPA), scintillation plate technology and filter binding assays. Useful substrates include recombinant proteins tagged with glutathione transferase, or synthetic peptide substrates tagged with biotin. Inhibitors of PKIN activity, such as small organic molecules, proteins or peptides, may be identified by such assays. **XVIII.** Enhancement/Inhibition of Protein Kinase Activity

Agonists or antagonists of PKIN activation or inhibition may be tested using assays described in section XVII. Agonists cause an increase in PKIN activity and antagonists cause a decrease in PKIN activity.

Various modifications and variations of the described methods and systems of the invention will be apparent to those skilled in the art without departing from the scope and spirit of the invention. Although the invention has been described in connection with certain embodiments, it should be understood that the invention as claimed should not be unduly limited to such specific embodiments. Indeed, various modifications of the described modes for carrying out the invention which are obvious to those skilled in molecular biology or related fields are intended to be within the scope of the following claims.

Table 1

### Table 2

Polypeptide SEQ ID NO:  2 3 4 5 6	Incyte Polypeptide ID 2011384CD1 2014888CD1 2258952CD1 7473244CD1 1242491CD1 2634875CD1 3951059CD1	GenBank ID NO:  9404634  913540326  92983205  913603881  93766209  92052189  92052189  92052189  92052189  93599507	Probability score 4.50E-60 1.00E-159 2.70E-08 0 0 8.40E-25 0 1.50E-272	serine/threonine kinase al (1994) Gene 139 (2), 235-239) ens] serine/threonine kinase FKS cus] ser/thr protein kinase al (1998) Nature 392 (6674), 35 ens] serine/threonine kinase 31 al, (2001) Nat. Genet. 27 (4), 4 IRE1 al (1998) EMBO J. 17 (19), 5708- ficus] serine/threonine kinase thalianal MAP3K delta-1 protein kinalianal MAP3K delta-1 protein kinase thal (1999) Gene 229 (1-2), 171- ens] skeletal myosin light chain kin et al (1990) J. Biol. Chem. 265, rho/rac-interacting citron kina thal (1998) J. Biol. Chem. 273 (
8 0	7475546CD1	g5815139 g3435114	0 1.80E-50	[Mus musculus] nuclear body associated kinase la [Homo sapiens] serine/threonine kinase ULK1 [Windstranger H of 2] (1908) Canomics 51 (1) 76.85)
10	7477076CD1 1874092CD1	g854733 g2511715	6.20E-198 4.00E-25	icus] casein kinase 1 gamma 1 i haliana] putative phosphatidyli
12	4841542CD1	g927732	3.30E-67	[Saccharomyces cerevisiae] Snflp: serine/threonine protein kinase;
13	7472695CD1	g1498250	1.10E-53	[Dictyostellum discoldeum] myosin light chain kinase (Tan, J.L. et al (1991) J. Biol. Chem. 266, 16044-16049)
14	7477966CD1	g3766209	0	g,X.Z. et al (1998) EMBO J. 17 (19), 57

Table 2 (cont.)

-				
SEQ ID NO:	incyte Polypeptide ID	NO:	score	GEILBALIK HOMOLOG
15	7163416CD1	g7649810	2.10E-135	[Homo sapiens] protein kinase PAK5
16	7472822CD1	g5081459	3.70E-241	rio] p55-relate
17	7477486CD1	g3217028	0	[5' incom][Homo sapiens] putative serine/threonine protein kinase
			į	(Stanchi, F. et al (2001) Yeast 18 (1), 69-80)
18	3773709CD1	93986088	6.70E-78	[Pyrococcus kodakaraensis] Glycerol Kinase
19	7477204CD1	g992672	7.30E-129	[Homo sapiens] G protein-coupled receptor kinase GRK4-
				Deca (Premont, R.T. et al (1996) J. Biol. Chem. 271 (11), 6403-6410)
		g4001826	0	[f]][Spermonhilus tridecemlineatus] G protein-coupled
			1	(K7) (1998) Mol 1745 //
	20100001		7000	1770) TOTT . 473. 47
0 7	SULBSBSCLL	G4521218	4./05-45	
21	63497CD1	g1213224	0	[Rattus norvegicus] SNF1-related kinase
				(6) 667 .
22	1625436CD1	g4096108	1.10E-252	[Homo sapiens] proline rich calmodulin-dependent
				protein kinase
		g206152	0	[fl][Rattus norvegicus] calmodulin-dependent protein
				Kinase II gamma subunit (EC 2./.I.3/)
23	3330646CD1	g406058	0	s] protein kinase
				den, P.D.
24	3562763CD1	g12830335	0	[5' incom][Homo sapiens] bA55008.2 (novel protein kinase)
	•	g1510182	9.80E-18	sculus] cyclin-dependent kinase 5
				(Ishizuka, T. et al (1995) Gene 166 (2), 267-271)
25	621293CD1	g2649941	4.50E-23	[Archaeoglobus fulgidus] adenylate kinase (adk) (Klenk,H.P. et al (1997) Nature 390 (6658), 364-370)
26	7480774CD1	g2463542	0	1,4,5-trisphosphate

Table .

Analytical	Methods and	Databases	BLAST_DOMO	HMMER_PFAM	PROFILESCAN	MOTIFS	MOTIFS	BLIMPS_PRINTS	HMMER_PFAM	BLAST_DOMO	BLAST_DOMO	BLAST_DOMO	BLAST_PRODOM	
Signature Sequences,			PROTEIN KINASE DOMAIN DM00004 P27448 58-297: R16-R255	Eukaryotic protein kinase domain pkinase: V12-L267	Protein kinases signatures and profile, protein kinase_tyr.prf: 0111-G163	Protein Kinase ATP binding site: I18-K41	Protein Kinase (serine/threonine): L131-L143	Tyrosine kinase catalytic domain signature: PR00109:Y125-L143 Y193-S215	Eukaryotic protein kinase domain pkinase: P135-L228	DM00004   P54744   13-263 PROTEIN KINASE DOMAIN: P113-L228 (P=1.1e-06)	PROTEIN KINASE DOMAIN DM00004 Q09499 536-784:P534-A784	KINASE; THREONINE; ATP; SERINE; DM06305 Q09499 786-924:V787-Y922	PROTEIN KINASE/ENDORIBONULCEASE PUTATIVE BLAST_PRODOM SERINE/THREONINE PROTEIN KINASE C41C4.4	CHROMOSOME II PRECURSOR TRANSFERASE PD152704:T170-L395,L61-E163
Potential	Glycosyla-	tion Sites									N200			
Potential	Phosphorylation	Sites	Y12 Y23 T17 S144 T30 S31 S237 S253						S190 S50 S51 T141 Y302		S207 S299 S500 S503 S580 S609	S65 S714 S814 S852 S857 T116	T128 T147 T175 T188 T202 T345	T55 T592 T658 T84 T895 T905 T936 Y146
Amino	Acid	Residues	273						329		938			
Incyte	Polypeptide	1D							2004888CD1		2258952CD1			
SEQ	A	:ON	H				<del>*</del>		2		ന_			OL YAME

Table 3 (cont.)

	Incyte	Amino	Potentia1	Potential	Signature Sequences,	Analytical
Pol	Polypeptide	Acid	Phosphorylation	Glycosyla-	Domains and Motifs	Methods and
H		Residues	Sites	tion Sites		Databases
					SERINE/THREONINE PROTEIN KINASE PRECURSOR TRANSMEMBRANE SIGNAL	BLAST_PRODOM
					TRANSFERASE ATP-BINDING PROTEIN IRE1	
				•	GLYCOPROTEIN PD032590:W794-Y922	
					Tyrosine kinase catalytic domain	BLIMPS_PRINTS
					PR00109: H639-I657, G694-L704, V716-	
					Protein kinases signatures and profile	PROFILESCAN
					protein_kinase_tyr.prf:	
					E625-G682	
					Eukaryotic protein kinase domain	HMMER_PFAM
	_				pkinase:	
					E532-E793	
					Protein_Kinase serine/theronine:	MOTIFS
					I645-I657	
77	7473244CD1	795	S2 S301	S35 N17 N331	Protein kinases signatures and profile	PROFILESCAN
			S423 S468 S485	N397 N398	protein_kinase_tyr.prf:	
			S486 S49 S524		Y133-6210	
			S546 S609 S666		Eukaryotic protein kinase domain	HMMER_PFAM
			8671 8699 8705		pkinase:	
_			S710 S776 T128		Y60-M311	
			T19 T282 T324		PROTEIN KINASE DOMAIN	BLAST_DOMO
	_		T333 T437 T504		DM00004   P27448   58-297:L62-L302	
			T568	_		
			T657			
ال			T680 T82 T9			

Table 3 (cont.)

Analytical	Methods and	Databases	BLAST_PRODOM	BLAST_PRODOM	BLAST_PRODOM		BLAST_PRODOM		BLIMPS_PRINTS		MOTIFS	MOTIFS	HMMER_PFAM		PROFILESCAN		MOTIFS			MOTIFS		
Signature Sequences,	Domains and Motifs		KINASE SERINE/THREONINEP ROTEIN PROTEIN TRANSFERASE ATP-BINDING SERINE/THREONINE PUTATIVE KIN1 EMK PAR1 PD004300:G682-L795	SERINE/THREONINE KINASE PD119193:1594-P665	KINASE SERINE/THREONINE PROTEIN SERINE/THREONINE PUTATIVE TRANSFERASE ATP-BINDING PROTEIN EMK P78 CDC25C	PD008571:S412-S595	KINASE SERINE/THREONINE PROTEIN PUTATIVE BLAST_PRODOM   SERINE/THREONINE TRANSFERASE ATP-BINDING	PROTEIN PAR1 KP78 EMK PD005838:M311-R411	Tyrosine kinase catalytic domain	PR00109: M136-V149, Y172-L190, V238- Q260	Protein Kinase_ATP binding site: I66-K89	Protein Kinase serine/theronine: I178-L190	Eukaryotic protein kinase domain:	L14-V257	Protein kinases signatures and profile:	L99-Q151	Protein kinases ATP-binding region	signature:	L14-K35	Serine/Threonine protein kinases active-	site signature:	I119-L131
Potential	Glycosyla-	tion Sites											N293 N424	N437								
Potentia1	Phosphorylation	Sites											S309 S42 S540	<b>S</b> 583	T270		T509		T623 T653			
Amino	Acid	Residues											959									
Incyte	lypeptide	ID			-								1242491CD1									
SEQ	A	 NO	4	<del></del>									5									

# Table 3 (cont.)

	Amino	Potential	Potential	Signature Sequences,	Analytical
Polypeptide Acid	Acid	Phosphorylation	Glycosyla-	nd Motifs	Methods and
	Residues	Sites	tion Sites		Databases
				Tyrosine kinase catalytic domain	BLIMPS_PRINTS
				signature	
				PR00109:M76-Q89, Y113-L131, A183-G205,	
				P232-S254	
				PROTEIN KINASE DOMAIN	BLAST_DOMO
				DM00004   P42679   236-470:L14-P252	
				DM00004   I49621   195-428:L14-P252	
				DM00004   I38044   100-349:L13-P252	
				DM00004 Q05609 553-797:L14-T197, L14-	
				T253	
2634875CD1	596	S107 S143 S157 S159 S184 S203	N278 N416	Eukaryotic protein kinase domain: M285-L540	HMMER_PFAM
		ıo		Tyrosine kinase catalytic domain	BLIMPS_PRINTS
	-	io		signature	
		T224 T247 T301		PR00109:M359-V372, F396-C414, T463-	
	*	T320 T351 T379		D485	
		T49 X376		Protein kinases ATP-binding region	MOTIFS
				signature:	
				L291-K314	
				Serine/Threonine protein kinases active-MOTIFS	MOTIFS
				site signature:	
				V402-C414	
				KINASE MYOSIN LIGHT CHAIN SKELETAL	BLAST_PRODOM
				MUSCLE MLCK TRANSFERASE SERINE/THREONINE	
				CALMODULIN BINDING	
				PD036174:A95-M285	
				PD027051:L540-V596	
				PD029157:A2-R82, A2-S90	

Table 3 (cont.)

Analytical	Methods and	Databases	BLAST_DOMO					HMMER_PFAM		HMMER_PFAM		BLIMPS_PRINTS	=		MOTIFS	•		MOTIFS			BLAST_PRODOM				BLAST_PRODOM			
Signature Sequences,			PROTEIN KINASE DOMAIN	DM00004   P07313   298-541: S287-A531	DM00004 JN0583 [727-969:K288-N530	DM00004   S07571   5152-5396:E289-M529	DM00004   P53355   15-257: E289-M529	Eukaryotic protein kinase domain:	F97-F360	ase C terminal domain:	S361-E390	Tyrosine kinase catalytic domain	signature	PR00109:M174-N187, S211-V229	Protein kinases ATP-binding region	signature:	V103-K126	Serine/Threonine protein kinases active-MOTIFS	site signature:	Y217-V229	RHO/RACINTERACTING CITRON KINASE SHORT	ISOFORM	PD154232:S422-V468	PD154360:M1-M43	KINASE RHO ASSOCIATED COILED COIL	PROTEIN FORMING RHO/RAC INTERACTING	CITRON ALPHA	PD007970:032-D96
Potential	syla-	tion Sites																										
Potential	osphorylation	Sites						S140 S248 S308	61	S410 S436 S445	S490 S81 S93	T279 T378 T83					-											
Amino		Residues					!	497																				
Incyte	Polypeptide Acid	ID						3951059CD1																				
SEQ	q	NO:	9					7	<b>780</b>			<b></b>									-							

# Table 3 (cont.)

Analytical Methods and Databases	BLAST_DOMO	HMMER_PFAM BLIMPS_PRINTS		MOTIFS	MOTIFS	BLAST_PRODOM	BLAST_PRODOM	BLAST_PRODOM
Signature Sequences, Domains and Motifs	PROTEIN KINASE DOMAIN  DM00004   Q09013   83-336: V99-L349  DM00004   S42867   75-498: S101-G241,  I258-S445  DM00004   S42864   41-325: E98-G241, N249- L349, D96-T153  DM00004   P38679   238-527:L102-G241,  I258-L349, E86-A124	Eukaryotic protein kinase domain: Y199-P420, R498-V527 Tyrosine kinase catalytic domain	signature PR00109:K314-L332	Protein kinases ATP-binding region signature: L205-K228	Serine/Threonine protein kinases active-MOTIFS site signature: L320-L332	PROTEIN KINASE NUCLEAR HOMEO DOMAIN INTERACTING DNA-BINDING SERINE/THREONINE PD141983:A573-C933 PD150874:A993-I1171	PROTEIN KINASE NUCLEAR SERINE/THREONINE HOMEO DOMAIN INTERACTING DNA-BINDING SERINE/THREONINE F20B6.8 PD042899:L425-P574	HOMEO DOMAIN INTERACTING PROTEIN KINASE 2 DNA-BINDING NUCLEAR PROTEIN PD184491:E872-P961
Potential Glycosyla- tion Sites		N140 N157 N271 N480 N562 N579			×111-2-4			
Potential Phosphorylation Sites		S121 S135 S178 S180 S254 S27 S37 S405 S649	S774 S804	S970 T119 T172 T221 T431 T450 T483 T517 T839	T867 T893 T995 T1022 S1027 S1099 Y443 Y468			
Amino Acid Residues		1171						
Incyte Polypeptide ID		7395890CD1						
SEQ ID NO:	2	∞	************					

Table 3 (cont.)

Analytical Methods and Databases	BLAST_DOMO	HMMER_PFAM BLIMPS_PRINTS	PROFILESCAN	MOTIFS	MOTIFS	BLAST_PRODOM	BLAST_DOMO
Signature Sequences, Domains and Motifs	PROTEIN KINASE DOMAIN  DM00004   P14680   371-694: V201-P518  DM00004   Q09815   519-804: E200-L473,  F500-T517  DM00004   P49657   101-409: L205-P518  DM00004   Q09690   700-985: E200-P444,  F500-P518	Eukaryotic protein kinase domain: F14-V270 Tyrosine kinase catalytic domain signature PR01-H104 F127-F145 F239-F261	Protein kinases signatures and profile: V113-P166	Protein kinases ATP-binding region signature: L20-K44	Serine/Threonine protein kinases active-MOTIFS site signature: I133-L145	KINASE PROTEIN TRANSFERASE ATP BINDING SERINE/THREONINE RECEPTOR TYROSINE PRECURSOR TRANSMEMBRANE PD000001:S176-P255, I15-F93, P237-W269, F117-M164, L20-K34	PROTEIN KINASE DOMAIN  DM00004   P53104   26-315: P151-F261, E18- E111, F117-S147  DM00004   S54788   154-400:L20-S260  DM00004   P27448   58-297: L16-R258  DM00004   P49673   31-267: L20-I259
Potential Glycosyla- tion Sites		N132					
Potential Phosphorylation Sites		\$217 \$219 \$227 \$256 \$260 \$339 \$361 \$406 \$447	7 T61				
Amino Acid Residues		470					
Incyte Polypeptide ID		7475546CD1					
SEQ ID NO:	ω	o					

Table 3 (cont.)

									_				- 40 later
Analytical Methods and Databases	HMMER_PFAM	PROFILESCAN	MOTIFS	MOTIFS	BLAST_PRODOM		BLAST_PRODOM		BLAST_DOMO			BLAST_PRODOM	
Signature Sequences, Domains and Motifs	Eukaryotic protein kinase domain pkinase: F44-E276	Protein kinases signatures and profile: T140-P197	Protein kinases ATP-binding region signature: I50-K73	Serine/Threonine protein kinases active-MOTIFS site signature:	CASEIN KINASE I GAMMA ISOFORM TPANSFERASE SERINE/THREONINE ATP RINDING	MULTIGENE PD015080:F315-T393	GAMMA 1 ISOF TRANSFERASE	SERINE/THREONINE PROTEIN ATP BINDING MULTIGENE PD049080:M1-N43	PROTEIN KINASE DOMAIN	DM00004 A56711 46-303:V46-Y304 DM00004 C56711 45-301:V46-Y304	DM00004 B56711 48-303:V46-Y304 DM00004 D56406 31-276:V46-V293	PROTEIN PHOSPHATIDYL INOSITOL 4-PHOSPHATE 5-KINASE PUTATIVE T22C1.7	ISOLOG ATPIP5K1 T4C15.16 PD149995: L13-D204
Potential Glycosyla- tion Sites													
Potential Phosphorylation Sites	S124 S150 S229 S96 T137 T14 T199 T214 T258	69 T273										S121 S132 S78 T197 T84	
Amino Acid Residues	1											240	
Incyte Polypeptide ID	7477076CD1										44	1874092CD1	
SEQ ID NO:	10						· · · · · · · · · · · · · · · · · · ·					11	

Table 3 (cont.)

Analytical	Methods and	Databases	BLAST_PRODOM	BLAST_DOMO	BLIMPS_PRINTS	HMMER_PFAM	MOTIFS	MOTIFS	нммек_рғам
Signature Sequences,	Domains and Motifs		KINASE PROTEIN TRANSFERASE ATP-BINDING SERINE/THREONINE PROTEIN PHOSPHORYLATION RECEPTOR TYROSINE PROTEIN PRECURSOR TRANSMEMBRANE PD000001: K3-S163, S178-F216, P236- W268 (P=1.2e-09)	PROTEIN KINASE DOMAIN  DM00004   P27448   58-297: L22-L260  DM00004   P06782   57-296: L22-L260  DM00004   JC1446   20-261: T24-L260  DM00004   P54645   17-258: E23-L260	Tyrosine kinase catalytic domain signature PR00109: M95-S108, Y131-L149, V197- H219	Eukaryotic protein kinase domain pkinase: Y19-V269	Protein_Kinase_ATP L25-K47	Protein_Kinase_Serine/Threonine V137-L149	Eukaryotic protein kinase domain pkinase: Y75-L340
Potential	Glycosyla-	tion Sites	N542 N87		AR				N172 N370 N397 N54
Potential	Phosphorylation	Sites	S108 S114 S293 S297 S305 S306 S339 S343 S382 S40 S427 S48 S489 S572 S88 S99 T193 T255	T259 T357 T477 T544 T582 Y425					\$128 \$170 \$208 \$233 \$255 \$285 \$30 \$308 \$347 \$346 \$379 \$39 \$340 \$432 \$46 \$1143 \$129 \$130 \$1409 \$1418 \$1469 \$193
Amino	Acid	idues	594						473
Incyte	lypeptide	ID	4841542CD1						7472695CD1
SEQ	AI.	NO:	12		·				13

Table 3 (cont.)

Analytical	Methods and	Databases	BLAST_DOMO			BLAST_PRODOM			BLIMPS_PRINTS		BLIMPS_PRINTS		MOTIFS	MOTIFS	PROFILESCAN	HMMER_PFAM						
Signature Sequences,	Domains and Motifs		PROTEIN KINASE DOMAIN DM00004 S57347 21-266: F77-T330	DM00004 S46283 13-259: G78-A331	DM00004 S54788 154- 400: G78-A331	KINASE PROTEIN TRANSFERASE ATP-BINDING	SEKINE/IRKEONINE FROIEIN FROSFROKILALION RECEPTOR TYROSINE PROTEIN PRECURSOR	TRANSMEMBRANE P197-L299, R79-D156	Ι.	PR00109: M151-D164, Y187-V205, C263-	Phosphorylase kinase family	FKIUIU49: UI04-II84	Protein_Kinase_ATP L81-K104	Protein_Kinase_Serine/Threonine I193-V205	protein_kinase_tyrosine.profile: E173-A228	Eukaryotic protein kinase domain	pkinase: F541-F802					
Potential	Glycosyla-	tion Sites														N200						
Potential	Phosphorylation	Sites														207 \$299	S511 S589 S618 S65 S723 S823	86	128	188	52	T84 T904 T914 T945 Y146
Amino	Acid	Residues														947				-		
Incyte	lypeptide	日														7477966CD1						
SEQ	日	NO:	13					****************			×					14				م ہے		

Table 3 (cont.)

Analytical Methods and Databases	BLAST_DOMO	BLAST_DOMO	BLAST_PRODOM	BLAST_PRODOM	BLIMPS_PRINTS	BLIMPS_PRINTS	MOTIFS	PROFILESCAN	HMMER_PFAM
Signature Sequences,  Domains and Motifs		; THREONINE; ATP; SERINE;   Q09499   786-924: V796-Y931   P32361   972-1114: Q795-L928	PROTEIN KINASE/ENDORIBONULCEASE PUTATIVE BLAST_PRODOM SERINE/THREONINE PROTEIN KINASE C41C4.4 CHROMOSOME II PRECURSOR TRANSFERASE PD152704: T170-L395, L61-E163	IRE1	Tyrosine kinase catalytic domain signature PR00109: H648-I666, G703-L713, V725- D747	phorylase kinase family signature 11049: P794-R805	Protein_Kinase_Serine/Threonine: 1 1654-1666	protein kinase_tyrosine.profile: E634-G691	Eukaryotic protein kinase domain pkinase: L407-Y601
Potential Glycosyla- tion Sites									N288
Potential Phosphorylation Sites									S107 S135 S165 S189 S248 S255 S276 S290 S332 S351 S429 S560 S624 T106 T107 T124 T212 T238 T24 T322 T46 T505 T580 T99
Amino Acid Residnes									641
Incyte Polypeptide Th									7163416CD1
SEQ ID NO:	14	*				- Andrews			17

Table 3 (cont.)

Analytical	Methods and	Databases	BLAST_DOMO				BLAST_PRODOM					BLIMPS PRINTS				MOTIFS		HMMER_PFAM		BLAST_DOMO						
Signature Sequences,	Domains and Motifs		PROTEIN KINASE DOMAIN DM00004 P35465 271-510: Y410-S628	DM00004 I49376 270-509: K412-S628	DM00004 Q03497 622-861: V411-S628	DM00004 P50527 388-627: S409-S628	KINASE SERINE/THREONINE PROTEIN	TRANSFERASE ATP-BINDING PROTEIN	PHOSPHORYLATION P21 ACTIVATED ACTIVATED	HOMOLOG SYNDROME	PD002852: I12-L44 (P=3.0e-06)	Tyrosine kinase catalytic domain	signature	PR00109: M481-S494, Y516-L534, G563-	I573, V582-D604	Protein_Kinase_ATP	I413-K436	Guanylate kinase:	T404-N500	GUANYLATE KINASE	DM00755 A57653 370-570: P359-P570	I38757	DM00755 S32545 1-196: R369-K556	P31007	-	
Potential	Glycosyla-	tion Sites											•					N334								
Potential	Phosphorylation	Sites																S136	<b>S</b> 266	S313 S318 S323	\$336	<b>S</b> 206	T213		T470	Y482 Y59
Amino	Acid	Residues																9/5								
Incyte	Polypeptide	1D								-								7472822CD1								
SEQ	Q	:ON	15						_	-			-					16								

Table 3 (cont.)

Analytical	Methods and	Databases	GUANYLATE BLAST_PRODOM REPEAT GMP			IERAL PLASMA BLAST_PRODOM	ING CASK CAMGUK		PROTEIN MAGUK P55 SUBFAMILY MEMBER DISCS BLAST_PRODOM			LY MEMBER MPP3 BLAST_PRODOM			BLIMPS_BLOCKS	:8-R475	BLIMPS_PRINTS	1-P241, A252-		DHR or GLGF). HMMER_PFAM				HMMER_PFAM	HMMER_PFAM	HMMER_PFAM MOTIFS	HNMER_PFAM MOTIFS
Signature Sequences,	Domains and Motifs		PROTEIN DOMAIN SH3 KINASE GUANYLATE TRANSFERASE ATP-BINDING REPEAT GMP	MEMBRANE	PD001338: T403-E496	PROTEIN SH3 DOMAIN PERIPHERAL PLASMA	MEMBRANE CALMODULIN BINDING CASK CAMGUK	PD008238: M1-I139	PROTEIN MAGUK P55 SUBFAMI	LARGE HOMOLOG SH3 DOMAIN	PD152180: K230-R297	PROTEIN MAGUK P55 SUBFAMILY MEMBER MPP3	DISCS LARGE HOMOLOG SH3	PD090357: S318-T403	Guanylate kinase protein	BL00856: V400-I420, D42	SH3 domain signature	PR00452: R284-R296, M231-P241, A252-	<u>Q</u> 267	PDZ domain (Also known as	PDZ:	I139-G219	SH3 domain SH3:		M231-R296	M231-R296 Guanylate_Kinase:	M231-R296 Guanylate_Kinase: T403-I420
Potential	Glycosyla-	tion Sites	,																								
Potential	Phosphorylation																		_			,					
Amino	Acid	Residues																									
Incyte Amin	Polypeptide	ID																								477.7	700
SEQ	Ω	:OM	16						<u> </u>		=	·			×		· <u>··</u>					•			-		<u>-</u>

# Table 3 (cont.)

Analytical	Methods and Databases	BLAST_DOMO	BLAST_PRODOM	BLIMPS_PRINTS	HMMER_PFAM	MOTIFS	MOTIFS	PROFILESCAN	BLAST-DOMO	BLAST-PRODOM	BLAST-PRODOM	BLIMPS-BLOCKS	
Signature Sequences,	Domains and Motifs		SE ATP-BINDING PHOSPHORYLATION N PRECURSOR 44-A194 S209-	Tyrosine kinase catalytic domain signature PR00109: L126-V139, F162-L180, A228- D250, I270-L292	nase domain	Protein_Kinase_ATP L56-K79	Protein_Kinase_Serine/Threonine: 1168-L180	protein_kinase_tyrosine.profile: 18120-8201	XYLULOKINASE DM02388 P18157 1-492: F20-M498	GLYCEROL 3PHOSPHOTRANSFERASE GLYCEROKINASE GK PD001007: G239-A448	SIMILAR TO GLYCEROL KINASE PD130307: F20-K137	<pre>1y of carbohydrate kinase : F20-C43, Y54-P64, S159-N178,</pre>	G414-L429
Potential	Glycosyla- tion Sites								N131 N132 N178 N216				
Potential	Phosphorylation Sites	\$130 \$158 \$19 \$201 \$291 \$327 \$357 \$379 \$420 \$443 \$463 \$512 \$524 \$571 \$579							S117 S142 S152 S169 S232 S339	T333 T6 T9			
Amino	d idues	794							504				
Incyte	Polypeptide ID	7477486CD1				****			3773709CD1	,			
SEQ	OF ON	17							18				

Table 3 (cont.)

Analytical	Methods and Databases	PROFILESCAN	HMMER-PFAM	•	MOTIFS	BLAST-DOMO	BLAST-PRODOM	BLIMPS-PFAM	BLIMPS-PRINTS		CHICK COURT IN	BLIMPS-PRINTS	PROFILESCAN	HMMER-PFAM		HMMER-PFAM		MOTIFS	MOTIFS
Signature Sequences,	Domains and Motifs	FGGY family of carbohydrate kinases signatures prok_carb_kinases.prf: p350-K409	מַן	L92-R122, L172-D224, F238-A448	Fggy_Kinases_2: A366-E386	PROTEIN KINASE DOMAIN DM00004 P32298 157-401: F194-G440	RECEPTOR KINASE PD001932: K455-N531	Regulator of G-protein PF00615: F163-K179 V267-I280	Tyrosine kinase catalytic domain	PR00109: F419-S441, M268-Y281, H306-	1024, G324-1304, V3/2-1394	GPCR Kinase signature PR00717: Y172-Q184, K230-S248, P469- I486, V492-F505, N507-T524	Protein kinases signatures and profile protein_kinase_tyr.prf: R209-8345	Regulator of G protein signaling domain	RGS: N55-P78, P162-L176	Eukaryotic protein kinase domain	pkinase: F191-F454	Protein_Kinase_Atp: L197-K220	Protein_Kinase_St: I312-L324
Potential	Glycosyla- tion Sites					N418 N543													
Potential	Phosphorylation Sites					S187 S23 S36 S380 S399 S544	S58 T138 T139 T213 T348 T407	T537 T79 T85											
Amino	Acid Residues				Te me	553													
Incyte	Polypeptide ID					7477204CD1													
SEQ	9 <u>9</u>	118				119			-			-							

Table 3 (cont.)

SEQ	Incyte	Amino	Potential	Potential	Signature Sequences,	Analytical
ΩI	Polypeptide	Acid	Phosphorylation	Glycosyla-	Domains and Motifs	Methods and
NO:	i A	Residues	Sites	tion Sites		Databases
20	3016969CD1	871	21 S12 53 S16	N211	PROTEIN KINASE DOMAIN DM00004 S07571 5152-5396: Q580-P812	BLAST-DOMO
			93		Tyrosine PR00109: Y684-I702, T751-E773, I581-	BLIMPS-PRINTS
			S652 S665 S667 S727 S81 T172 T417 T516 T526		Eukaryotic protein kinase domain pkinase: F575-L827	HMMER-PFAM
···			9		Protein_Kinase_Tyr: 1690-1702	MOTIFS
21	063497CD1	765	S162 S181 S259 S286 S291 S410	N219 N289 N588 N618	Eukaryotic protein kinase domain:	HMMER_PFAM
		v <del> </del>	31 8437		Tyrosine kinase catalytic domain	BLIMPS_PRINTS
			S479 S495 S531 S539 S544 S550		signature PB00109:I.92-M105. V129-F147. V238-I.260	
-			69 S576			BLAST PRODOM
			39 \$646		PD127501:0346-D579	
			76 T172		PD070820:T715-I765, E642-G693, I345-	
			T365 T474 T478		P370	
			T50 T543 T622 T623 T684 T714		ZK524.4 PROTEIN SNF1RELATED KINASE PD156028:I282-I345	BLAST_PRODOM
			T716		KINASE TRANSFERASE ATP BINDING SERINE/	BLAST_PRODOM
					THREONINE PHOSPHORYLATION RECEPTOR	-
s					TYROSINE TRANSMEMBRANE	
_					PD000001:L18-V145, V238-W268, G168-	
					F215	
					PROTEIN KINASE DOMAIN	BLAST_DOMO
					DM00004 P27448 58-297:K20-L260	
					UMUUUU4   UCT440   20-201: UT0-UZ00	

Table 3 (cont.)

Analytical	Methods and Databases	MOTIFS	ctive- MOTIFS		HMMER_PFAM	file: PROFILESCAN		BLIMPS_PRINTS			DENT BLAST_PRODOM	IN		Т396-		BLAST_DOMO					MOTIFS	3.1 <del>11</del>		active-MOTIFS	
Signature Sequences,	Domains and Motifs	Protein kinases ATP-binding region signature:	Serine/Threonine protein kinases active-MOTIFS	site signature: V135-F147	Eukaryotic protein kinase domain: V14-V272	Protein kinases signatures and profile:	F85-E167	Tyrosine kinase catalytic domain	signature	PR00109:H126-L144	KINASE II CALCIUM/CALMODULIN DEPENDENT	SUBUNIT TRANSFERASE SERINE/THREONINE		001779:R456-V499, V272-S329,	A417	PROTEIN KINASE DOMAIN	DM00004 P11798 15-261:L16-A263	DM00004 JU0270 16-262:E18-A263	DM00004 A44412 16-262:E18-A263	DM00004 S57347 21-266:L20-T262	Protein kinases ATP-binding region	signature:	L20-K43	Serine/Threonine protein kinases ad	site signature:
Potential	Glycosyla- tion Sites			:	N313 N394 N407 N424																				
Potential	Phosphorylation Sites				S109 S355 S356 S36 S427 S433	S51 S557 S79	T262 T383 T408	T409 T410 T47	T488 T94			-	-						-						
Amino	Acid Residues				588																				
Incyte	Polypeptide ID				1625436CD1			-																	
SEQ	9 9 9	21			22			<b></b> -														····			

Table 3 (cont.)

Analytical	Methods and Databases		
Signature Sequences,	Domains and Motifs	F512-F785	
Potential	Glycosyla-	M1252 M1293 M1293	
Potential	Phosphorylation	0. \$130 \$1100 66 \$171 \$1205 00 \$234 \$1100 65 \$171 \$1205 00 \$204 \$1195 30 \$1214 \$1200 03 \$253 \$1214 \$1200 03 \$258 \$1225 00 \$236 \$1225 00 \$236 \$1225 00 \$236 \$1225 00 \$236 \$1250 \$236 \$1250 \$236 \$1	T1765
Amino	Acid	o n -	
	Polypeptide Th	T T T T T T T T T T T T T T T T T T T	
SEQ	0 C		

Table 3 (cont.)

Analytical	Methods and	Databases	HMMER_PFAM		profile: PROFILESCAN		BLIMPS_PRINTS		706-		SPECIFIC BLAST_PRODOM					IN4 BLAST_PRODOM	CIFIC		BLAST_DOMO		8,		9		8,		active-MOTIFS	
Signature Sequences,	Domains and Motifs		PDZ domain:	P1104-L1191	Protein kinases signatures and pr	F579-M659	Tyrosine kinase catalytic domain	signature	PR00109:M589-K602, Y625-I643, V706-	D728	MICROTUBULE ASSOCIATED TESTIS SPE	SERINE/THREONINE PROTEIN KINASE	PD142315:H1313-T1798	PD135564:V61-Y320, L1151-P1363	PD182663:E863-H1139	PROTEIN KINASE SERINE/THREONINE KIN4	MICROTUBULE ASSOCIATED TESTIS SPECIFIC	PD041650: K321-D511	PROTEIN KINASE DOMAIN	DM00004 A54602 455-712: T514-G772	DM08046 P05986 1-397: S508-K658,	V685-E829, D268-P291	DM00004 S42867 75-498: I515-T666,	H672-F813	DM00004 S42864 41-325: E513-K658,	H672-T773	e protein kinases	site signature:
Potential		tion Sites					-			, ,									, ,									
Potential	Phosphorylation	Sites																										
Amino	Acid	Residues												•						-								
Incyte	Polypeptide Acid	Ωī																										
SEQ	A	NO:	23			· · · · · · · · · · · · · · · · · · ·													·-						_	•		

Table 3 (cont.)

SEQ	Incyte	Amino	Potential	Potential	Signature Sequences,	Analytical
	Polypeptide Acid	Acid	Phosphorylation	Glycosyla-	Domains and Motifs	Methods and
NO:	ID	Residues	Sites	tion Sites		Databases
	3562763CD1	362	<b>ω</b> 1	N110 N165	ne domain:	HMMER
			S325 S81 T164		A263-D283	
			T197 T260 T280		Eukaryotic protein kinase domain:	HMMER_PFAM
	-		T286 T324 T353		Y30-L351	
					Protein kinases signatures and profile:	PROFILESCAN
					Tyrosine kinase catalytic domain	BLIMPS_PRINTS
					signature	
					PR00109: M143-L156, F178-I196, M326-	
					A348	
					PROTEIN KINASE DOMAIN	BLAST_DOMO
					DM00004 Q02723 16-259: K111-V215,	
					N232-V304	
					DM00004 A54602 455-712:N110-L316, I36-	
			•		DM00004 P23573 10-277: L139-K214,	
					E35-L102, F248-A348	
					DM00004[A57459 417-662:Y138-S325, E35-	
					L73	
					Protein kinases ATP-binding region	MOTIFS
					signature:	
					I36-K59	
					Serine/Threonine protein kinases active-MOTIFS	MOTIFS
					site signature:	
					I184-I196	

Table 3 (cont.)

SEQ	Incyte	Amino	Potential	Potential	Potential Signature Sequences,	Analytical
1		Acid	Phosphorylation	Glycosyla-	Phosphorylation Glycosyla- Domains and Motifs	Methods and
NO:	CI CI	Residues Sites	Sites	tion Sites		Databases
25	621293CD1	275			Adenylate kinase:	HMMER_PFAM
	······································				L69-P205	
					Adenylate kinase proteins.	BLIMPS_BLOCKS
	_				BL00113:L68-L84, N92-R135, C141-L155	
					Adenylate kinase signature	BLIMPS_PRINTS
					PR00094:L68-A81, G96-G110, W146-N162	
26	7480774CD1 660	099	S104 S106 S167	N177	INOSITOL 3 KINASE 1D MYOINOSITOL	BLAST_PRODOM
			S199 S226 S325		TRISPHOSPHATE 5 TRISPHOSPHATE IP3K IP3	
•			S338 S339 S343		TRANSFERASE KINASE CALMODULIN BINDING	
******			S355 S381 S458		PD010031:Q446-Q659, P377-Q442	
			S46 S629 S96		CALMODULIN-BINDING DOMAIN	BLAST_DOMO
			T117 T151 T160		DM07435   P42335   210-672:E315-Q659	
			T183 T210 T468		DM07435   P23677   1-461:G261-Q659	
			T500 T83 T90 T99			

### Table 4

Polynucleotide SEQ ID NO:	Incyte Polynucleotide ID	Sequence Length	Selected Fragment(s)	Sequence Fragments	5' Position	3' Position
27	2011384CB1	822	282-377	6829315H1 (SINTNOR01)	44	743
				g2954208	1	282
28	2004888CB1	1376	1349-1376, 499- 635	5545302T6 (TESTNOC01)	713	1376
				674588R6 (CRBLNOT01)	517	1256
				5562195F8 (BRSTDIT01)	Н	644
29.	2258952CB1	3468	1-983, 1461- 1908, 3369-3468	3219989H1 (COLNNON03)	3223	3468
				2258952T6 (OVARTUT01)	2757	3353
				FL2258952_g7458755_ 000012_g3766209	33	2849
				7126256H1 (COLNDIY01)	2527	3076
				g1633937	2718	3385
				7677920H1 (NOSETUE01)	-1	601
30	7473244CB1	2831	1-243, 834-1782	2660853T6 (LUNGTUT09)	2249	2831
				5216205F6 (BRSTNOT35)	1789	2681
				6854507F8 (BRAIFEN08)	763	1471
	- The state of the			55057226H1	354	1145
				5911008F6 (BRAIFEN05)	1299	1988
				2074751F6 (ISLTNOT01)	1626	2118
				688153571 (BRAHTDR03)	7	582
31	1242491CB1	2693	1-317, 2569-2693	70006068D1	1296	1838
				7934296H1 (COLADISO2)	2109	2693
				70003021D1	1740	2337
				7226035H1 (LUNGTMC01)	725	1187

Polynucleotide SEO ID NO:	Incyte Polynucleotide ID	Sequence Length	Selected Fragment(s)	Sequence Fragments	5' Position	3' Position
				5755513H1 (LUNGNOT35)	672	1102
1				70004229D1	1874	2338
32	2634875CB1	2973	1-1353, 2203-	2505294/HI 4009430F6 (MTSCNOT10)	959	1432
			9	5168601H1 (MUSCDMT01)	1691	1965
				5672440H1 (MUSLTDT01)	2213	2414
				6903523H1 (MUSLTDR02)	1833	2344
				55052146J1	1475	1654
		<del></del>		6217472F6 (MUSCDIT06)	2263	2973
				3585116F6 (293TF4T01)	623	1126
				GBI.g7242443_000006 .edit	1059	1585
	•			55052619J1	1	807
				2634875H1 (BONTNOT01)	1521	1764
33	3951059CB1	2066	532-772, 1830- 1886, 1966-2066	6882814J1 (BRAHTDR03)	1489	2066
				55058330J1	396	1316
				FL452484_00001	-	970
				71179403V1	1052	1745
34	7395890CB1	3975	1-326, 3951- 3975, 2980-3355,	6771964H1 (BRAUNOR01)	715	1432
			3666-3731, 1813- 2074, 1066-1098	6770122H1 (BRAUNOR01)	1471	2040
		<u> </u>		6771964J1 (BRAUNOR01)	2028	2713
			* * * * * * * * * * * * * * * * * * *	7393659H1 (BRABDIE02)	186	799
				55052405H1	1	218
			q	2570554R6 (HIPOAZT01)	2495	3012
				7660364H1 (OVARNOE02)	1861	2459
		-		FL034583_00001	2778	3584

Table 4 (cont.)

ion 3' Position	968	3162	1481	3975	1999	1287	1107	1918	580	771	282	1689	922	1476	1054	633	3360	3032	2379	1632	1346	3142	2126	200
5' Position	256	2715	968	3181	1327	655	497	1254	r-1	216	1	1077	558	820	604	1	2797	2345	1719	1036	783	2494	1597	
Sequence Fragments	7395271H1 (BRABDIE02)	6200064H1 (PITUNON01)	7395911H1 (BRABDIE02)	GNN.g8439948_000007 .edit2.comp	6873077H1 (BRAGNON02)	6623984J1 (UTRMTMR02)	7192851H2 (BRATDIC01)	6810083J1 (SKIRNOR01)	7013748H1 (KIDNNOC01)	7190770H1 (BRATDICO1)	55051332H1	6819441H1 (OVARDIR01)	7758313J1 (SPLNTUE01)	GNN:g807680_edit	1874092F6 (LEUKNOT02)	7315561H1 (SYNODIN02)	71224917V1	70858292V1	8045106H1 (OVARTUE01)	7617315J1 (KTDNTUE01)	7609838J1 (KTDCTME01)	70856122V1	71225608V1	5505385641
Selected Fragment(s)	, i .		,			1-46, 658-1061				1-66		4			1-30		1-172, 2484-	2523, 650-1457,	2247-2417	,		-		
Sequence Length						1918				1689					1054		3360							
Incyte Polynucleotide ID						7475546CB1				7477076CB1					1874092CB1		4841542CB1							
Polynucleotide SEO ID NO:						35				36			occ a.		37		38							

7472695CB1 2240 7477966CB1 3340 7163416CB1 2539	Polynucleotide SEO ID NO:	Incyte Polynucleotide ID	Sequence	Selected Fragment(s)	Sequence Fragments	5' Position	3' Position
7477966CB1 3340 1-980, 1504- 1710, 3315-3340 1-63416CB1 2539 1-228, 913-1225, 1994-2539 7472822CB1 2377 2341-2377, 1093- 1463, 1625-2081	39	7472695CB1	2240	1-20, 101-131, 704-1001	7191541F6 (BRATDIC01)	1	906
7477966CB1 3340 1-980, 1504- 1710, 3315-3340 1-228, 913-1225, 1994-2539 1-228, 913-1225, 1994-2539 1-228, 1994-2539 1994-2539					71872279V1	911	1501
7477966CB1 3340 1-980, 1504- 1710, 3315-3340 1-228, 913-1225, 1934-2539 1-228, 913-1225, 1994-2539 1463, 1625-2081					4211726T8 (BRONDIT01)	1466	2181
7477966CB1 3340 1-980, 1504- 1710, 3315-3340 7163416CB1 2539 1-228, 913-1225, 1994-2539 7472822CB1 2377 2341-2377, 1093- 1463, 1625-2081				* .	71870527V1	1717	2240
7477966CB1 3340 1-980, 1504- 1710, 3315-3340 7163416CB1 2539 1-228, 913-1225, 1994-2539 7472822CB1 2377 2341-2377, 1093-					71870095V1	699	1374
7477966CB1 3340 1-980, 1504- 1710, 3315-3340 7163416CB1 2539 1-228, 913-1225, 1994-2539 7472822CB1 2377 2341-2377, 1093- 1463, 1625-2081					2013786T6 (TESTNOT03)	1551	2217
7163416CB1 2539 1-228, 913-1225, 1994-2539 1994-2539 7472822CB1 2377 2341-2377, 1093-1463, 1625-2081	40	7477966CB1	3340		1513994T6 (PANCTUT01)	2768	3340
7163416CB1 2539 1-228, 913-1225, 1994-2539 1994-2539 7472822CB1 2377 2341-2377, 1093-1463, 1625-2081					6802962H1 (COLENOR03)	2241	2824
7163416CB1 2539 1-228, 913-1225, 1994-2539 1994-2539 7472822CB1 2377 2341-2377, 1093-1463, 1625-2081					55052773H1	1376	2254
7163416CB1 2539 1-228, 913-1225, 1994-2539 1994-2580 1994-2580 1994-2580 1994-2580 1994-2580 1994-2580 1994-2580 1994-2580 1994-2580 1994-2580 1994-2580 1994-2580 1994-2580 1994-2580 199					1513994F6 (PANCTUT01)	2155	2776
7163416CB1 2539 1-228, 913-1225, 1994-2539 1994-2377 1994-25577 1994-2577 1994-2577 1994-2577 1994-25					55052765H1	894	1745
7163416CB1 2539 1-228, 913-1225, 1994-2539 1994-2539 7472822CB1 2377 2341-2377, 1093-1463, 1625-2081					7607337J1 (COLRTUE01)	594	1258
7163416CB1 2539 1-228, 913-1225, 1994-2539 1994-2558 1994-2588 1994-2588 1994-2588 1994-2588 1994-2588 1994-2588 1994-2588 1994-2588 1994-2588 1994-2588 1994-2588 1994-2588 1994-2588 1994-2588 1994-2588 1994-2588 1994-2588 1994-2588 1994-2588 199					6802518H1 (COLENOR03)	551	858
7163416CB1 2539 1-228, 913-1225, 1994-2539 1994-2585 1994-2585 1994-2585 1994-2585 1994-2585 1994-2585 1994-2585 1994-2585 1994-2585 1994-2585 1994-2585 1994-2585 1994-2585 1994-2585 1994-2585 1994-2585 199					7677920H1 (NOSETUE01)	₩.	598
7472822CB1 2377 2341-2377, 1093-	41	7163416CB1	2539	1-228, 913-1225, 1994-2539	7715351J1 (SINTFEE02)	<del></del> 1	649
7472822CB1 2377 2341-2377, 1093-					1625532H1 (COLNPOT01)	1779	1993
7472822CB1 2377 2341-2377, 1093-					7163416F8 (PLACNOR01)	1888	2539
7472822CB1 2377 2341-2377, 1093-					7701682J1 (PENHTUE02)	815	1434
7472822CB1 2377 2341-2377, 1093-					7715351H1 (SINTFEE02)	399	1037
7472822CB1 2377 2341-2377, 1093-				***	7077243H1 (BRAUTDR04)	1306	1979
	42	7472822CB1	2377	2341-2377, 1093-	71982976V1	913	1546
71986606V1 55052941J1 71983943V1				1463, 1625-2081	71983661V1	793	1520
55052941J1 71983943V1					71986606V1	1494	2168
7100226					55052941J1	1551	2193
T/00005T/					71983660V1	1642	2377

Polynucleotide SEQ ID NO:	Incyte Polynucleotide ID	Sequence Length	Selected Fragment(s)	Sequence Fragments	5' Position	3' Position
43	7477486CB1	2897	2698-2763, 1- 365, 2314-2623,	4029722F8 (BRAINOT23)	2042	2584
			1516-1614, 2804- 2897	6910737R6 (PITUDIR01)	462	1370
				7237528H1 (BRAINOY02)	2348	2897
				7674962H2 (NOSETUE01)	125	589
				71982594V1	1386	1991
				6629715R6 (HEALDIR01)	637	1476
				GNN.g6165121_004.ed it	T	506
			,	6950253H1 (BRAITDR02)	1480	2176
44	3773709CB1	3361	1-168, 1479- 1982, 3336-3361	6938382F6 (FTUBTUR01)	116	850
				4383108H1 (BRAVUTT02)	Ţ	257
				7365206H1 (OVARDIC01)	2019	2580
				55024481H1 (PKINDNV08)	791	1462
				4119492H1 (BRSTTUT25)	3104	3361
				70783206V1	1969	2579
. 2000				3432983T6 (SKINNOT04)	2555	3217
				70782455V1	1361	2005
				70143324V1	2631	3219
			- 1		1463	2006
45	7477204CB1	1662	854-1662, 1-807	GNN.g8139716_edit		1662
46	3016969CB1	3225	1-916, 1154-	71873834V1	1555	2122
May a gran			1362, 3144-3225	5751549F8 (LUNGNOT35)	2153	2740
				7718401J1 (SINTFEE02)	1341	2100
				7354408H1 (HEARNON03)	2779	3225
			J	71872969V1	1969	2707
				71875134V1	885	1440

Polynucleotide SEO ID NO:	Incyte Polynucleotide ID	Sequence Length	Selected Fragment(s)	Sequence Fragments	5' Position	3' Position
i .			ÿ	3016969T6 (MUSCNOT07)	2532	3211
				6200811F6 (PITUNON01)	808	1403
		*****		55052669H1	T	852
	063497CB1	4772	1-431, 4420- 4540, 2098-2130,	6581829H1 (HEACDIC01)	2823	3464
***************************************			3522-3599, 2875- 3036	7199634H1 (LUNGFER04)	602	1153
				6936880H1 (FTUBTUR01)	3000	3714
				1449223H1 (PLACNOT02)	4029	4248
				4787168H1 (BRATNOT03)	3705	3964
				7714789H1 (SINTFEE02)	1198	1849
				7714789J1 (SINTFEE02)	4189	4772
				063497H1 (PLACNOB01)	1661	1880
				8025257J1 (ENDMUNE01)	<b>←</b> 1	702
				7381417H1 (ENDMUNE01)	1790	2359
				4351289H1 (CONFIMT01)	3884	4222
	٠			5068175H1 (PANCNOT23)	3675	3946
				7380657H1 (ENDMUNE01)	772	1305
				4051307H1 (SINTNOT18)	2689	2972
				7627517J1 (GBLADIE01)	2393	2919
				7629590H1 (GBLADIE01)	1953	2559
48	1625436CB1	1880	948-1167	5772228H1 (BRAINOT20)	844	1420
				7228 <u>5173V1</u>	673	1148

Table 4 (cont.)

Polynucleotide SEQ ID NO:	Incyte Polynucleotide ID	Sequence Length	Selected Fragment(s)	Sequence Fragments	5' Position	3' Position
48				7353062H1 (HEARNON03)	₩.	610
				7154515H1 (BRAMNOA01)	1164	1839
-				6764194H1 (BRAUNOR01)	1370	1880
:				72284772V1	491	1135
49	3330646CB1	5747	1-1738, 2291- 2733, 3677-4763	8178538H2 (EYERNON01)	5053	5722
				7218734H1 (COLNTMC01)	4882	5570
				8013776H1 (HEARNOC04)	4245	4904
				8006864H1 (PENIFEC01)	442	1064
				7711762H2 (TESTTUE02)	889	1292
				55124907H1	1301	2151
				8009629H1 (NOSEDIC02)	3681	4314
				7054991H1 (BRALINON02)	5099	5747
				55124907J1	1250	2101
				8267426H1 (MIXDUNE03)	2739	3511
No.				8054655J1 (ESOGTUE01)	2905	3529
				7930953H1 (COLNDIS02)	4339	4966
				7978939H1 (LSUBDMC01)	<b>L</b> I	504
	٠			7719236J1 (SINTFEE02)	2085	2746
				60215898V1	2234	2776
			-	6779321J1 (OVARDIR01)	3439	4230
50	3562763CB1	3418	1564-1627, 1-	55053205H1	523	1210
			376, 975-1073, 3066-3418	7321924H1 (NOSETUE01)	1843	2392
				7278180H1 (BMARTXE01)	2873	3418

Polynucleotide	Polynucleotide   Incyte	Sequence	Selected	Sequence Fragments	5' Position 3' Position	3' Position
SEQ ID NO:	Polynucleotide ID	Length	Fragment(s)			
50				400518R6	873	1430
				(PITUNOT02)		
				6816641J1	1297	1981
				(ADRETUR01)		
***				g2963935	1	383
				55143790J1	2257	3143
				55067380J2	314	579
				55143774J1	2577	3148
51	621293CB1	995	1-372, 410-468	72335268V1	<b>←</b> 1	508
	!			71870548V1	477	994
52	7480774CB1	2459	1664-2459, 1-110	71440281V1	685	1345
- Table 1				71438714V1	652	1226
		-		7082565H1		688
				(STOMTMR02)		
				71432228V1	1798	2459
				71431941V1	1257	1972
				6472388H1	1352	1985
				(PLACFEB01)		

Table 5

Polynucleotide	Incyte	Representative Library
SEQ ID NO:	Project ID	
2.7	2011384CB1	SINTNOR01
28	2004888CB1	TESTNOT03
29	2258952CB1	COLENOR03
30	7473244CB1	ISL'INOT01
31	1242491CB1	LUNGNOT02
32	2634875CB1	MUSCNOT07
33	3951059CB1	DRGCNOT01
34	7395890CB1	BRABDIE02
35	7475546CB1	CORPNOT02
36	7477076CB1	BRATDIC01
37	1874092CB1	LEUKNOT02
38	4841542CB1	KIDNNOT05
39	7472695CB1	TESTNOT03
40	7477966CB1	COLENOR03
41	7163416CB1	ESOGTME01
77	7472822CB1	BRABDIR03
43	7477486CB1	BRAITDR03
<b>ተ</b> ቸ	3773709CB1	SINTNOR01
46	3016969CB1	COLNNOT41
47	063497CB1	ENDMUNE 0.1
48	1625436CB1	BRACNOK02
49	3330646CB1	HNT2AGT01
20	3562763CB1	BRAHNOE01
51	621293CB1	KIDNNOT09
52	7480774CB1	BLADTUT02

### Table (

-	1.6	
Library	Vector	rary Describtion
BLADTUTUZ	PLNCY	Library was constructed using kNA isolated from bladder tumor tissue removed from an 80-year-old Caucasian female during a radical cystectomy and lymph node excision. Pathology indicated grade 3 invasive transitional cell carcinoma. Family history included acute renal failure, osteoarthritis, and atherosclerosis.
BRABDIE02	pincy	This 5' biased random primed library was constructed using RNA isolated from diseased cerebellum tissue removed from the brain of a 57-year-old Caucasian male who died from a cerebrovascular accident. Serologies were negative. Patient history included Huntington's disease, emphysema, and tobacco abuse (3-4 packs per day, for 40 years).
BRABDIR03	pincy	Library was constructed using RNA isolated from diseased cerebellum tissue removed from the brain of a 57-year-old Caucasian male who died from a cerebrovascular accident. Serologies were negative. Patient history included Huntington's disease, emphysema, and tobacco abuse (3-4 packs per day for 40 years).
BRACNOK02	PSPORT1	s amplified and normalized library was constructed using RNA isolate terior cingulate tissue removed from an 85-year-old Caucasian female m myocardial infarction and retroperitoneal hemorrhage. Pathology in erosclerosis, moderate to severe, involving the circle of Willis, mi ebral, basilar and vertebral arteries; infarction, remote, left dent leus; and amyloid plaque deposition consistent with age. There was merate leptomeningeal fibrosis, especially over the convexity of the There was mild generalized atrophy involving all lobes. The white the convexity of the seriest bightly reduced. The substantia nigra pars compacta appeasingmented. Patient history included COPD, hypertension, and recurrent ous thrombosis. 6.4 million independent clones from this amplified normalized in one round using conditions adapted Soares et al., PN 9228-9232 and Bonaldo et al., Genome Research 6 (1996):791.  Frary was constructed RNA isolated from posterior hippocampus tissue an a 45-year-old Caucasian female who died from a dissecting aortic a hemic bowel disease. Pathology indicated mild arteriosclerosis involubral cortical white matter and abasal ganglia. Grossly, there was mildigal librosis and mild focal atherosclerotic plaque in the middle ery, as well as vertebral arteries bilaterally. Microscopically, the ispheres, brain stem and cerebellum reveal focal areas in the white rephages in the Virchow-Robin space. In addition, there were scatter rephages in the Virchow-Robin space. In addition, there were scatter
		neurofibrillary tangles within the basolateral nuclei of the amygdala. Patient

Library	Vector	Library Description
		history included mild atheromatosis of aorta and coronary arteries, bowel and liver infarct due to aneurysm, physiologic fatty liver associated with obesity, mild diffuse emphysema, thrombosis of mesenteric and portal veins, cardiomegaly due to hypertrophy of left ventricle, arterial hypertension, acute pulmonary edema, splenomegaly, obesity (300 lb.), leiomyoma of uterus, sleep apnea, and iron deficiency anemia.
BRAITDR03	PCDNA2.1	This random primed library was constructed using RNA isolated from allocortex, cingulate posterior tissue removed from a 55-year-old Caucasian female who died from cholangiocarcinoma. Pathology indicated mild meningeal fibrosis predominately over the convexities, scattered axonal spheroids in the white matter of the cingulate cortex and the thalamus, and a few scattered neurofibrillary tangles in the entorhinal cortex and the periaqueductal gray region. Pathology for the associated tumor tissue indicated well-differentiated cholangiocarcinoma of the liver with residual or relapsed tumor. Patient history included cholangiocarcinoma, post-operative Budd-Chiari syndrome, biliary ascites, hydorthorax, dehydration, malnutrition, oliguria and acute renal failure. Previous surgeries included cholecystectomy and resection of 85% of the liver.
BRATDIC01	pINCY	This large size-fractionated library was constructed using RNA isolated from diseased brain tissue removed from the left temporal lobe of a 27-year-old Caucasian male during a brain lobectomy. Pathology for the left temporal lobe, including the mesial temporal structures, indicated focal, marked pyramidal cell loss and gliosis in hippocampal sector CA1, consistent with mesial temporal sclerosis. The left frontal lobe showed a focal deep white matter lesion, characterized by marked gliosis, calcifications, and hemosiderin-laden macrophages, consistent with a remote perinatal injury. The frontal lobe tissue also showed mild to moderate generalized gliosis, predominantly subpial and subcortical, consistent with chronic seizure disorder. GRAP was positive for astrocytes. The patient presented with intractable epilepsy, focal epilepsy, hemiplegia, and an unspecified brain injury. Patient history included cerebral palsy, abnormality of gait, depressive disorder, and tobacco abuse in remission. Previous surgeries included tendon transfer. Patient medications included minocycline hydrochloride, Tegretol, phenobarbital, vitamin C, Pepcid, and Pevaryl. Family history included brain cancer in the father.
COLENOR03	PCDNA2.1	Library was constructed using RNA isolated from colon epithelium tissue removed from a 13-year-old Caucasian female who died from a motor vehicle accident.
COLNNOT41	pincy	Library was constructed using RNA isolated from colon tissue removed from a 37-year-old female during a partial gastrojejunectomy. Pathology indicated a portion

Library	Vector	Library Description
1		of stomach and jejunum with an intact anastomotic site. The stomach showed a mild chronic gastritis without helicobacter pylori organisms. Normal appearing submucosal and myenteric plexus ganglion cells were noted. The jejunum had no significant abnormality.
CORPNOT02	pincy	Library was constructed using RNA isolated from diseased corpus callosum tissue removed from the brain of a 74-year-old Caucasian male who died from Alzheimer's disease.
DRGCNOT 01	pincy	Library was constructed using RNA isolated from dorsal root ganglion tissue removed from the cervical spine of a 32-year-old Caucasian male who died from acute pulmonary edema and bronchopneumonia, bilateral pleural and pericardial effusions, and malignant lymphoma (natural killer cell type). Patient history included probable cytomegalovirus, infection, hepatic congestion and steatosis, splenomegaly, hemorrhagic cystitis, thyroid hemorrhage, and Bell's palsy. Surgeries included colonoscopy, large intestine biopsy, adenotonsillectomy, and nasopharyngeal endoscopy and biopsy; treatment included radiation therapy.
ENDMUNE 0.1	PINCY	This 5' biased random primed library was constructed using RNA isolated from untreated umbilical artery endothelial cell tissue removed from a Caucasian male (Clonetics) newborn.
ESOGTME01	PSPORT1	This 5' biased random primed library was constructed using RNA isolated from esophageal tissue removed from a 53-year-old Caucasian male during a partial esophageal tissue removed from a 53-year-old Caucasian male during a partial esophagectomy, proximal gastrectomy, and regional lymph node biopsy. Pathology indicated no significant abnormality in the non-neoplastic esophagus. Pathology for the matched tumor tissue indicated invasive grade 4 (of 4) adenocarcinoma, forming a sessile mass situated in the lower esophagus, 2 cm from the quastroesophageal junction and 7 cm from the proximal margin. The tumor invaded through the muscularis propria into the adventitial soft tissue. Metastatic carcinoma was identified in 2 of 5 paragastric lymph nodes with perinodal extension. The patient presented with dysphagia. Patient history included membranous nephritis, hyperlipidemia, benign hypertension, and anxiety state. Previous surgeries included an adenotonsillectomy, appendectomy, and inguinal hernia repair. The patient was not taking any medications. Family history included at abdominal aortic aneurysm rupture in the father; breast cancer in the mother; a myocardial infarction and atherosclerotic coronary artery disease in the sibling(s); and myocardial infarction and atherosclerotic coronary artery disease in the grandparent(s).
HNT2AGT01	PBLUESCRIPT	Library was constructed at Stratagene (STR937233), using RNA isolated from the

Library	Vector	Library Description
		hNT2 cell line derived from a human teratocarcinoma that exhibited properties characteristic of a committed neuronal precursor. Cells were treated with retinoic
		acid for 5 weeks and with mitotic inhibitors for two weeks and allowed to mature
		for an additional 4 weeks in conditioned medium.
ISLTNOT01	pincy	Library was constructed using RNA isolated from a pooled collection of pancreatic   islet cells.
KIDNNOT05	PSPORT1	Library was constructed using RNA isolated from the kidney tissue of a 2-day-old
		Hispanic female, who died from cerebral anoxia. Family history included congenital heart disease.
KIDMNOT09	pINCY	Library was constructed using RNA isolated from the kidney tissue of a Caucasian
LEUKNOT02	DINCY	Library was constructed using RNA isolated from white blood cells of a 45-year-old
	ŧ	female with blood type 0+. The donor tested positive for cytomegalovirus (CMV).
LUNGNOT02	PBLUESCRIPT	Library was constructed using RNA isolated from the lung tissue of a 47-year-old
		Caucasian male, who died of a subarachnoid hemorrhage.
MUSCNOT07	DINCY	Library was constructed using RNA isolated from muscle tissue removed from the
	:	forearm of a 38-year-old Caucasian female during a soft tissue excision. Pathology
		for the associated tumor tissue indicated intramuscular hemangioma. Family history
		included breast cancer, benign hypertension, cerebrovascular disease, colon
		cancer, and type II diabetes.
SINTNORO1	PCDNA2.1	This random primed library was constructed using RNA isolated from small intestine
		tissue removed from a 31-year-old Caucasian female during Roux-en-Y gastric
		bypass. Patient history included clinical obesity.
TESTNOT03	PBLUESCRIPT	Library was constructed using RNA isolated from testicular tissue removed from a
		37-year-old Caucasian male, who died from liver disease. Patient history included
		cirrhosis, jaundice, and liver failure.

Program	Description	Reference	Parameter Threshold
ABIFACTURA	A program that removes vector sequences and masks ambiguous bases in nucleic acid sequences.	Applied Biosystems, Foster City, CA.	
ABIPARACEL FDF	A Fast Data Finder useful in comparing and annotating amino acid or nucleic acid sequences.	Applied Biosystems, Foster City, CA; Paracel Inc., Pasadena, CA.	Mismatch <50%
ABI AutoAssembler	A program that assembles nucleic acid sequences.	Applied Biosystems, Foster City, CA.	
BLAST	A Basic Local Alignment Search Tool useful in sequence similarity search for amino acid and nucleic acid sequences. BLAST includes five functions: blastp, blastn, blastx, tolastn, and tolastx.	Altschul, S.F. et al. (1990) J. Mol. Biol. 215:403-410; Altschul, S.F. et al. (1997) Nucleic Acids Res. 25:3389-3402.	ESTs: Probability value= 1.0E-8 or less Full Length sequences: Probability value= 1.0B-10 or less
FASTA	A Pearson and Lipman algorithm that searches for similarity between a query sequence and a group of sequences of the same type. FASTA comprises as least five functions: fasta, fasta, fastx, tfastx, and ssearch.	Pearson, W.R. and D.J. Lipman (1988) Proc. Natl. Acad Sci. USA 85:2444-2448; Pearson, W.R. (1990) Methods Enzymol. 183:63-98; and Smith, T.F. and M.S. Waterman (1981) Adv. Appl. Math. 2:482-489.	ESTs: fasta B value=1.06B-6 Assembled ESTs: fasta Identity= 95% or greater and Match length=200 bases or greater, fastx B value=1.0B-8 or less Full Length sequences: fastx score=100 or greater
· BLIMPS	A BLocks IMProved Searcher that matches a sequence against those in BLOCKS, PRINTS, DOMO, PRODOM, and PFAM databases to search for gene families, sequence homology, and structural fingerprint regions.	Henikoff, S. and J.G. Henikoff (1991) Nucleic Acids Res. 19:6565-6572; Henikoff, J.G. and S. Henikoff (1996) Methods Brzymol. 266:88-105; and Attwood, T.K. et al. (1997) J. Chem. Inf. Comput. Sci. 37:417-424.	Probability value= 1.0B-3 or less
HMMER	An algorithm for searching a query sequence against hidden Markov model (HMM)-based databases of protein family consensus sequences, such as PFAM.	Krogh, A. et al. (1994) J. Mol. Biol. 235:1501-1531; Somhammer, E.L.L. et al. (1988) Nucleic Acids Res. 26:320-322; Durbin, R. et al. (1998) Our World View, in a Nutshell, Cambridge Univ. Press, pp. 1-350.	PFAM hits: Probability value=1.0E-3 or less Signal peptide hits: Score=0 or greater

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Program	Description	Reference	Parameter Threshold
ProfileScan	An algorithm that searches for structural and sequence motifs in protein sequences that match sequence patterns defined in Prosite.	Gribskov, M. et al. (1988) CABIOS 4:61-66; Gribskov, M. et al. (1989) Methods Bnzymol. 183:146-159; Bairoch, A. et al. (1997) Nucleic Acids Res. 25:217-221.	Normalized quality scorez GCG-specified "HIGH" value for that particular Prosite motif. Generally, score=1.4-2.1.
Phred	A base-calling algorithm that examines automated sequencer traces with high sensitivity and probability.	Ewing, B. et al. (1998) Genome Res. 8:175-185; Ewing, B. and P. Green (1998) Genome Res. 8:186-194.	
Phrap	A Phils Revised Assembly Program including SWAT and CrossMatch, programs based on efficient implementation of the Smith-Waterman algorithm, useful in searching sequence homology and assembling DNA sequences.	Smith, T.F. and M.S. Waterman (1981) Adv. Appl. Math. 2:482-489; Smith, T.F. and M.S. Waterman (1981) J. Mol. Biol. 147:195-197; and Green, P., University of Washington, Seattle, WA.	Score= 120 or greater; Match length= 56 or greater
Consed	A graphical tool for viewing and editing Phrap assemblies.	Gordon, D. et al. (1998) Genome Res. 8:195-202.	
SPScan	A weight matrix analysis program that scans protein sequences for the presence of secretory signal peptides.	Nielson, H. et al. (1997) Protein Engineering 10:1-6; Claverie, J.M. and S. Audic (1997) CABIOS 12:431-439.	Score=3.5 or greater
TMAP	A program that uses weight matrices to delineate transmembrane segments on protein sequences and determine orientation.	Persson, B. and P. Argos (1994) J. Mol. Biol. 237:182-192; Persson, B. and P. Argos (1996) Protein Sci. 5;363-371.	
TMHMMER	A program that uses a hidden Markov model (HMM) to delineate transmembrane segments on protein sequences and determine orientation.	Sonnhammer, E.L. et al. (1998) Proc. Sixth Inti. Conf. on Intelligent Systems for Mol. Biol., Glasgow et al., eds., The Am. Assoc. for Artificial Intelligence Press, Menlo Park, CA, pp. 175-182.	
Motifs	A program that searches amino acid sequences for patterns that matched those defined in Prosite.	Bairoch, A. et al. (1997) Nucleic Acids Res. 25:217-221; Wisconsin Package Program Manual, version 9, page M51-59, Genetics Computer Group, Madison, WI.	7-221; age

What is claimed is:

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1. An isolated polypeptide selected from the group consisting of:

- a) a polypeptide comprising an amino acid sequence selected from the group consisting of
   SEQ ID NO:1-26,
  - b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-26,
  - c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, and
- d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-26.
  - 2. An isolated polypeptide of claim 1 selected from the group consisting of SEQ ID NO:1-26.

3. An isolated polynucleotide encoding a polypeptide of claim 1.

- 4. An isolated polynucleotide encoding a polypeptide of claim 2.
- 5. An isolated polynucleotide of claim 4 selected from the group consisting of SEQ ID NO:27-52.
  - 6. A recombinant polynucleotide comprising a promoter sequence operably linked to a polynucleotide of claim 3.

7. A cell transformed with a recombinant polynucleotide of claim 6.

- 8. A transgenic organism comprising a recombinant polynucleotide of claim 6.
- 9. A method for producing a polypeptide of claim 1, the method comprising:a) culturing a cell under conditions suitable for expression of the polypeptide, wherein said
  - cell is transformed with a recombinant polynucleotide, and said recombinant polynucleotide comprises a promoter sequence operably linked to a polynucleotide encoding the polypeptide of claim 1, and
- b) recovering the polypeptide so expressed.

- 10. An isolated antibody which specifically binds to a polypeptide of claim 1.
- 11. An isolated polynucleotide selected from the group consisting of:
- a) a polynucleotide comprising a polynucleotide sequence selected from the group consisting
   of SEQ ID NO:27-52,
  - b) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:27-52,
    - c) a polynucleotide complementary to a polynucleotide of a),
    - d) a polynucleotide complementary to a polynucleotide of b), and
- 10 e) an RNA equivalent of a)-d).
  - 12. An isolated polynucleotide comprising at least 60 contiguous nucleotides of a polynucleotide of claim 11.
- 13. A method for detecting a target polynucleotide in a sample, said target polynucleotide having a sequence of a polynucleotide of claim 11, the method comprising:
  - a) hybridizing the sample with a probe comprising at least 20 contiguous nucleotides comprising a sequence complementary to said target polynucleotide in the sample, and which probe specifically hybridizes to said target polynucleotide, under conditions whereby a hybridization complex is formed between said probe and said target polynucleotide or fragments thereof, and
  - b) detecting the presence or absence of said hybridization complex, and, optionally, if present, the amount thereof.
    - 14. A method of claim 13, wherein the probe comprises at least 60 contiguous nucleotides.

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- 15. A method for detecting a target polynucleotide in a sample, said target polynucleotide having a sequence of a polynucleotide of claim 11, the method comprising:
- a) amplifying said target polynucleotide or fragment thereof using polymerase chain reaction amplification, and
- b) detecting the presence or absence of said amplified target polynucleotide or fragment thereof, and, optionally, if present, the amount thereof.
  - 16. A composition comprising a polypeptide of claim 1 and a pharmaceutically acceptable excipient.

17. A composition of claim 16, wherein the polypeptide has an amino acid sequence selected from the group consisting of SEQ ID NO:1-26.

- 18. A method for treating a disease or condition associated with decreased expression of
   functional PKIN, comprising administering to a patient in need of such treatment the composition of claim 16.
  - 19. A method for screening a compound for effectiveness as an agonist of a polypeptide of claim 1, the method comprising:
    - a) exposing a sample comprising a polypeptide of claim 1 to a compound, and
    - b) detecting agonist activity in the sample.

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20. A composition comprising an agonist compound identified by a method of claim 19 and a pharmaceutically acceptable excipient.

21. A method for treating a disease or condition associated with decreased expression of functional PKIN, comprising administering to a patient in need of such treatment a composition of claim 20.

- 22. A method for screening a compound for effectiveness as an antagonist of a polypeptide of claim 1, the method comprising:
  - a) exposing a sample comprising a polypeptide of claim 1 to a compound, and
  - b) detecting antagonist activity in the sample.
- 23. A composition comprising an antagonist compound identified by a method of claim 22 and a pharmaceutically acceptable excipient.
  - 24. A method for treating a disease or condition associated with overexpression of functional PKIN, comprising administering to a patient in need of such treatment a composition of claim 23.
  - 25. A method of screening for a compound that specifically binds to the polypeptide of claim 1, said method comprising the steps of:
  - a) combining the polypeptide of claim 1 with at least one test compound under suitable conditions, and

b) detecting binding of the polypeptide of claim 1 to the test compound, thereby identifying a compound that specifically binds to the polypeptide of claim 1.

- 26. A method of screening for a compound that modulates the activity of the polypeptide of 5 claim 1, said method comprising:
  - a) combining the polypeptide of claim 1 with at least one test compound under conditions permissive for the activity of the polypeptide of claim 1,
    - b) assessing the activity of the polypeptide of claim 1 in the presence of the test compound, and
  - c) comparing the activity of the polypeptide of claim 1 in the presence of the test compound with the activity of the polypeptide of claim 1 in the absence of the test compound, wherein a change in the activity of the polypeptide of claim 1 in the presence of the test compound is indicative of a compound that modulates the activity of the polypeptide of claim 1.
- 27. A method for screening a compound for effectiveness in altering expression of a target polynucleotide, wherein said target polynucleotide comprises a sequence of claim 5, the method comprising:
  - a) exposing a sample comprising the target polynucleotide to a compound, under conditions suitable for the expression of the target polynucleotide,
    - b) detecting altered expression of the target polynucleotide, and

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- c) comparing the expression of the target polynucleotide in the presence of varying amounts of the compound and in the absence of the compound.
  - 28. A method for assessing toxicity of a test compound, said method comprising:
  - a) treating a biological sample containing nucleic acids with the test compound;
- b) hybridizing the nucleic acids of the treated biological sample with a probe comprising at least 20 contiguous nucleotides of a polynucleotide of claim 11 under conditions whereby a specific hybridization complex is formed between said probe and a target polynucleotide in the biological sample, said target polynucleotide comprising a polynucleotide sequence of a polynucleotide of claim 11 or fragment thereof;
  - c) quantifying the amount of hybridization complex; and
  - d) comparing the amount of hybridization complex in the treated biological sample with the amount of hybridization complex in an untreated biological sample, wherein a difference in the amount of hybridization complex in the treated biological sample is indicative of toxicity of the test compound.

29. A diagnostic test for a condition or disease associated with the expression of PKIN in a biological sample comprising the steps of:

- a) combining the biological sample with an antibody of claim 10, under conditions suitable
   for the antibody to bind the polypeptide and form an antibody:polypeptide complex; and
  - b) detecting the complex, wherein the presence of the complex correlates with the presence of the polypeptide in the biological sample.
    - 30. The antibody of claim 10, wherein the antibody is:
- a) a chimeric antibody,
  - b) a single chain antibody,
  - c) a Fab fragment,
  - d) a F(ab')<sub>2</sub> fragment, or
  - e) a humanized antibody.

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- 31. A composition comprising an antibody of claim 10 and an acceptable excipient.
- 32. A method of diagnosing a condition or disease associated with the expression of PKIN in a subject, comprising administering to said subject an effective amount of the composition of claim 31.
  - 33. A composition of claim 31, wherein the antibody is labeled.
- 34. A method of diagnosing a condition or disease associated with the expression of PKIN in
  a subject, comprising administering to said subject an effective amount of the composition of claim
  33.
  - 35. A method of preparing a polyclonal antibody with the specificity of the antibody of claim 10 comprising:
- a) immunizing an animal with a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, or an immunogenic fragment thereof, under conditions to elicit an antibody response;
  - b) isolating antibodies from said animal; and

c) screening the isolated antibodies with the polypeptide, thereby identifying a polyclonal antibody which binds specifically to a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-26.

- 5 36. An antibody produced by a method of claim 35.
  - 37. A composition comprising the antibody of claim 36 and a suitable carrier.
- 38. A method of making a monoclonal antibody with the specificity of the antibody of claim 10 comprising:
  - a) immunizing an animal with a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-26, or an immunogenic fragment thereof, under conditions to elicit an antibody response;
    - b) isolating antibody producing cells from the animal;
- c) fusing the antibody producing cells with immortalized cells to form monoclonal antibodyproducing hybridoma cells;
  - d) culturing the hybridoma cells; and

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- e) isolating from the culture monoclonal antibody which binds specifically to a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-26.
- 39. A monoclonal antibody produced by a method of claim 38.
  - 40. A composition comprising the antibody of claim 39 and a suitable carrier.
- 41. The antibody of claim 10, wherein the antibody is produced by screening a Fab expression library.
  - 42. The antibody of claim 10, wherein the antibody is produced by screening a recombinant immunoglobulin library.
  - 43. A method for detecting a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-26 in a sample, comprising the steps of:
  - a) incubating the antibody of claim 10 with a sample under conditions to allow specific binding of the antibody and the polypeptide; and

b) detecting specific binding, wherein specific binding indicates the presence of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-26 in the sample.

- 5 44. A method of purifying a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-26 from a sample, the method comprising:
  - a) incubating the antibody of claim 10 with a sample under conditions to allow specific binding of the antibody and the polypeptide; and
- b) separating the antibody from the sample and obtaining the purified polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-26.
  - 45. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:1.
  - 46. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:2.
  - 47. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:3.
  - 48. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:4.
- 20 49. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:5.

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- 50. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:6.
- 51. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:7.
- 52. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:8.
- 53. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:9.
- 30 54. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:10.
  - 55. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:11.
  - 56. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:12.

57. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:13. 58. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:14. 5 59. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:15. 60. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:16. 61. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:17. 10 62. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:18. 63. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:19. 15 64. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:20. 65. A polypeptide of claim 1, comprising the amino acid sequence of SEO ID NO:21. 66. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:22. 20 67. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:23. 68. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:24. 25 69. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:25. 70. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:26. 71. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID 30 NO:27. 72. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:28.

73. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:29.

- 74. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ IDNO:30.
  - 75. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:31.
- 10 76. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:32.
  - 77. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:33.
  - 78. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:34.

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- 79. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ IDNO:35.
  - 80. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:36.
- 25 81. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:37.
  - 82. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:38.
  - 83. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:39.

84. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:40.

- 85. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ IDNO:41.
  - $86.\,$  A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:42.
- 10 87. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:43.
  - 88. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:44.

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- 89. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:45.
- 90. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:46.
  - 91. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:47.
- 25 92. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:48.
  - 93. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:49.
  - 94. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:50.

95. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:51.

96. A polynucleotide of claim 11, comprising the polynucleotide sequence of SEQ ID NO:52.

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				Thr 350					355				•	360
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Ser	Gln	Trp	Leu	Leu 380	Ile	Gly	His	His		Leu	Pro	Pro	Val	
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Met	Thr	His	Ser	Pro 185	Thr	Asp	Pro	Arg	Pro 190	Ala	Lys	Ala	Glu	Glu 195
Gly	Lys	Asn	Ile	Leu 200	Ala	Glu	Ser	Gln	Lys 205	Glu	Val	Gly	Glu	Lys 210
Thr	Pro	Gly	Gln	Ala 215	Gly	Gln	Ala	Lys	Met 220	Gln	Gly	Asp	Thr	Ser 225
Arg	Gly	Ile	Glu		Gln	Ala	Val	Pro		Glu	Lys	Ser	Glu	
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Met Leu Val Asp Pro Ser Arg Gln Pro Tyr Arg Val Lys Val Ile
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Asp	Phe	Gly	Ser	Ala 350	Ser	His	Val	Ser	Lys 355	Ala	Val	Суз	Ser	Thr 360
Tyr	Leu	Gln	Ser	Arg	Tyr	Tyr	Arg	Ala		Glu	Ile	Ile	Leu	
				∙3 б5					370					375
ьeu ·	Pro	Pne	cys	Glu 380	Ата	TTE	Asp	Met	385	ser	ьeu	GIA	cys	390
Ile	Ala	Glu	Leu	Phe	Leu	Gly	Trp	Pro		Tyr	Pro	Gly	Ala	
G111	Ͳvr	Asp	Gln	395 Ile	Ara	Tvr	Tle	Ser	400 Gln	Thr	Gln	Glv	Leu	405 Pro
				410					415					420
Ala	G1u	Tyr	Leu	Leu 425	Ser	Ala	GIY	Thr	ьуя 430	Thr	ınr	Arg	Pne	Pne 435
Asn	Arg	Asp	Thr	Asp	Ser	Pro	Tyr	Pro		Trp	Arg	Leu	Lys	
Pro	Asp	Asp	His	440 Glu	Ala	Glu	Thr	Gly	445 Ile	Lys	Ser	Lys	Glu	450 Ala
7	T		<b>-</b> 1 -	455 Di-	70	<b>C</b>	T	7	460	3/	70.7 -	Q1 m	T 7 ~ 7	465
Arg	гуѕ	TYT	TTE	Phe 470	Asn	Суѕ	ьеи	Asp	475	мес	Ala	GIII	Val	480
Met	Thr	Thr	Asp	Leu 485	Glu	Gly	Ser	Asp	Met 490	Leu	Val	Glu	Lys	Ala 495
Asp	Arg	Arg	Glu	Phe 500	Ile	Asp	Leu	Leu	Lys 505	Lys	Met	Leu	Thr	Ile 510
Asp	Ala	Asp	Lys	Arg	Ile	Thr	Pro	Ile	Glu	Thr	Leu	Asn	His	Pro
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Val	Lys	Ser	CAz	Phe 545	Gln	Asn	Met	Glu	Ile 550	Суѕ	Lys	Arg	Arg	Val 555
Asn	Met	Туг	Asp	Thr 560	Val	Asn	Gln	Ser		Thr	Pro	Phe	Ile	
His	Val	Ala	Pro	Ser 575	Thr	Ser	Thr	Asn		Thr	Met	Thr	Phe	
Asn	Gln	Leu	Thr	Thr	Val	His	Asn	Gln	Pro 595	Ser	Ala	Ala	Ser	
Ala	Ala	Val	Ala	590 Gln 605	Arg	Ser	Met	Pro		Gln	Thr	Gly	Thr	600 Ala 615
Gln	Ile	Суѕ	Ala	Arg 620	Pro	Asp	Pro	Phe		Gln	Ala	Leu	Ile	
Cys	Pro	Pro	Gly	Phe 635	Gln	Gly	Leu	Gln		Ser	Pro	Ser	Lys	
Ala	Gly	Tyr	Ser	Val 650	Arg	Met	Glu	Asn		Val	Pro	Ile	Val	Thr 660
Gln	Ala	Pro	Gly	Ala 665	Gln	Pro	Leu	Gln		Gln	Pro	Gly	Leu	
Ala	Gln	Gln	Ala	Trp	Pro	Ser	Gly	Thr		Gln	Ile	Leu	Leu	Pro 690
Pro	Ala	Trp	Gln	680 Gln	Leu	Thr	Gly	Val	685 Ala	Thr	His	Thr	Ser	
Gln	His	Ala	Thr	695 Val	Ile	Pro	Glu	Thr	700 Met	Ala	Gly	Thr	Gln	705 Gln
				710					715					720
			-	Arg 725					730	_				735
Pro	Ile	Met	Gln	Gln 740	Pro	Ala	Leu	Leu	Thr 745	Gly	His	Val	Thr	Leu 750
Pro	Ala	Ala	Gln	Pro 755	Leu	Asn	Val	G1y	Val 760	Ala	Hìs	Va1	Met	Arg 765
Gln	Gln	Pro	Thr	Ser 770	Thr	Thr	Ser	Ser		Lys	Ser	Lys	Gln	
Gln	Ser	Ser	Val	Arg	Asn	Val	Ser	Thr		Glu	Val	Ser	Ser	
Gln	Ala	Ile	Ser	785 Ser	Pro	Gln	Arg	Ser	Lys	Arg	Val	Lys	Glu	Asn
Thr	Pro	Pro	Arg	800 Cys	Ala	Met	Val	His		Ser	Pro	Ala	Cys	
ጥኮኮ	Ser	Val	Thr	815 Cys	Glv	Trp	Glv	Asp	820 Val	Ala	Ser	Ser	Thr	825 Thr
	~			830	~-1	~-1	7		835					840

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Arg Glu Arg Gln Arg Gln Thr Ile Val Ile Pro Asp Thr Pro Ser
                845
                                     850
Pro Thr Val Ser Val Ile Thr Ile Ser Ser Asp Thr Asp Glu Glu
                 860
                                     865
                                                          870
Glu Glu Gln Lys His Ala Pro Thr Ser Thr Val Ser Lys Gln Arg
                875
                                     880
                                                          885
Lys Asn Val Ile Ser Cys Val Thr Val His Asp Ser Pro Tyr Ser
                890
                                     895
                                                          900
Asp Ser Ser Ser Asn Thr Ser Pro Tyr Ser Val Gln Gln Arg Ala
                905
                                     910
                                                          915
Gly His Asn Asn Ala Asn Ala Phe Asp Thr Lys Gly Ser Leu Glu
                920
                                     925
                                                          930
Asn His Cys Thr Gly Asn Pro Arg Thr Ile Ile Val Pro Pro Leu
                935
                                     940
Lys Thr Gln Ala Ser Glu Val Leu Val Glu Cys Asp Ser Leu Val
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                                     955
                                                          960
Pro Val Asn Thr Ser His His Ser Ser Ser Tyr Lys Ser Lys Ser
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                                     970
                                                          975
Ser Ser Asn Val Thr Ser Thr Ser Gly His Ser Ser Gly Ser Ser
                980
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                                                          990
Ser Gly Ala Ile Thr Tyr Arg Gln Gln Arg Pro Gly Pro His Phe
                995
                                    1000
                                                         1005
Gln Gln Gln Gln Pro Leu Asn Leu Ser Gln Ala Gln Gln His Ile
                                    1015
               1010
                                                         1020
Thr Thr Asp Arg Thr Gly Ser His Arg Arg Gln Gln Ala Tyr Ile
               1025
                                    1030
                                                         1035
Thr Pro Thr Met Ala Gln Ala Pro Tyr Ser Phe Pro His Asn Ser
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                                    1045
                                                         1050
Pro Ser His Gly Thr Val His Pro His Leu Ala Ala Ala Ala Ala
               1055
                                    1060
Ala Ala His Leu Pro Thr Gln Pro His Leu Tyr Thr Tyr Thr Ala
               1070
                                    1075
Pro Ala Ala Leu Gly Ser Thr Gly Thr Val Ala His Leu Val Ala
               1085
                                    1090
Ser Gln Gly Ser Ala Arg His Thr Val Gln His Thr Ala Tyr Pro
               1100
                                    1105
Ala Ser Ile Val His Gln Val Pro Val Ser Met Gly Pro Arg Val
               1115
                                    1120
                                                         1125
Leu Pro Ser Pro Thr Ile His Pro Ser Gln Tyr Pro Ala Gln Phe
               1130
                                    1135
Ala His Gln Thr Tyr Ile Ser Ala Ser Pro Ala Ser Thr Val Tyr
               1145
                                    1150
                                                         1155
Thr Gly Tyr Pro Leu Ser Pro Ala Lys Val Asn Gln Tyr Pro Tyr
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Leu Thr Glu Arg Leu Gly Ser Gly Thr Tyr
                                         Ala Thr Val Tyr Lys
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                                                           3.0
Ala Tyr Ala Lys Lys
                    Asp Thr Arg Glu Val Val Ala Ile Lys Cys
                  35
                                      40
                                                           45
Val Ala Lys Lys Ser Leu Asn Lys Ala Ser Val Glu Asn Leu Leu
                 50
                                      55
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Thr Glu Ile Glu Ile Leu Lys Gly Ile Arg His Pro His Ile Val
                                      70
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Gln Leu Lys Asp Phe Gln Trp Asp Ser Asp Asn Ile Tyr Leu Ile
Met Glu Phe Cys Ala Gly Gly Asp Leu Ser Arg Phe Ile His Thr
                                     100
                                                         105
Arg Arg Ile Leu Pro Glu Lys Val Ala Arg Val Phe Met Gln Gln
                110
                                     115
Leu Ala Ser Ala Leu Gln Phe Leu His Glu Arg Asn Ile Ser His
                125
                                     130
                                                         135
Leu Asp Leu Lys Pro Gln Asn Ile Leu Leu Ser Ser Leu Glu Lys
                140
                                     145
                                                         150
Pro His Leu Lys Leu Ala Asp Phe Gly Phe Ala Gln His Met Ser
                155
                                     160
                                                         165
Pro Trp Asp Glu Lys His Val Leu Arg Gly Ser Pro Leu Tyr Met
                170
                                     175
Ala Pro Glu Met Val Cys Gln Arg Gln Tyr Asp Ala Arg Val Asp
                185
                                     190
                                                         195
Leu Trp Ser Met Gly Val Ile Leu Tyr Glu Ala Leu Phe Gly Gln
                200
                                     205
                                                         210
Pro Pro Phe Ala Ser Arg Ser Phe Ser Glu Leu Glu Glu Lys Ile
                215
                                     220
                                                         225
Arg Ser Asn Arg Val Ile Glu Leu Pro Leu Arg Pro Leu Leu Ser
                230
                                     235
Arg Asp Cys Arg Asp Leu Leu Gln Arg Leu Leu Glu Arg Asp Pro
                245
                                     250
                                                         255
Ser Arg Arg Ile Ser Phe Gln Asp Phe Phe Ala His Pro Trp Val
                260
                                     265
                                                          270
Asp Leu Glu His Met Pro Ser Gly Glu Ser Leu Gly Arg Ala Thr
                275
                                     280
                                                         285
Ala Leu Val Val Gln Ala Val Lys Lys Asp Gln Glu Gly Asp Ser
                290
                                     295
Ala Ala Ala Leu Ser Leu Tyr Cys Lys Ala Leu Asp Phe Phe Val
                305
                                     310
                                                         315
Pro Ala Leu His Tyr Glu Val Asp Ala Gln Arg Lys Glu Ala Ile
                320
                                     325
                                                         330
Lys Ala Lys Val Gly Gln Tyr Val Ser Arg Ala Glu Glu Leu Lys
                335
                                     340
Ala Ile Val Ser Ser Ser Asn Gln Ala Leu Leu Arg Gln Gly Thr
                                     355
                                                          360
Ser Ala Arg Asp Leu Leu Arg Glu Met Ala Arg Asp Lys Pro Arg
                365
                                     370
                                                         375
Leu Leu Ala Ala Leu Glu Val Ala Ser Ala Ala Met Ala Lys Glu
                380
                                     385
                                                         390
Glu Ala Ala Gly Gly Glu Gln Asp Ala Leu Asp Leu Tyr Gln His
                395
                                     400
                                                         405
Ser Leu Gly Glu Leu Leu Leu Leu Ala Ala Glu Pro Pro Gly
                410
                                     415
                                                         420
Arg Arg Glu Leu Leu His Thr Glu Val Gln Asn Leu Met Ala
                425
                                     430
Arg Ala Glu Tyr Leu Lys Glu Gln Met Arg Glu Ser Arg Trp Glu
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                                     445
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Ala Asp Thr Leu Asp Lys Glu Gly Leu Ser Glu Ser Val Arg Ser
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Ser Cys Thr Leu Gln
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Ser Ser Ser Ser Gly Val Leu Met Val Gly Pro Asn Phe Arg
                 35
                                      40
Val Gly Lys Lys Ile Gly Cys Gly Asn Phe Gly Glu Leu Arg Leu
                 50
                                      55
Gly Lys Asn Leu Tyr Thr Asn Glu Tyr Val Ala Ile Lys Leu Glu
                 65
                                      70
Pro Ile Lys Ser Arg Ala Pro Gln Leu His Leu Glu Tyr Arg Phe
                 80
                                      85
                                                          90
Tyr Lys Gln Leu Gly Ser Ala Gly Glu Gly Leu Pro Gln Val
                                                         Tyr
                 95
                                     100
Tyr Phe Gly Pro Cys Gly Lys Tyr Asn Ala Met Val Leu Glu Leu
                110
                                     115
Leu Gly Pro Ser Leu Glu Asp Leu Phe Asp Leu Cys Asp Arg Thr
                125
                                     130
                                                         135
Phe Thr Leu Lys Thr Val Leu Met Ile Ala Ile Gln Leu Leu Ser
                140
                                     145
                                                         150
Arg Met Glu Tyr Val His Ser Lys Asn Leu Ile Tyr Arg Asp Val
                155
                                     160
                                                         165
Lys Pro Glu Asn Phe Leu Ile Gly Arg Gln Gly Asn Lys Lys Glu
                170
                                     175
His Val Ile His Ile Ile Asp Phe Gly Leu Ala Lys Glu Tyr Ile
                185
                                     190
                                                         195
Asp Pro Glu Thr Lys Lys His Ile Pro Tyr Arg Glu His Lys Ser
                200
                                     205
                                                         210
Leu Thr Gly Thr Ala Arg Tyr Met Ser Ile Asn Thr His Leu Gly
                215
                                     220
                                                         225
Lys Glu Gln Ser Arg Arg Asp Asp Leu Glu Ala Leu Gly His Met
                230
                                     235
Phe Met Tyr Phe Leu Arg Gly Ser Leu Pro Trp Gln Gly Leu Lys
                245
                                     250
                                                         255
Ala Asp Thr Leu Lys Glu Arg Tyr Gln Lys Ile Gly Asp Thr Lys
                260
                                     265
                                                         270
Arg Asn Thr Pro Ile Glu Ala Leu Cys Glu Asn Phe Pro Glu Glu
                275
                                     280
                                                         285
Met Ala Thr Tyr Leu Arg Tyr Val Arg Arg Leu Asp Phe Phe Glu
                290
                                     295
                                                         300
Lys Pro Asp Tyr Glu Tyr Leu Arg Thr Leu Phe Thr Asp Leu Phe
                305
                                     310
                                                         315
Glu Lys Lys Gly Tyr Thr Phe Asp Tyr Ala Tyr Asp Trp Val Gly
                320
                                     325
                                                         330
Arg Pro Ile Pro Thr Pro Val Gly Ser Val His Val Asp Ser Gly
                335
                                     340
                                                         345
Ala Ser Ala Ile Thr Arg Glu Ser His Thr His Arg Asp Arg Pro
                350
                                     355
Ser Gln Gln Pro Leu Arg Asn Gln Val Val Ser Ser Thr Asn
                365
                                     370
Gly Glu Leu Asn Val Asp Asp Pro Thr Gly Ala His Ser Asn Ala
                380
                                     385
                                                         390
Pro Ile Thr Ala His Ala Glu Val Glu Val Glu Glu Ala Lys
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                                     400
Cys Cys Cys Phe Phe Lys Arg Lys Arg Lys Thr Ala Gln Arg
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His Lys
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<sup>&</sup>lt;223> Incyte ID No: 1874092CD1

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Gly Trp Asp Arg Lys Ala Gln Arg Asn Gly Leu Arg Ser Gln Val
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                                                           3.0
Tyr Ala Val Asn Gly Asp Tyr Tyr Val Gly Glu Trp Lys Asp Asn
                 35
                                      40
                                                           45
Val Lys His Gly Lys Gly Thr Gln Val Trp
                                        Lys Lys Lys Gly Ala
                 50
                                      55
                                                           60
Ile Tyr Glu Gly Asp Trp Lys Phe Gly Lys Arg Asp Gly Tyr Gly
                 65
                                      70
                                                           75
Thr Leu Ser Leu Pro Asp Gln Gln Thr Gly Lys Cys Arg Arg Val
                 80
                                      85
Tyr Ser Gly Trp Trp Lys Gly Asp Lys Lys Ser Gly Tyr Gly Ile
                 95
                                     100
Gln Phe Phe Gly Pro Lys Glu Tyr Tyr Glu Gly Asp Trp Cys Gly
                 110
                                     115
Ser Gln Arg Ser Gly Trp Gly Arg Met Tyr Tyr Ser Asn Gly Asp
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                                     130
                                                          135
Ile Tyr Glu Gly Gln Trp Glu Asn Asp Lys Pro Asn Gly Glu Gly
                                                          150
                 140
                                     145
Met Leu Arg Leu Lys Asn Gly Asn Arg Tyr Glu Gly Cys Trp Glu
                155
                                     160
                                                          165
Arg Gly Met Lys Asn Gly Ala Gly Arg Phe Phe His Leu Asp His
                170
                                     175
                                                          180
Gly Gln Leu Phe Glu Gly Phe Trp Val Asp Asn Met Ala Lys Cys
                                     190
                                                          195
                185
Gly Thr Met Ile Asp Phe Gly Arg Asp Glu Ala Pro Glu Pro Thr
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                                     2.05
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Gln Phe Pro Ile Pro Glu Val Lys Ile Leu Asp Pro Asp Gly Val
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Leu Ala Glu Ala Leu Ala Met Phe Arg Lys Thr Glu Glu Gly Asp
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Arg His Arg Tyr Glu Phe Leu Glu Thr Leu Gly Lys Gly Thr
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Gly Lys Val Lys Lys Ala Arg Glu Ser Ser Gly Arg Leu Val Ala
                                      40
                                                           45
Ile Lys Ser Ile Arg Lys Asp Lys Ile Lys Asp Glu Gln Asp Leu
                 50
                                      55
Met His Ile Arg Arg Glu Ile Glu Ile Met Ser Ser Leu Asn His
                                                           75
Pro His Ile Ile Ala Ile His Glu Val Phe Glu Asn Ser Ser Lys
                 80
                                      85
                                                           90
Ile Val Ile Val Met Glu Tyr Ala Ser Arg Gly Asp Leu Tyr Asp
                 95
                                     100
                                                          105
Tyr Ile Ser Glu Arg Gln Gln Leu Ser Glu Arg Glu Ala Arg His
                110
                                     115
                                                          120
Phe Phe Arg Gln Ile Val Ser Ala Val His Tyr Cys His Gln Asn
                125
                                     130
                                                          135
Arg Val Val His Arg Asp Leu Lys Leu Glu Asn Ile Leu Leu Gly
                140
                                     145
                                                          150
Ala Asn Gly Asn Ile Lys Ile Ala Asp Phe Gly Leu Ser Asn Leu
                155
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Tyr His Gln Gly Lys Phe Leu Gln Thr Phe Cys Gly Ser Pro Leu

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170
                                     175
Tyr Ala Ser Pro Glu Ile Val Asn Gly Lys Pro Tyr Thr Gly Pro
                185
                                     190
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Glu Val Asp Ser Trp Ser Leu Gly Val Leu Leu Tyr Ile Leu Val
                200
                                     205
His Gly Thr Met Pro Phe Asp Gly His Asp His Lys Ile Leu Val
                215
                                     220
                                                          225
Lys Gln Ile Ser Asn Gly Ala Tyr Arg Glu Pro Pro Lys Pro Ser
                230
                                     235
                                                          240
Asp Ala Cys Gly Leu Ile Arg Trp Leu Leu Met Val Asn Pro Thr
                245
                                     250
Arg Arg Ala Thr Leu Glu Asp Val Ala Ser His Trp Trp Val Asn
                260
                                     265
Trp Gly Tyr Ala Thr Arg Val Gly Glu Glu Ala Pro His Glu
                275
                                     280
Gly Gly His Pro Gly Ser Asp Ser Ala Arg Ala Ser Met Ala Asp
                290
                                     295
                                                          300
Trp Leu Arg Arg Ser Ser Arg Pro Leu Leu Glu Asn Gly Ala Lys
                305
                                     310
                                                          315
Val Cys Ser Phe Phe Lys Gln His Ala Pro Gly Gly Gly Ser Thr
                320
                                     325
                                                          330
Thr Pro Gly Leu Glu Arg Gln His Ser Leu Lys Lys Ser Arg Lys
                335
                                     340
Glu Asn Asp Met Ala Gln Ser Leu His Ser Asp Thr Ala Asp Asp
                350
                                     355
                                                          360
Thr Ala His Arg Pro Gly Lys Ser Asn Leu Lys Leu Pro Lys Gly
                365
                                     370
                                                          375
Ile Leu Lys Lys Lys Val Ser Ala Ser Ala Glu Gly Val Gln Glu
                380
                                     385
                                                          390
Asp Pro Pro Glu Leu Ser Pro Ile Pro Ala Ser Pro Gly Gln Ala
                395
                                     400
                                                          405
Ala Pro Leu Leu Pro Lys Lys Gly Ile Leu Lys Lys Pro Arg Gln
                410
                                     415
                                                          420
Arg Glu Ser Gly Tyr Tyr Ser Ser Pro Glu Pro Ser Glu Ser Gly
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                                                          435
Glu Leu Leu Asp Ala Gly Asp Val Phe Val Ser Gly Asp Pro Lys
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                                     445
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Glu Gln Lys Pro Pro Gln Ala Ser Gly Leu Leu Leu His Arg Lys
                455
                                     460
Gly Ile Leu Lys Leu Asn Gly Lys Phe Ser Gln Thr Ala Leu Glu
                470
                                     475
                                                          480
Leu Ala Ala Pro Thr Thr Phe Gly Ser Leu Asp Glu Leu Ala Pro
                485
                                     490
                                                          495
Pro Arg Pro Leu Ala Arg Ala Ser Arg Pro Ser Gly Ala Val Ser
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                                     505
                                                          510
Glu Asp Ser Ile Leu Ser Ser Glu Ser Phe Asp Gln Leu Asp Leu
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                                     520
                                                          525
Pro Glu Arg Leu Pro Glu Pro Pro Leu Arg Gly Cys Val Ser Val
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                                                          540
Asp Asn Leu Thr Gly Leu Glu Glu Pro Pro Ser Glu Gly Pro Gly
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                                     550
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Ser Cys Leu Arg Arg Trp Arg Gln Asp Pro Leu Gly Asp Ser
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Phe Ser Leu Thr Asp Cys Gln Glu Val Thr Ala Thr Tyr Arg Gln
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Ala Leu Arg Val Cys Ser Lys Leu Thr
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Arg	Lys	Asp	Leu	Pro 35	Ser	Arg	Thr	Ser	Asn 40	Val	Glu	Arg	Lys	
Ser	Gln	Gln	Gln	Trp 50	Gly	Arg	Gly	Asn	Phe 55	Thr	Glu	Gly	Lys	Val 60
Pro	His	Ile	Arg	Ile 65	Glu	Asn	Gly	Ala	Ala 70	Ile	Glu	Glu	Ile	Tyr 75
Thr	Phe	Gly	Arg	Ile 80	Leu	Gly	Lys	Gly	Ser 85	Phe	Gly	Ile	Val	Ile 90
Glu	Ala	Thr	Asp	Lys 95	Glu	Thr	Glu	Thr	Lys 100	Trp	Ala	Ile	Lys	Lys 105
Val	Asn	Lys	Glu	Lys 110	Ala	Gly	Ser	Ser	Ala 115	Val	Lys	Leu	Leu	Glu 120
Arg	Glu	Val	Asn	Ile 125	Leu	Lys	Ser	Val	Lys 130	His	Glu	His	Ile	Ile 135
His	Leu	Glu	Gln	Val 140	Phe	Glu	Thr	Pro	Lys 145	Lys	Met	Tyr	Leu	Val 150
				Glu 155					160					165
				Ser 170					175					180
				Ile 185					190					195
				Leu 200					205					210
	_		Asn	215					220	-		Thr	_	225
				Lys 230					235					240
		_	Gly	245			-		250			Val		255
				Ser 260				_	265				_	270
				Leu 275		-	_		280					285
				Leu 290					295					300
			Ala	305	1				310	,		Ala		315
				Leu 320 Leu		_		_	325			_		330
				335 Pro					340					345
				350 Glu					355					360
				365 Thr					370					375
_		_		380 Asp					385					390
				395 Ala			_		400	_				405
				410 Met					415					420
_			_	425 Ile	-				430				-	435
				440 Thr	_	_			445					450
				455 Lys				-J <del>-</del>	460		-4-		<u>x</u>	465
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Leu	His	Pro	Glu	Glu 440	Lys	Thr	Pro	Asp	Ser 445	Tyr	Leu	Gly	Leu	Gly 450
Pro	Gln	Asp	Leu		Ala	Ala	Ser	Leu		Ala	Val	Leu	Leu	-
Gly	Trp	Ile	Leu		Val	Met	Arg	Gln		Gln	Pro	Gln	Val	Val 480
Glu	Lys	Gln	Gln		Thr	Pro	Leu	Ala		Ala	Asp	Phe	Ala	
Ile	Ser	Gln	Asp	Ala 500	Gln	Ser	Leu	His		Gly	Ala	Ser	Arg	Arg 510
Ser	Gln	Lys	Arg	Leu 515	Gln	Ser	Pro	Ser		Gln	Ala	Gln	Pro	Leu 525
Asp	Asp	Pro	Glu	Ala 530	Glu	Gln	Leu	Thr		Val	Gly	Lys	Ile	
Phe	Asn	Pro	Lys		Val	Leu	Gly	Arg		Ala	Gly	Gly	Thr	Phe 555
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Leu	Gln	Glu	Ser		Arg	His	Pro	Asn		Leu	Arg	Tyr	Phe	
Thr	Glu	Arg	Gly		Gln	Phe	His	Tyr		Ala	Leu	Glu	Leu	
Arg	Ala	Ser	Leu		Glu	Tyr	Val	Glu		Pro	Asp	Leu	Asp	
Gly	Gly	Leu	Glu		Glu	Val	Val	Leu		Gln	Leu	Met	Ser	
Leu	Ala	His	Leu		Ser	Leu	His	Ile		His	Arg	Asp	Leu	
Pro	Gly	Asn	Ile		Ile	Thr	Gly	Pro		Ser	Gln	Gly	Leu	
Arg	Val	Val	Leu		Asp	Phe	Gly	Leu		Lys	Lys	Leu	Pro	
Gly	Arg	Суз	Ser		Ser	Leu	His	Ser		Ile	Pro	Gly	Thr	
Gly	Trp	Met	Ala		Glu	Leu	Leu	Gln		Leu	Pro	Pro	Asp	Ser 720
Pro	Thr	Ser	Ala		Asp	Ile	Phe	Ser		Gly	Cys	Val	Phe	
Tyr	Val	Leu	Ser		Gly	Ser	His	Pro		Gly	Asp	Ser	Leu	
Arg	Gln	Ala	Asn		Leu	Thr	Gly	Ala		Cys	Leu	Ala	His	
Glu	Glu	Glu	Val		Asp	Lys	Val	Val		Arg	Asp	Leu	Val	
Ala	Met	Leu	Ser		Leu	Pro	Gln	Pro		Pro	Ser	Ala	Pro	
Val	Leu	Ala	His		Phe	Phe	Trp	Ser		Ala	Lys	Gln	Leu	Gln
Phe	Phe	Gln	Asp	Val	Ser	Asp	Trp	Leu	Glu	Lys	Glu	Ser	Glu	
Glu	Pro	Leu	Val		Ala	Leu	Glu	Ala		Gly	Суз	Ala	Val	
Arg	Asp	Asn	Trp	830 His 845	Glu	His	Ile	Ser		Pro	Leu	Gln	Thr	
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Leu	Arg	Ala	Val		Asn	Lys	Lys	His		Tyr	Arg	Glu	Leu	
Val	Glu	Val	Arg		Ala	Leu	Gly	Gln		Pro	Asp	Gly	Phe	
Gln	Tyr	Phe	Thr		Arg	Phe	Pro	Arg		Leu	Leu	His	Thr	
Arg	Ala	Met	Arg		Cys	Ala	Ser	Glu		Leu	Phe	Leu	Pro	-
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21/61

Ala Leu Ala Gly Glu Asp Thr Gly Val Val Thr His Glu Gln Phe

Lys Ala Ala Leu Arg Met Val Val Asp Gln Gly Asp Pro Arg Leu

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Ser Lys Glu Gly Lys Met Phe Ile Lys Ala Leu Phe Asp Tyr Asn
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Type	Gly	Gly	Glu	Glu	Glu	Glu	Ala	Glu	Glu	Arg	Gly	Arg	His	Ala	Gln
The Cly Leu   Lys   Leu   Cly   Val   His   Cys   Ile   Thr   Cly   Cln   Lys   Cas   Ca	Tyr	Val	Gly	Pro	Tyr	Arg	Leu	Glu	Lys	Thr	Leu	Gly	Lys	Gly	Gln
Name   Line   Lys   Line   Val   Ash   Ash   Glu   Lys   Leu   Ser   Glu   Ser   Val   90   90   90   90   90   90   90   9	Thr	Gly	Leu	Val	Lys	Leu	Gly	Val	His	Cys	Ile	Thr	Gly	Gln	Lys
Met   Lys   Call   Arg   Glu   Le   Ala   Ite   Leu   Lys   Leu   Ite   Glu   Se   Se   Se   Se   Se   Se   Se   S	Val	Ala	Ile	Lys	Ile	Val	Asn	Arg	Glu	Lys	Leu	Ser	Glu	Ser	Val
His   Pro   His   Val   Leu   Lys   Leu   His   Asp   Val   15   15   17   12   12   12   12   12   12   12	Leu	Met	Lys	Val	Glu	Arg	Glu	Ile	Ala,	Ile	Leu	Lys	Leu	Ile	Glu
Tyr         Leu         Val         Leu         Val         Ser         Gly         Gly         Gly         Leu         Phe         135           Asp         Tyr         Leu         Val         Lys         Lys         Lys         Ru         Leu         Thr         Pro         Lys         Glu         Ala         Arg         Leu         Thr         Pro         Lys         Glu         Lala         Arg         150	His	Pro	His	Val	Leu	Lys	Leu	His	Asp	Val	Tyr	Glu	Asn	Lys	Lys
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Type	Asp	Tyr	Leu	Val		Lys	Gly	Arg	Leu		Pro	Lys	Glu	Ala	
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195   195					170					175		Asn	Leu	Leu	
Ser   Ser	_		_		185		_			190		_			
Arg   Arg   Ala   Asp   Ala   Asp   Ala   Asp   Arg   Arg					200					205		_	_		210
Leu Val Gly Ala Leu Pro Phe Asp					215					220					225
Leu   Glu   Lys   Val   Lys   Arg   Gly   Val   Phe   His   Met   Pro   His   Phe   Ile   270   275   280   285				_	230	_		_	_	235					240
Pro   Pro   Asp   Cys   Gln   Ser   Leu   Leu   Arg   Gly   Met   Ile   Glu   Val   Glu   285					245					250					255
Pro   Clu   Lys   Arg   Leu   Ser   Leu   Clu   Clu   Clu   Tle   Clu   Lys   His   Pro   Trp   290   295   300     Tyr   Leu   Cly   Cly   Lys   His   Clu   Pro   Asp   Pro   Cys   Leu   Clu   Pro   Ala   305   315     Pro   Cly   Arg   Arg   Val   Ala   Met   Arg   Ser   Leu   Pro   Ser   Asn   Cly   Clu   315     Pro   Cly   Arg   Arg   Val   Ala   Met   Arg   Ser   Leu   Pro   Ser   Asn   Cly   Clu   315     Leu   Asp   Pro   Asp   Val   Leu   Clu   Ser   Met   Ala   Ser   Leu   Cly   Cys   Phe   335     Arg   Asp   Arg   Clu   Arg   Leu   His   Arg   Clu   Leu   Arg   Ser   Clu   Clu   Clu   350     Asn   Cln   Clu   Lys   Met   Tle   Tyr   Tyr   Leu   Leu   Leu   Asp   Arg   Lys   Clu   370     Asn   Cln   Clu   Asp   Arg   Clu   Asp   Clu   Asp   Asp   Asp   Asp   Asp   Asp   380     Val   Asp   Pro   Pro   Arg   Lys   Arg   Val   Asp   Ser   Pro   Met   Leu   Ser   Asp			_		260		_			265					270
Tyr Leu Gly Gly Lys His Glu Pro Asp Pro Cys Leu Glu Pro Ala 300  Tyr Leu Gly Gly Lys His Glu Pro Asp Pro Cys Leu Glu Pro Ala 315  Pro Gly Arg Arg Val Ala Met Arg Ser Leu Pro Ser Asn Gly Glu 320  Leu Asp Pro Asp Val Leu Glu Ser Met Ala Ser Leu Gly Cys Phe 335  Arg Asp Arg Glu Arg Leu His Arg Glu Leu Arg Ser Glu Glu Glu 360  Asn Gln Glu Lys Met 350  Arg Tyr Pro Ser Cys Glu Asp Gln Asp Leu Leu Asp Arg Lys Glu 370  Arg Tyr Pro Ser Cys Glu Asp Gln Asp Leu Leu Asp Arg Lys Glu 370  Arg Tyr Pro Ser Cys Glu Asp Gln Asp Lys Ser Met Glu Val Leu Ser Arg 390  Val Asp Pro Pro Arg Arg Pro Glu Arg Lys Ser Met Glu Val Leu Ser Arg 400  His Gly Lys Arg Arg Pro Glu Arg Lys Ser Met Glu Val Leu Ser Arg 420  Ile Thr Asp Ala Gly Gly Gly Gly Ser Pro Val Pro Thr Arg Arg 435  Ala Leu Glu Met Ala Gln His Ser Gln Arg Ser Arg Ser Val Ser 450  Gly Ala Ser Thr Gly Leu Ser Ser Ser Pro Leu Ser Arg Ser Val Ser 465  Ser Pro Val Phe Ser Phe Ser Pro Glu Pro Gly Ala Gly Asp Glu Arg 465  Ser Pro Val Pro Gly Ser Pro Thr Ser Lys Thr Gln Thr Leu Pro 485  Arg Gly Gly Fro Arg Gly Gly Gly Gly Ala Gly Glu Gln Pro Pro Pro Pro Arg 490			_	-	275				_	280					285
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Arg       Tyr       Pro       Ser       Cys       Glu       Asp       Gln       Asp       Leu       Pro       Pro       Arg       Asp       390         Val       Asp       Pro       Pro       Arg       Lys       Arg       Val       Asp       Ser       Pro       Met       Leu       Ser       Arg       Arg       405         His       Gly       Lys       Arg       Lys       Ser       Met       Glu       Val       Leu       Ser       Arg       405         Ile       Thr       Asp       Ala       Gly       Gly       Gly       Ser       Pro       Val       Pro       Thr       Arg       A					350					355 Leu					360
Val         Asp         Pro         Pro         Arg         Lys         Arg         Val         Asp         Ser         Pro         Met         Leu         Ser         Arg         Arg         405         Arg         Arg         Arg         Pro         Glu         Arg         Lys         Ser         Met         Glu         Val         Leu         Ser         Arg         Arg <td>Arg</td> <td>Tyr</td> <td>Pro</td> <td>Ser</td> <td>Cys</td> <td>Glu</td> <td>Asp</td> <td>Gln</td> <td>Asp</td> <td>Leu</td> <td>Pro</td> <td>Pro</td> <td>Arg</td> <td>Asn</td> <td></td>	Arg	Tyr	Pro	Ser	Cys	Glu	Asp	Gln	Asp	Leu	Pro	Pro	Arg	Asn	
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Ile Thr Asp Ala Gly Gly Gly Gly Gly Ser Pro Val Pro Thr Arg Arg 435         Ala Leu Glu Met Ala Gln His Ser Gln Arg Ser Arg Ser Val Ser 450         Gly Ala Ser Thr Gly Leu Ser Ser Ser Pro Leu Ser Ser Pro Arg 465         Ser Pro Val Phe Ser Phe Ser Pro Glu Pro Gly Ala Gly Asp Glu 470         Ala Arg Gly Gly Gly Ser Pro Gly Ala Gly Gly Ser Pro Arg 490         Ser Arg Gly Pro Arg Gly Gly Gly Gly Gly Ala Gly Glu Gln Pro Pro Pro	His	Gly	Lys	Arg	Arg	Pro	Glu	Arg	Lys	Ser	Met	Glu	Val	Leu	Ser
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Gln Thr Thr Gly Gly Phe Tyr Pro Leu Ile Gly Trp Lys Ile Gly
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Gln Glu Val Val Cys Leu Ala Glu Ser Asn Ala Gly Asp Thr Gly
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                                     310
                                                          315
Thr Ala Ile Lys Trp Ala Gln Gln Leu Asp Leu Phe Thr Asp Ala
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                                     325
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Ala Glu Thr Glu Lys Met Ala Lys Ser Leu Glu Asp Ser Glu Gly
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                                     340
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Val Cys Phe Val Pro Ser Phe Ser Gly Leu Gln Ala Pro Leu Asn
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                                     355
                                                          360
Asp Pro Trp Ala Cys Ala Ser Phe Met Gly Leu Lys Pro Ser Thr
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                                     370
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Ser Lys Tyr His Leu Val Arg Ala Ile Leu Glu Ser Ile Ala Phe
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                                     385
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Arg Asn Lys Gln Leu Tyr Glu Met Met Lys Lys Glu Ile His Ile
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                                     400
                                                          405
Pro Val Arg Lys Ile Arg Ala Asp Gly Gly Val Cys Lys Asn Gly
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Phe Val Met Gln Met Thr Ser Asp Leu Ile Asn Glu Asn Ile Asp
                425
                                     430
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Arg Pro Ala Asp Ile Asp Met Ser Cys Leu Gly Ala Ala Ser Leu
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                                     445
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Ala Gly Leu Ala Val Gly Phe Trp Thr Asp Lys Glu Glu Leu Lys
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                                     460
Lys Leu Arg Gln Ser Glu Val Val Phe Lys Pro Gln Lys Lys
                                                         Cys
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Gln Glu Tyr Glu Met Ser Leu Glu Asn Trp Ala Lys Ala Val Lys
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Arg Ser Met Asn Trp Tyr Asn Lys Thr
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Gln Arg Arg Arg Ser Leu Ala Leu Pro Gly Leu Gln Gly Cys
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Glu Gln Gln Pro Ile Gly Arg Arg Leu Phe Arg Asp Phe Leu Ala
Thr Val Pro Thr Phe Arg Lys Ala Ala Thr Phe Leu Glu Asp Val
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                                      85
Gln Asn Trp Glu Leu Ala Glu Glu Gly Pro Thr Lys Asp Ser Ala
                 95
                                     100
Leu Gln Gly Leu Val Ala Thr Cys Ala Ser Ala Pro Ala Pro Gly
                110
                                     115
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Asn Pro Gln Pro Phe Leu Ser Gln Ala Val Ala Thr Lys Cys Gln
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Ala Ala Thr Thr Glu Glu Glu Arg Val Ala Ala Val Thr Leu Ala
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Lys Ala Glu Ala Met Ala Phe Leu Gln Glu Gln Pro Phe Lys Asp
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Phe Val Thr Ser Ala Phe Tyr Asp Lys Phe Leu Gln Trp Lys Leu
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Phe Glu Met Gln Pro Val Ser Asp Lys Tyr Phe Thr Glu Phe Arg
                185
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Val Leu Gly Lys Gly Gly Phe Gly Glu Val Cys Ala Val Gln Val
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                                     205
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Lys Asn Thr Gly Lys Met Tyr Ala Cys Lys Leu Asp Lys Lys
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Arg Leu Lys Lys Gly Gly Glu Lys Met Ala Leu Leu Glu Lys
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Glu Ile Leu Glu Lys Val Ser Ser Pro Phe Ile Val Ser Leu Ala
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Tyr Ala Phe Glu Ser Lys Thr His Leu Cys Leu Val Met Ser Leu
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Met Asn Gly Gly Asp Leu Lys Phe His Ile Tyr Asn Val Gly Thr
                275
                                     280
Arg Gly Leu Asp Met Ser Arg Val Ile Phe Tyr Ser Ala Gln Ile
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                                     295
Ala Cys Gly Met Leu His Leu His Glu Leu Gly Ile Val Tyr Arg
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                                     310
Asp Met Lys Pro Glu Asn Val Leu Leu Asp Asp Leu Gly Asn Cys
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                                     325
Arg Leu Ser Asp Leu Gly Leu Ala Val Glu Met Lys Gly Gly Lys
                335
                                     340
Pro Ile Thr Gln Arg Ala Gly Thr Asn Gly Tyr Met Ala Pro Glu
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Ile Leu Met Glu Lys Val Ser Tyr Ser Tyr Pro Val Asp Trp Phe
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                                     370
Ala Met Gly Cys Ser Ile Tyr Glu Met Val Ala Gly Arg Thr Pro
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Phe Lys Asp Tyr Lys Glu Lys Val Ser Lys Glu Asp Leu Lys Gln
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                                     400
Arg Thr Leu Gln Asp Glu Val Lys Phe Gln His Asp Asn Phe Thr
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Glu Glu Ala Lys Asp Ile Cys Arg Leu Phe Leu Ala Lys Lys Pro
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Glu Gln Arg Leu Gly Ser Arg Glu Lys Ser Asp Asp Pro Arg Lys
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His His Phe Phe Lys Thr Ile Asn Phe Pro Arg Leu Glu Ala Gly
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Leu Ile Glu Pro Pro Phe Val Pro Asp Pro Ser Val Val Tyr Ala
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Lys Asp Ile Ala Glu Ile Asp Asp Phe Ser Glu Val Arg Gly Val
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                                     490
                                                         495
Glu Phe Asp Asp Lys Asp Lys Gln Phe Phe Lys Asn Phe Ala Thr
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                                     505
Gly Ala Val Pro Ile Ala Trp Gln Glu Ile Ile Glu Thr Gly
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                                     520
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Leu Phe Glu Glu Leu Asn Asp Pro Asn Arg Pro Thr Gly Cys Glu
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Glu Gly Asn Ser Ser Lys Ser Gly Val Cys Leu Leu
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Val Ser Ser Leu Arg Val Gly Ser Ser Gln Val Gly Thr Glu Pro
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                                                           45
Gly Pro Ser Leu Asp Ala Glu Gly Trp Thr Gln Glu Ala Glu Asp
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                                      55
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Leu Ser Asp Ser Thr Pro Thr Leu Gln Arg Pro Gln Glu Gln Val
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                                      70
                                                           75
Thr Met Arg Lys Phe Ser Leu Gly Gly Arg Gly Gly Tyr Ala Gly
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                                      85
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Val Ala Gly Tyr Gly Thr Phe Ala Phe Gly Gly Asp Ala Gly Gly
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                                     100
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Met Leu Gly Gln Gly Pro Met Trp Ala Arg Ile Ala Trp Ala Val
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                                     115
                                                          120
Ser Gln Ser Glu Glu Glu Glu Glu Glu Ala Arg Ala Glu Ser
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                                     130
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Gln Ser Glu Glu Gln Glu Ala Arg Ala Glu Ser Pro Leu Pro
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Gln Val Ser Ala Arg Pro Val Pro Glu Val Gly Arg Ala Pro Thr
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                                                          165
Arg Ser Ser Pro Glu Pro Thr Pro Trp Glu Asp Ile Gly Gln Val
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                                     175
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Ser Leu Val Gln Ile Arg Asp Leu Ser Gly Asp Ala Glu Ala Ala
                 185
                                     190
Asp Thr Ile Ser Leu Asp Ile Ser Glu Val Asp Pro Ala Tyr Leu
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Asn Leu Ser Asp Leu Tyr Asp Ile Lys Tyr Leu Pro Phe Glu Phe
                 215
                                     220
                                                          225
Met Ile Phe Arg Lys Val Pro Lys Ser Ala Gln Pro Glu Pro Pro
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                                     235
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Ser Pro Met Ala Glu Glu Glu Leu Ala Glu Phe Pro Glu Pro Thr
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Trp Pro Trp Pro Gly Glu Leu Gly Pro His Ala Gly Leu Glu Ile
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Thr Glu Glu Ser Glu Asp Val Asp Ala Leu Leu Ala Glu Ala Ala
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                                     280
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Val Gly Arg Lys Arg Lys Trp Ser Ser Pro Ser Arg Ser Leu Phe
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His Phe Pro Gly Arg His Leu Pro Leu Asp Glu Pro Ala Glu Leu
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                                     310
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Gly Leu Arg Glu Arg Val Lys Ala Ser Val Glu His Ile Ser Arg
                                     325
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Ile Leu Lys Gly Arg Pro Glu Gly Leu Glu Lys Glu Gly Pro Pro
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                                     340
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Arg Lys Lys Pro Gly Leu Ala Ser Phe Arg Leu Ser Gly Leu Lys
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Ser Trp Asp Arg Ala Pro Thr Phe Leu Arg Glu Leu Ser Asp Glu
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Thr Val Val Leu Gly Gln Ser Val Thr Leu Ala Cys Gln Val Ser
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Ala Gln Pro Ala Ala Gln Ala Thr Trp Ser Lys Asp Gly Ala Pro
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                                     400
Leu Glu Ser Ser Ser Arg Val Leu Ile Ser Ala Thr Leu Lys Asn
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Phe	Gln	Leu	Leu	410 Thr 425	Ile	Leu	Val	Val	415 Val 430	Ala	Glu	Asp	Leu	420 Gly 435
Val	Tyr	Thr	Cys	Ser 440	Val	Ser	Asn	Ala		Gly	Thr	Val	Thr	
Thr	Gly	Val	Leu	Arg 455	Lys	Ala	Glu	Arg		Ser	Ser	Ser	Pro	
Pro	Asp	Ile	Gly	Glu 470	Val	Tyr	Ala	Asp		Val	Leu	Leu	Val	
Lys	Pro	Val	Glu		Tyr	Gly	Pro	Val		Tyr	Ile	Val	Gln	
Ser	Leu	Glu	Gly	Gly 500	Ser	Trp	Thr	Thr		Ala	ser	Asp	Ile	
Asp	Cys	Cys	Tyr	Leu 515	Thr	Ser	Lys	Leu		Arg	Gly	Gly	Thr	Tyr 525
Thr	Phe	Arg	Thr	Ala 530	Cys	Val	Ser	Lys		Gly	Met	Gly	Pro	
Ser	Ser	Pro	Ser	Glu 545	Gln	Va1	Leu	Leu	Gly 550	Gly	Pro	Ser	His	
Ala	Ser	Glu	Glu	Glu 560	Ser	Gln	Gly	Arg	Ser 565	Ala	Gln	Pro	Leu	
Ser	Thr	Lys	Thr	Phe 575	Ala	Phe	Gln	Thr	Gln 580	Ile	Gln	Arg	Gly	Arg 585
Phe	Ser	Val	Val	Arg 590	Gln	Cys	Trp	Glu	Lys 595	Ala	Ser	Gly	Arg	Ala 600
Leu	Ala	Ala	Lys	Ile 605	Ile	Pro	Tyr	His	Pro 610	Lys	Asp	Lys	Thr	Ala 615
Val	Leu	Arg	Glu	Tyr 620	Glu	Ala	Leu	Lys	Gly 625	Leu	Arg	His	Pro	His 630
Leu	Ala	Gln	Leu	His 635	Ala	Ala	Tyr	Leu	Ser 640	Pro	Arg	His	Leu	Val 645
Leu	Ile	Leu	Glu	Leu 650	Суѕ	Ser	Gly	Pro	Glu 655	Leu	Leu	Pro	Cys	Leu 660
		_	Ala	665			Glu		670				_	Leu 675
				Ser 680					685					Ile 690
				Leu 695					700					705
				Val 710					715					720
				Leu 725					730					735
				Glu 740				_	745	_				750
	_		Trp	755			Val		760					765
				Val 770			•		775					780
				Gly 785					790					795
				Ala 800					805				_	Ala 810
				Arg 815					820	_		Gln	_	Pro 825
				Glu 830					835					840
				Ala 845					850					855
Lys	Arg	Arg	Ala	Leu 860	Leu	Tyr	ГĀг	Arg	His 865	Asn	Leu	Ala	Gln	Val 870
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Glu Asp Glu Glu Asp Lys Lys Pro Met Ser Leu Ser Thr Gln Val

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Lys Ser Ala Pro Val Leu Asn Gln Ile Phe Glu Glu Gly Glu Ser
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Asp Asp Glu Phe Asp Met Asp Glu Asn Leu Pro Pro Lys Leu Ser
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                                     505
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Arg Leu Lys Met Asn Ile Ala Ser Pro Gly Thr Val His Lys Arg
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                                     520
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Tyr His Arg Arg Lys Ser Gln Gly Arg Gly Ser Ser Cys Ser
                                                         Ser
                530
                                     535
                                                         540
Ser Glu Thr Ser Asp Asp Ser Glu Ser Arg Arg Leu Asp
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                                     550
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Lys Asp Ser Gly Phe Thr Tyr Ser Trp His Arg Arg Asp Ser
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Glu Gly Pro Pro Gly Ser Glu Gly Asp Gly Gly Gln Ser Lys
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Pro Ser Asn Ala Ser Gly Gly Val Asp Lys Ala Ser Pro Ser Glu
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Asn Asn Ala Gly Gly Gly Ser Pro Ser Ser Gly Ser Gly Asn
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Pro Thr Asn Thr Ser Gly Thr Thr Arg Arg Cys Ala Gly Pro Ser
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Asn Ser Met Gln Leu Ala Ser Arg Ser Ala Gly Glu Leu Val Glu
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Ser Leu Lys Leu Met Ser Leu Cys Leu Gly Ser Gln Leu His Gly
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Ser Thr Lys Tyr Ile Ile Asp Pro Gln Asn Gly Leu Ser Phe Ser
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Ser Val Lys Val Gln Glu Lys Ser Thr Trp Lys Met Cys Ile Ser
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Ser Thr Gly Asn Ala Gly Gln Val Pro Ala Val Gly Gly Ile Lys
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Phe Phe Ser Asp His Met Ala Asp Thr Thr Thr Glu Leu Glu Arg
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Ile Lys Ser Lys Asn Leu Lys Asn Asn Val Leu Gln Leu Pro Leu
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Cys Glu Lys Thr Ile Ser Val Asn Ile Gln Arg Asn Pro Lys Glu
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Asn Thr Lys Lys Leu Ser Ala Arg Asp His Gln Lys Leu Glu Arg
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Glu Ala Arg Ile Cys Arg Leu Leu Lys His Pro Asn Ile Val Arg
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                                      70
Leu His Asp Ser Ile Ser Glu Glu Gly Phe His Tyr Leu Val Phe
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Asp Leu Val Thr Gly Gly Glu Leu Phe Glu Asp Ile Val Ala Arg

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				Glu 110					115	-				120
Leu	Glu	Ser	Val	Asn 125	His	Ile	His	Gln	His 130	Asp	Ile	Val	His	Arg 135
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Ala	Ala	Val	Lys	Leu 155	Ala	Asp	Phe	Gly	Leu 160	Ala	Ile	Glu	Val	Gln 165
Gly	Glu	Gln	Gln	Ala 170	Trp	Phe	Gly	Phe	Ala 175	Gly	Thr	Pro	Gly	
Leu	ser	Pro	Glu	Val 185	Leu	Arg	Lys	Asp		Tyr	Gly	Lys	Pro	
qzA	Ile	Trp	Ala	Cys 200	Gly	Val	Ile	Leu		Ile	Leu	Leu	Val	
Tyr	Pro	Pro	Phe	Trp	qzA	Glu	Asp	Gln	His 220	Lys	Leu	Tyr	Gln	
Ile	Lys	Ala	Gly	Ala 230	Tyr	Asp	Phe	Pro		Pro	Glu	Trp	Asp	
Val	Thr	Pro	Glu	Ala 245	Lys	Asn	Leu	Ile		Gln	Met	Leu	Thr	
Asn	Pro	Ala	Lys	Arg 260	Ile	Thr	Ala	Asp	Gln 265	Ala	Leu	Lys	Tyr	
Trp	Val	Cys	Gln	Arg 275	Ser	Thr	Val	Ala	Ser 280	Met	Met	His	Arg	Gln 285
Glu	Thr	Val	Glu	Cys 290	Leu	Arg	Lys	Phe	Asn 295	Ala	Arg	Arg	Lys	
Lys	Gly	Ala	Ile	Leu 305	Thr	Thr	Met	Leu	Val 310	Ser	Arg	Asn	Phé	
Val	Gly	Arg	Gln	ser 320	ser	Ala	Pro	Ala	ser 325	Pro	Ala	Ala	Ser	Ala 330
Ala	Gly	Leu	Ala	Gly 335	Gln	Ala	Ala	Lys	Ser 340	Leu	Leu	Asn	Lys	Lys 345
Ser	Asp	Gly	Gly	Val 350	Lys	Lys	Arg	Lys	Ser 355	Ser	Ser	Ser	Val	His 360
Leu	Met	Pro	Gln	Ser 365	Asn	Asn	Lys	Asn	Ser 370	Leu	Val	Ser	Pro	Ala 375
				Pro 380					385					390
Val	Val	His	Asn	Ala 395	Thr	Asp	Gly	Ile	Lys 400	Gly	Ser	Thr	Glu	Ser 405
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Arg	Thr	Gly	Asn	Gly 425	Ser	Ser	Val	Pro	Glu 430	Gly	Arg	Ser	Ser	Arg 435
Asp	Arg	Thr	Ala	Pro 440	Ser	Ala	Gly	Met	Gln 445	Pro	Gln	Pro	Ser	Leu 450
Cys	Ser	Ser	Ala	Met 455	Arg	Lys	Gln	Glu	Ile 460	Ile	Lys	Ile	Thr	Glu 465
Gln	Leu	Ile	Glu	Ala 470	Ile	Asn	Asn	Gly	Asp 475	Phe	Glu	Ala	Tyr	Thr 480
Lys	Ile	Cys	Asp	Pro 485	Gly	Leu	Thr	Ser	Phe 490	Glu	Pro	Glu	Ala	Leu 495
Gly	Asn	Leu	Val	Glu 500	Gly	Met	qzA	Phe	His 505	Lys	Phe	Tyr	Phe	Glu 510
Asn	Leu	Leu	Ser	Lys 515	Asn	Ser	Lys	Pro	Ile 520	His	Thr	Thr	Ile	Leu 525
Asn	Pro	His	Val	His 530	Val	Ile	Gly	Glu	Asp 535	Ala	Ala	Cys	Ile	Ala 540
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Thr	Ser	Gln	Ser	Glu 560	Glu	Thr	Arg	Val	Trp 565	His	Arg	Arg	Asp	Gly 570
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Pro	Leu	Gln												

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Ser Leu Pro Pro Arg Arg Arg Ala Pro Pro Gly Arg Gln Arg Leu
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Glu Glu Arg Thr Gly Pro Ala Gly Pro Glu Gly Lys Glu Gln Asp
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Val Ala Thr Gly Val Ser Pro Leu Leu Phe Arg Lys Leu Ser Asn
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Pro Asp Ile Phe Ser Ser Thr Gly Lys Val Lys Leu Gln Arg Gln
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Leu Ser Gln Asp Asp Cys Lys Leu Trp Arg Gly Asn Leu Ala Ser
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Ser Leu Ser Gly Lys Gln Leu Leu Pro Leu Ser Ser Ser Val His
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Ser Ser Val Gly Gln Val Thr Trp Gln Ser Ser Gly Glu Ala Ser
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Asn Leu Val Arg Met Arg Asn Gln Ser Leu Gly Gln Ser Ala Pro
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Ser Leu Thr Ala Gly Leu Lys Glu Leu Ser Leu Pro Arg Arg Gly
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Ser Phe Cys Arg Thr Ser Asn Arg Lys Ser Leu Ile Val Thr Ser
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Ser Thr Ser Pro Thr Leu Pro Arg Pro His Ser Pro Leu His Gly
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                185
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His Thr Gly Asn Ser Pro Leu Asp Ser Pro Arg Asn Phe Ser Pro
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Asn Ala Pro Ala His Phe Ser Phe Val Pro Ala Arg Arg Thr Asp
                215
                                     220
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Gly Arg Arg Trp Ser Leu Ala Ser Leu Pro Ser Ser Gly
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Thr Asn Thr Pro Ser Ser Thr Val Ser Ser Ser Cys Ser Ser Gln
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Glu Lys Leu His Gln Leu Pro Phe Gln Pro Thr Ala Asp Glu Leu
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                                     265
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His Phe Leu Thr Lys His Phe Ser Thr Glu Ser Val Pro Asp Glu
                275
                                     280
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Glu Gly Arg Gln Ser Pro Ala Met Arg Pro Arg Ser Arg Ser Leu
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                                     295
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Ser Pro Gly Arg Ser Pro Val Ser Phe Asp Ser Glu Ile Ile Met
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Met Asn His Val Tyr Lys Glu Arg Phe Pro Lys Ala Thr Ala Gln
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Met Glu Glu Arg Leu Ala Glu Phe Ile Ser Ser Asn Thr Pro Asp
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Ser Val Leu Pro Leu Ala Asp Gly Ala Leu Ser Phe Ile His His
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Gln Val Ile Glu Met Ala Arg Asp Cys Leu Asp Lys Ser Arg Ser
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Gly Leu Ile Thr Ser Gln Tyr Phe Tyr Glu Leu Gln Glu Asn Leu
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Glu Lys Leu Leu Gln Asp Ala His Glu Arg Ser Glu Ser Ser Glu
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Val Ala Phe Val Met Gln Leu Val Lys Lys Leu Met Ile Ile Ile
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Ala Arg Pro Ala Arg Leu Leu Glu Cys Leu Glu Phe Asp Pro Glu
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Gly	Gln	Gly	Íle		Суз	Asp	Ile	Pro		Tyr	Ile	Val	Ser	
Leu	Gly	Leu	Thr	_	Asp	Pro	Leu	Glu		Met	Ala	Gln	Leu	
Ser	Cys	Asp	Ser	Pro 485	Asp	Thr	Pro	Glu		Asp	Asp	Ser	Ile	
Gly	His	Gly	Ala	Ser 500	Leu	Pro	Ser	Lys	Lys 505	Thr	Pro	Ser	Glu	Glu 510
Asp	Phe	Glu	Thr	Ile 515	Lys	Leu	Ile	Ser	Asn 520	Gly	Ala	Tyr	Gly	Ala 525
Val	Phe	Leu	Val	Arg 530	His	Lys	Ser	Thr	Arg 535	Gln	Arg	Phe	Ala	Met 540
Lys	Lys	Ile	Asn	Lys 545	Gln	Asn	Leu	Ile	Leu 550	Arg	Asn	Gln	Ile	Gln 555
Gln	Ala	Phe	Val		Arg	Asp	Ile	Leu		Phe	Ala	Glu	Asn	
Phe	Val	Val	Ser	Met 575	Phe	Cys	Ser	Phe	Asp 580	Thr	Lys	Arg	His	
Суѕ	Met	Val	Met	Glu 590	Tyr	Val	Glu	Gly	Gly 595	Asp	Cys	Ala	Thr	Leu 600
Leu	Lys	Asn	Ile	Gly 605	Ala	Leu	Pro	Val	Asp 610	Met	Val	Arg	Leu	Tyr 615
			Thr	620					625				_	Gly 630
			Arg	635					640					Ser 645
Met	Gly	His	Ile	Lуs 650					655					660
			Leu	665			Leu	_	670	_				675
		_	Glu	680		_	_		685	_	_			690
Tyr	Ile	Ala	Pro	Glu 695	Val	Ile	Leu	Arg	Gln 700	Gly	Tyr	Gly	Lys	Pro 705
Va1	Asp	Trp	Trp	Ala 710	Met	Gly	Ile		715	-				720
	_		Pro	725		_	-		730				Phe	735
			Ser	740				_	745		_	_		750
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			Glu	770		_		-	775		-			780
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Glu	Asp	Glu	Glu	Glu 830	Val	Ser	Glu	qaA	Gly 835	Суз	Leu	Glu	Ile	Arg 840
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Met	Glu	Arg	Leu	Ser 860	Leu	Leu	Glu	Glu	Arg 865	Arg	Thr	Pro	Pro	Pro 870
Thr	Lys	Arg	Ser	Leu 875	Ser	Glu	Glu	Lys	Glu 880	Asp	His	Ser	Asp	Gly 885
Leu	Ala	Gly	Leu		Gly	Arg	Asp	Arg		Trp	Val	Ile	Gly	
Pro	Glu	Ile	Leu		Lys	Arg	Leu	Ser		Ser	Glu	Ser	Ser	
Thr	Glu	Ser	Asp		Ser	Pro	Pro	Met		Val	Arg	Arg	Arg	

Ser Gly Leu Leu Asp Ala Pro Arg Phe Pro Glu Gly Pro Glu Glu Ala Ser Ser Thr Leu Arg Gln Pro Gln Glu Gly Ile Trp Val Leu Thr Pro Pro Ser Gly Glu Gly Val Ser Gly Pro Val Thr Glu His Ser Gly Glu Gln Arg Pro Lys Leu Asp Glu Glu Ala Val Gly Arg Ser Ser Gly Ser Ser Pro Ala Met Glu Thr Arg Gly Arg Gly Thr Ser Gln Leu Ala Glu Gly Ala Thr Ala Lys Ala Ile Ser Asp Leu Ala Val Arg Arg Ala Arg His Arg Leu Leu Ser Gly Asp Ser Thr Glu Lys Arg Thr Ala Arg Pro Val Asn Lys Val Ile Lys Ser Ala Ser Ala Thr Ala Leu Ser Leu Leu Ile Pro Ser Glu His His Thr Cys Ser Pro Leu Ala Ser Pro Met Ser Pro His Ser Gln Ser Ser Asn Pro Ser Ser Arg Asp Ser Ser Pro Ser Arg Asp Phe Leu Pro Ala Leu Gly Ser Met Arg Pro Pro Ile Ile Ile His Arg Ala Lys Lys Tyr Gly Phe Thr Leu Arg Ala Ile Arg Val Tyr Met Gly Asp Ser Asp Val Tyr Thr Val His His Met Val Trp His Val Glu Asp Gly Gly Pro Ala Ser Glu Ala Gly Leu Arg Gln Gly Asp Leu Ile Thr His Val Asn Gly Glu Pro Val His Gly Leu Val His Thr Glu Val Val Glu Leu Ile Leu Lys Ser Gly Asn Lys Val Ala Leu Glu Asn Thr Ser Ile Lys Val Gly Pro Ile Ser Thr Thr Pro Ala Arg Lys Gly Ser Tyr Lys Ala Lys Met Ala Arg Arg Ser Lys Arg Ser Arg Gly Lys Asp Gly Gln Glu Ser Arg Lys Arg Ser Ser Leu Phe Arg Lys Ile Thr Lys Gln Ala Ser Leu Leu His Thr Ser Ser Leu Ser Ser Leu Asn Arg Ser Leu Ser Ser Gly Glu Ser Arq Gly Pro Gly Ser Pro Thr His Ser His Ser Leu Ser Pro Arg Ser Thr Gln Gly Tyr Arg Val Thr Pro Asp Ala Val His Ser Val Gly Gly Asn Ser Ser Gln Ser Ser Ser Pro Ser Ser Ser Val Pro Ser Ser Pro Ala Gly Ser Gly His Thr Arg Pro Ser Ser Leu His Gly Leu Ala Pro Lys Leu Gln Arg Gln Tyr Arg Ser Pro Arg Arg Lys Ser Ala Gly Ser Ile Pro Leu Ser Pro Leu Ala His Thr Pro Thr Ala Ser Pro Gln Arg Ser Pro Ser Pro Ser Pro Pro Pro Pro Leu Ser Gly His Val Ala Gln Ala Phe Pro Thr Lys Leu His Leu Ser Pro Pro Leu Gly Arg Gln Leu Ser Arg Pro Lys Ser Ala Glu Pro Pro Arg Ser Pro Leu Leu Lys Arg Val Gln Ser Ala Glu Lys Leu Ala Ala Ala Leu Ala Ala Ser Glu Lys Lys Leu Ala Thr Ser Arg Lys His Ser Leu Asp Leu Pro His Ser Glu Leu Lys Lys Glu

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Asn Leu Ser Pro Arg Glu Gln Gly Lys Thr Gln Pro Pro Ser Ala
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                                    1705
Pro Arg Leu Ala His Pro Ser Tyr Glu Asp Pro Ser Gln Gly Trp
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Leu Trp Glu Ser Glu Cys Ala Gln Ala Val Lys Glu Asp Pro Ala
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Leu Ser Ile Thr Gln Val Pro Asp Ala Ser Gly Asp Arg Arg Gln
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Asp Val Pro Cys Arg Gly Cys Pro Leu Thr Gln Lys Ser Glu Pro
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Pro Cys Tyr Leu Trp Phe Val Met Glu Phe Cys Glu Gly Gly Asp
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Lys Asn His Ile Val His Arg Asp Leu Lys Pro Asp Asn Ile Leu
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Gly Asn Gln Asp Asn Lys Asn Val Asn Val Asn Lys Tyr Trp Leu
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Ser Ser Ala Cys Gly Ser Asp Phe Tyr Met Ala Pro Glu Val Trp
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Glu Gly His Tyr Thr Ala Lys Ala Asp Ile Phe Ala Leu Gly Ile
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Ile Ile Trp Ala Met Ile Glu Arg Ile Thr Phe Ile Asp Ser Glu
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Thr Lys Lys Glu Leu Leu Gly Thr Tyr Ile Lys Gln Gly Thr Glu
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Leu His Ile Pro Gln Lys Arg Arg Thr Ser Met Ser Glu Gly Ile
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Lys Gln Leu Lys Asp Met Leu Ala Ala Asn Pro Gln Asp Arg
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Gly Lys Ser Leu Gln Ala Ala Leu Leu Ala Gln Lys Tyr Arg Leu
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Tyr Asn Pro Asn Arg Val Phe Phe Leu Asn Val Pro Phe Asp Ser
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Gln Ala Arg Leu Leu Gln Asn Pro Lys Asp Ala Glu Glu Gln Val
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Lys Leu Lys Met Asp Leu Phe Tyr Arg Asn Ser Ala Asp Leu Glu
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Gln Gln Asp Ile Glu Gly Pro Trp Thr Glu Pro Tyr Thr Asp Gly
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Leu Cys Pro Val Pro Arg Leu Ile Ile Thr Pro Glu Thr Pro Glu
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Phe Gly Lys Thr Val Ala Leu Pro Asp His Gln Thr Leu Ser His
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